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## DRAFT TECHNICAL REPORT

# Lower Duwamish Triad Sampling Event

Prepared for  
US EPA Region 10

Prepared by

Herrera Environmental Consultants, Inc.  
2200 Sixth Avenue, Suite 1100  
Seattle, Washington 98121  
Telephone: 206/441-9080

And

U.S. Army Corps of Engineers  
Seattle District  
4735 East Marginal Way South  
Seattle, Washington 98134

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## **Introduction**

This revised and expanded document describes field-related events and results from a 2004 investigation to define patterns and determine source(s) of polychlorinated biphenyls (PCBs) in a reach of the Lower Duwamish River, Seattle, Washington. These results are being used to guide Remedial Investigation/Feasibility Study (RI/FS) and Corrective Measures Study (CMS) decisions. The investigation is associated with two regulatory authorities, RCRA and CERCLA. A variety of parameters were analyzed, including grain-size, total organic carbon (TOC), semivolatile organic compounds (SVOC), metals, and PCB, in order to determine exceedances of Washington State Sediment Management Standards. The distribution of PCB up-river from a known source was not understood in relation to other potential sources. The PCB data are being used to resolve PCB patterns and to permit EPA to make a "boundary" decision concerning the influence of two different source areas. Immunoassay analyses of sediments from shallow borings were also used to direct placement of deeper cores. Samples were split into Aroclor samples (for off-site laboratory analysis) and immunoassay samples (for rapid-turnaround analysis at a Navy laboratory in San Diego, CA), resulting in a large collaborative data set (97 stations) that permitted a "Demonstration of Method Applicability" (DMA) for the immunoassay in a complex urban sediment mixture. That evaluation is reported elsewhere. (Section PCB results section for a link.)

## **Summary of Field Sampling Activities**

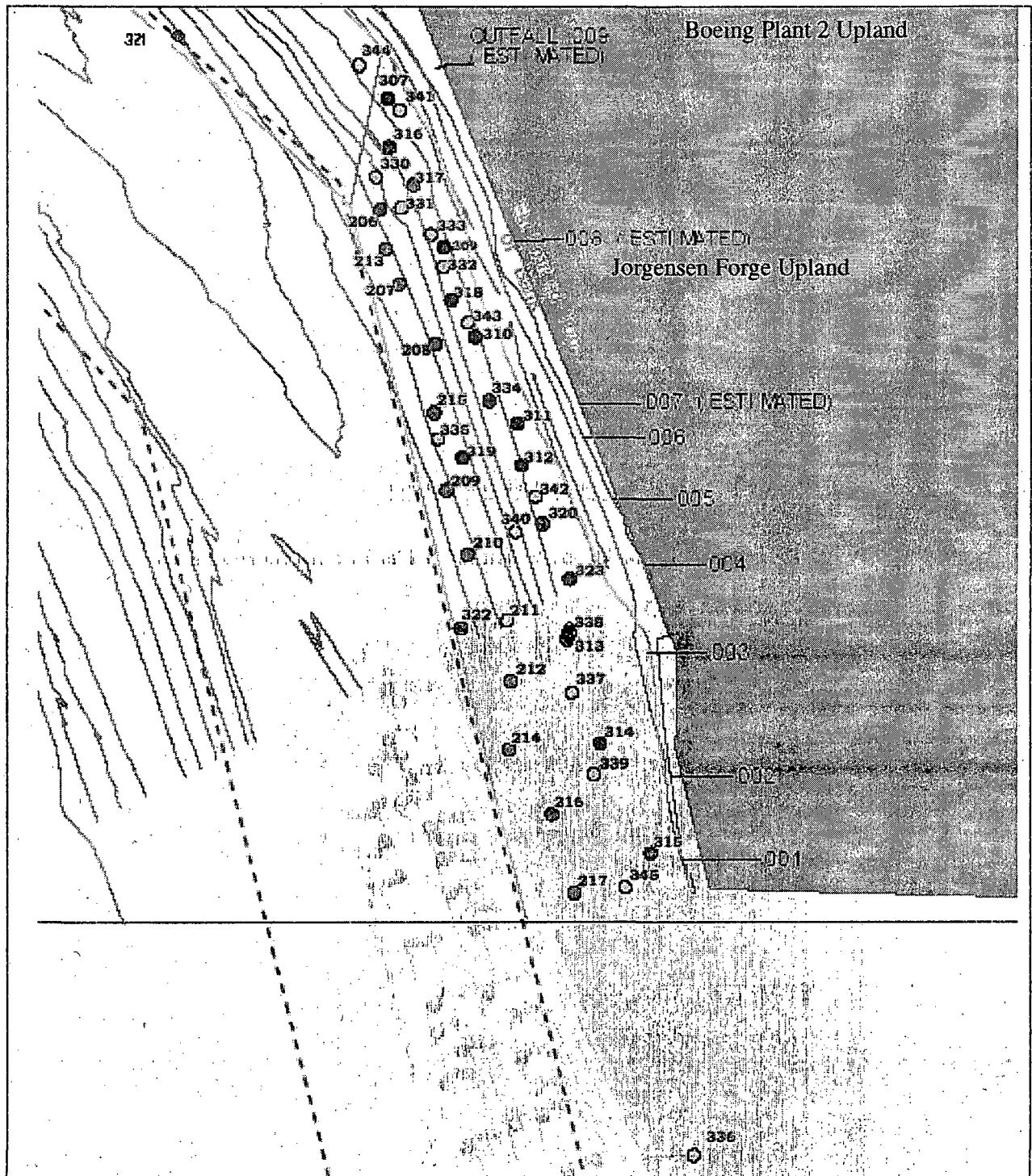
Figure 1 shows a map of the reach near river mile 3.8 on the Lower Duwamish River, in Seattle, Washington. In the figure, the stations designated 200 and purple coded are stations previously sampled by the Boeing Company; stations coded 300 were sampled in this investigation as surface and subsurface cores. Yellow stations were surface sampled using an adaptive fill methodology to address areas of greatest uncertainty following evaluation using Visual Sample Plan version 3.0.

The sampling activities resulted in 97 discrete surface and subsurface samples. All 97 samples were collected using a single methodology. The digital data for all 97 samples were collected using a handheld personal digital assistant (PDA). The PDA data were imported into a Microsoft® Access® relational database. Access was used to create histograms, sample statistics, stratifications, and to generate longitudinal and cross-sectional maps of the area. The PDA was programmed with a specific methodology for the site, which included surface sampling (rapid turnaround) and sampling in San Diego, CA, resulting in a large collaborative data set that permitted a "Demonstration of Method Applicability" (DMA) for the immunoassay in a complex urban sediment mixture. This evaluation is reported elsewhere. (Section PCB results section for a link.)

### **4.3 Detailed Sampling Activities**

Sampling of the reach near river mile 3.8 on the Lower Duwamish River, all sampling sites, station designations, and purple coded stations (300) are detailed below. The USACE/Herrera Environmental Consultants (Herrera) collected all the stations as surface samples using an adaptive fill methodology, except for areas of greatest uncertainty following evaluation using Visual Sample

Figure 1. Sample Locations on the Lower Duwamish River, Near River Mile 3.8



Field sampling activities were performed by Herrera Environmental Consultants (Herrera) and staff from the Army Corps of Engineers (USACE) between August 16 and 27, 2004. Phase 1 (a comparison of a limited number of PCB samples to both immunoassay by Method 4020 and Method 8082-GC/ECD) was completed at an earlier date. Sediment sampling for this investigation was completed in two phases, as directed by the project specific Quality Assurance Project Plan (QAPP, USACE 2004):

- Phase 2 – Surface and subsurface sediment samples were collected at 16 station locations. These locations were identified as EPA bank/outfall locations, EPA “hot-spot” locations, and EPA spatial locations.
- Phase 3 – Adaptive (“Triad”) surface sediment samples were collected at additional locations. These locations were identified as previous Boeing sample locations and “filler” sample locations between Phase 2 locations.

One of the purposes of the data collection activity was to increase the amount of data to better determine the correspondence between the two analyses, in order to determine the usefulness of the immunoassay in “guiding” future sampling for this suite of compounds.

Planned sampling activities are described in the QAPP; actual field activities are described below, and documented in log books and on sediment log sheets (Appendix A). Photographic documentation representing a subset of sediment samples is presented in Appendix B.

The site specific Health and Safety Plan (HASP) (Herrera 2004) was followed and no health and safety concerns occurred during the field sampling event.

Surface and subsurface sediment samples were collected between the river bank and navigation channel using the vessel R/V Kittiwake to deploy sampling equipment. Surface sediment samples were collected using a single van Veen sampler and subsurface sediment samples were collected using a Vibracorer. Sampling equipment was deployed and retrieved from the stern of the boat using a winch.

Phase 2 surface sediment samples were collected during the first two days of field activity. Subsurface core samples were collected over two days following surface sample collection. An attempt was made to collect surface and subsurface samples at the same location. Due to subsurface debris, most core locations were shifted from the corresponding surface sample locations.

Sample locations were electronically recorded using an onboard differential global positioning system (DGPS) with an accuracy of  $\pm 2$  meters. A comparison of actual (final) field station locations to planned station locations is presented in Table 1. Station locations were adjusted in the field if refusal was encountered at the planned station location coordinates. Thirteen surface station locations varied from planned station locations by greater than 1 meter, as shown in Table 1. Refusals occurred due to the presence of rocks and other debris or due to insufficient water depth for the boat to maneuver without disturbing the sediment surface. Final field locations for

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subsurface core samples varied by greater than one meter from the corresponding final surface grab station locations due to refusal by subsurface debris 15 out of 16 times.

Planned and final station location coordinates are presented in Appendix C.

**Table 1. Comparison of planned station location to final field station locations.**

Station ID	Distance of Final Surface Grab Station Location from Planned Station Location	Distance of Final Subsurface Core Station Location from Final Surface Grab Station Location	Comments
<b>Phase 2 stations</b>			
SD-DUW-307	0.6 meters south	5.7 meters southwest	2 subsurface refusals
SD-DUW-309	0.3 meters north	5.9 meters southwest	
SD-DUW-310	0.3 meters southwest	4.1 meters southwest	1 surface refusal
SD-DUW-311	0.9 meters southeast	3.2 meters northwest	
SD-DUW-312	5.0 meters southwest	2.0 meters east	3 surface refusals
SD-DUW-313	8.2 meters northwest	4.3 meters southwest	5 surface and 2 subsurface refusals
SD-DUW-314	7.3 meters northwest	0.8 meters northwest	5 surface and 1 subsurface refusals
SD-DUW-315	0.3 meters northwest	1.4 meters south	1 surface refusal
SD-DUW-316	0.5 meters south	5.5 meters southwest	
SD-DUW-317	0.4 meters south	5.5 meters northwest	1 subsurface refusal
SD-DUW-318	0.3 meters northwest	2.8 meters northeast	
SD-DUW-319	1.7 meters southwest	1.3 meters northwest	
SD-DUW-320	0.9 meters northeast	1.6 meters southwest	
SD-DUW-321	0.8 meters southeast	2.3 meters northeast	
SD-DUW-322	0.3 meters southwest	2.1 meters southwest	1 subsurface refusal
SD-DUW-323	2.2 meters southwest	2.3 meters southwest	1 surface refusal
<b>Phase 3 stations</b>			
SD-DUW-206	0.9 meters east	NA	
SD-DUW-207	0.3 meters south	NA	
SD-DUW-208	0.6 meters northeast	NA	
SD-DUW-209	3.4 meters southeast	NA	1 refusal
SD-DUW-210	0.2 meters south	NA	
SD-DUW-211	1.1 meters southwest	NA	
SD-DUW-212	0.6 meters east	NA	
SD-DUW-213	0.8 meters northeast	NA	
SD-DUW-214	0.5 meters southeast	NA	1 surface and 1 subsurface refusal
SD-DUW-215	1.0 meters southeast	NA	1 surface and 1 subsurface refusal
SD-DUW-216	0.6 meters northeast	NA	1 surface refusal
SD-DUW-217	0.4 meters northwest	NA	
SD-DUW-330	0.7 meters northeast	NA	1 surface refusal
sd_081020_main text.doc	14 October 2008	14 October 2008	
October 2008	14 October 2008	14 October 2008	USACE/Herrera Environmental Consultants

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Station ID	Distance of Final Surface Grab Station Location from Planned Station Location	Distance of Final Subsurface Core Station Location from Final Surface Grab Station Location	Comments
SD-DUW-331	0.8 meters southwest	NA	
SD-DUW-332	1.6 meters southeast	NA	
SD-DUW-333	2.9 meters southwest	NA	
SD-DUW-334	1.0 meters west	NA	
SD-DUW-335	0.8 meters northeast	NA	
SD-DUW-336	0.4 meters southeast	NA	
SD-DUW-337	0.0 meters	NA	
SD-DUW-338	14.5 meters northwest	NA	5 refusals
SD-DUW-339	12.4 meters northwest	NA	5 refusals
SD-DUW-340	0.6 meters southeast	NA	
SD-DUW-341	5.0 meters northwest	NA	2 refusals
SD-DUW-342	0.1 meters	NA	
SD-DUW-343	0.4 meters south	NA	
SD-DUW-344	6.4 meters northwest	NA	3 refusals
SD-DUW-345	0.1 meters west	NA	

\* Coordinates are North American Datum (NAD) of 1983 State Plane Washington North.  
NA Not applicable. (Phase 3 involved surface sampling only.)

All sample handling and custody procedures outlined in the QAPP were followed. Field activities and deviations from the QAPP for surface and subsurface sampling are discussed in more detail below.

#### **Surface Sediment Sampling**

Surface sediment samples for Phase 2 sampling were collected on August 16 and 17, 2004. Phase 3 surface sediment samples were collected on August 26 and 27, 2004. All samples were collected from the top 10 centimeters of sediment using a single van Veen sampler as described in the QAPP.

No deviations from the QAPP, other than station location adjustments due to rocks or debris at the sediment surface discussed above, were performed during field sampling activities.

#### **Subsurface Sampling**

Subsurface sediment samples were collected using a Vibrocorer on August 18 and 19, 2004. Samples were collected from all 16 stations identified for the Phase 2 sampling event.

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Deviations from the QAPP for subsurface sediment field sampling activities included the following:

- Overlying water was decanted from the top opening of the core tubes rather than drilling holes through the core tube at the sediment/water interface. Because sediment was not collected for analysis from the 0 to 1 foot interval, any disturbance to the sediment surface caused by decanting from the top of the core tube did not affect sample quality.
- Cores were processed at Herrera's laboratory, where a walk-in cooler was not available. Core tubes were stored vertically, packed in ice, and wrapped in plastic until processed. Samples were not adversely affected.
- Adequate sediment core depth was not achieved at two sample stations due to refusal. Samples were not collected from the 2- to 3-foot depth at SD-DUW-318 and samples were not submitted for the 4- to 5- or 5- to 6-foot depths at SD-DUW-321.
- An additional (unplanned) subsurface sample was collected from station SD-DUW-313 because a sheen was observed in this core. The sample collected from the 1- to 2-foot sample interval was submitted to the EPA Region 10 Laboratory for analysis of semivolatile organic compounds, and results are presented later in this report. Deeper samples at this location could not be collected due to refusal and the sampling station was moved four meters to the west.

Accurate depth of penetration could not be measured, partly due to tubes bending as they were driven into the sediment and because physical indication of penetration depth was not always evident on the core tubes once they were retrieved. On the second day of coring activities, the 10-foot core tubes were cut down to 7-foot lengths prior to sampling. This prevented the tubes from bending, and the entire length of the tube could then penetrate the sediment.

The estimated and measured percent recovery for each station is presented on the sediment log forms (Appendix A). Sample collection depths (i.e., 1 to 2 feet, 2 to 3 feet, etc.) were not adjusted based on the calculated percent recovery. An "estimated" qualifier (E) was applied to percent recovery values associated with cores that did not fully penetrate the sediment and only physical indications (markings on the core tube) or Vibracorer cable payout estimates were used to calculate recoveries.

#### Investigation Derived Waste

All excess sediment was collected in 42.5-gallon buckets. Two representative samples were collected from the 5-gallon buckets containing sediment from stations identified in Table 2 and analyzed by the toxicity characteristic leaching procedure (TCLP) for lead to determine disposal requirements. Sample concentrations were below the Resource Conservation and Recovery Act

(RCRA) limit of 5 milligrams per liter (mg/L), indicating that the sediment does not need to be disposed of as hazardous waste.

**Table 2. Investigation derived waste (IDW) lead results by toxicity characteristic leaching procedure (TCLP) for the Lower Duwamish Triad project.**

Sample ID	Result (mg/L)	IDW bucket identification
WD01	0.4	SD-312 and SD-316
WD02	0.1 U	SD-210 and SD-211

mg/L milligrams per liter

U The material was analyzed for, but was not detected. The associated numerical value is the reporting limit.

## Analytical Results

A total of 70 Phase 2 sediment samples, including five field duplicate samples and one rinsate sample, were submitted for laboratory analysis; a total of 31 Phase 3 sediment samples, including three field duplicate samples, were submitted for laboratory analysis (Appendix D, Tables D-1 and D-2).

Analytical Resources, Incorporated (ARI) analyzed sediment samples for total organic carbon (TOC), grain size distribution, semivolatile organic compounds (SVOCs), and mercury. ARI analytical results are presented in Appendix E; a data quality review is provided separately. Analytical results and associated data quality issues are discussed below.

The Environmental Division of the US Navy's SPAWARSCEN Laboratory analyzed 100 samples for PCBs by Method 4020 (immunoassay). Results are provided in Appendix F and Table 8.

The EPA Region 10 Manchester Laboratory analyzed 32 sediment samples plus two duplicate samples for the metals arsenic, cadmium, chromium, copper, lead, nickel, silver, and zinc.

### Total Organic Carbon

ARI analyzed 99 sediment samples, including seven field duplicates for TOC using EPA method 9060 modified by PSEP for sediments (PSEP 1996). Analytical results are presented in Tables E-1 (Phase 2 samples) and E-2 (Phase 3 samples); no data quality issues were associated with the TOC analyses.

Surface sediment samples collected in both Phase 2 and Phase 3 resulted in TOC concentrations ranging from 1.45 to 2.78 percent. The highest TOC value occurred at station SD-DUW-214, located near the navigation channel; the lowest TOC value occurred at station SD-DUW-342, located along the river bank. In general, TOC concentrations for subsurface sediment samples were lower at stations located along the bank and higher concentrations were found at sample stations located closer to the navigation channel.

Seven field duplicate samples were analyzed for TOC (Table 3). All relative percent difference (RPD) values (ranging from 0.9 to 30.0 percent) met the control limit (less than 50 percent) established in the QAPP.

**Table 3. Comparison of sample and field duplicate total organic carbon (TOC) results for the Lower Duwamish Triad project.**

Sample ID	Field Duplicate ID	Sample Result (percent)	Field Duplicate Result (percent)	RPD
SD-314-0000	SD-325-0000	1.66	1.73	4.1
SD-320-0003	SD-328-0003	1.83	1.72	6.2
SD-321-0000	SD-324-0000	2.25	2.23	0.9
SD-323-0001	SD-326-0001	1.65	1.58	4.3
SD-207-0000	SD-433	2.11	1.56	30.0
SD-333-0000	SD-431	2.50	2.24	11.0
SD-338-0000	SD-432	1.81	1.92	5.9

RPD Relative percent difference

### Mercury

ARI analyzed 19 sediment samples for mercury using EPA method 7471A (U.S. EPA 1986). Samples from 1-2 feet were eliminated during the planned number of samples due to resource decisions. Analytical results are presented in Table E-4 and Table 5, below. All samples were analyzed 11 to 22 days outside of the recommended holding time. Holding time exceedances for each sample are noted in Table E-4. Therefore, samples that had a detected concentration of mercury above the laboratory reporting limit were qualified as estimated (flagged J), to indicate that sample results may be biased low. Mercury was not detected in the sediment sample from station SD-DUW-341 at the laboratory reporting limit; the result was rejected (flagged R), in accordance with EPA data review guidelines (U.S. EPA 2004). Because the reported concentration is expected to be lower than the actual concentration (associated with the extended holding time prior to analysis), the presence or absence of the analyte cannot be verified.

Mercury concentrations in surface samples ranged from 0.09 to 0.39 mg/kg. The SQS criterion for mercury is 0.41 mg/kg and the CSL criterion for mercury is 0.59 mg/kg. No project samples exceeded SQS or CSL criteria.

Two field duplicates were analyzed for mercury (Table 4). The RPD values met the established control limit (less than 50 percent) established in the QAPP.

**Table 4. Comparison of inorganic compounds laboratory duplicate results for the Lower Duwamish Triad project. (Note: there were no field duplicates analyzed.)**

Compound	Sample ID	Field Duplicate ID:	Sample Result (mg/kg)	Field Duplicate		Relative Percent Difference	
				SD-	Q		
Ag	309-0001	309-0001D	1	U	0.99	U	0%
Ag	320-0001	320-0001D	1.3		1.3		0%
As	309-0001	309-0001D	11		12		4%
As	320-0001	320-0001D	20		21		2%
Cd	309-0001	309-0001D	1.2		1.4		8%
Cd	320-0001	320-0001D	2.02		1.74		7%
Cr	309-0001	309-0001D	35.2		34.7		1%
Cr	320-0001	320-0001D	86.1	J	83.2	J	2%
Cu	309-0001	309-0001D	55.1		57.1		2%
Cu	320-0001	320-0001D	86.1		88.8		2%
Hg	321-0000	324-0000	0.18	J	0.2	J	11%
Hg	314-0000	325-0000	0.23	J	0.2	J	14%
Ni	309-0001	309-0001D	29.5		28.7		1%
Ni	320-0001	320-0001D	35.5		34.1		2%
Pb	309-0001	309-0001D	44.8		45.9		1%
Pb	320-0001	320-0001D	433		421		1%
Zn	309-0001	309-0001D	127		125		1%
Zn	320-0001	320-0001D	351		328		3%

mg/kg milligrams per kilogram

RPD Relative percent difference

J The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.

**Other Inorganics**

EPA Region 10's Manchester Laboratory results are provided in Appendix G and summarized in Table 5.

Table 5. Results for Inorganic Compounds

	Sediment Quality Standards * (SQS)	Cleanup Screening Levels Criteria * (CSL)	307- 0000	307- 0001	309- 0000	309- 0001	Duplic- cate	310- 0000	310- 0001	311- 0000	311- 0001	312- 0000	312- 0001	313- 0000
As	57	93	15	5.2	14	11	12	12	18	13	15	24.5	26.7	37
Cd	5.1	6.7	1.76	0.5 U	1.3	1.2	1.4	1.1	1.65	1	1.87	1.56	2.29	2.5
Cr	260	270	43.2	11.7	81.7	35.2	34.7	59.9	34.3	69.7	69.2	78.7	64	584
Cu	390	390	45.9	12.1	58	55.1	57.1	52.5	43.4	60.3	79.5	71	71.1	128
Pb	450	530	209	2.8	108	44.8	45.9	287	26.7	169	500	196	574	632
Ni	LAET = 140		50	9.07	48.3	29.5	28.7	37.9	22.8	58	51.1	53.1	28.4	129
Ag	6.1	6.1	1 U	1 U	0.99 U	1 U	0.99 U	0.99 U	1 U	0.99 U	1 U	1 U	0.99 U	5 U
Zn	410	960	1300	298	157	127	125	142	157	160	216	174	457	529
Hg	0.41	0.59	0.09 J	J	0.24 J			0.14 J		0.11 J		0.20 J		0.39 J
As	57	93	13	24	4.5U	12	4.5 U	14	12	15	13	5.9	13	
Cd	5.1	6.7	0.9	0.75	0.5 U	0.54	0.5 U	0.8	1.2	1.1	1.5	0.57	0.5 U	0.71
Cr	260	270	23.3	108	11.7	77.7	9.96	39.7	35.4	44.2	35.9	54.1	11.4 J	42.5
Cu	390	390	38.4	72	9.53	68.8	7.32	52.3	56.1	59.6	67.4	56.4	10.7	48.9
Pb	450	530	13.9	81.8	2.5U	67.6	2.5 U	59.00	57.5	69.1	81.7	82.6	2.5U	204
Ni	LAET = 140		19	47.6	8.39	57.5	6.92	28.7	27	27.6	26.1	35.9	7.79	25.5
Ag	6.1	6.1	1 U	0.99 U	1 U	0.99 U	0.99 U	1 U	0.99 U	1 U	1 U	0.99 U	1 U	0.99 U
Zn	410	960	67.1	217	23.9	133	20.2	211	150	175	172	153	21	162
Hg	0.41	0.59	0.16 J	0.23 J		0.09 J		0.15 J		0.10 J		0.10 J		0.16 J
As	57	93	21	20	20	21	12	13	14	12	32	22.8		
Cd	5.1	6.7	1.1	0.78	2.02	1.74	0.58	1.1	0.5 U	0.78	2.5U	0.51		
Cr	260	270	30.5	87.1 J	86.1 J	83.2 J	32.1	23.3	30.1	27.9	1070	32		
Cu	390	390	39.1	72.7	86.1	88.8	53.1	29.2	53.1	46.2	123	53.7		
Pb	450	530	23.6	211	433	421	28	14	25.5	37.7	2350	171		
Ni	LAET = 140		20.3	48.9	35.5	34.1	23.2	16.6	22	19.8	144	31		
Ag	6.1	6.1	0.99 U	1.1	1.3	1.3	0.99 U	0.99 U	0.99 U	0.99 U	5U	0.99 U		
Zn	410	960	133	237	351	328	112	120	115	110	1590	189		
Hg	0.41	0.59		0.21 J			0.18 J		0.14 J		0.20 J			

\* Washington State Sediment Management Standards, Sediment Quality Standards (SQS) and Cleanup Screening Level (CSL), WAC Chapter 173-204. LAET = Lowest Apparent Effects Threshold. (LAET is not a regulatory standard.)

*Italic and Underline:* exceeds SQS

**Bold:** exceeds CSL or LAET

## Semivolatile Organic Compounds

ARI analyzed 19 sediment samples, including two field duplicates, for SVOCs using EPA method 8270B (U.S. EPA 1986). SVOC analytical results are presented in Table E-3; no data quality issues were associated with the SVOC analyses, with the exceptions noted below.

Two field duplicate samples were analyzed for SVOCs (Table 6). The relative percent difference (RPD) was calculated for all detected compounds identified greater than five times the reporting limit (RL). The difference was calculated if the detected compound concentration was less than five times the RL in either the sample or field duplicate. A control limit of less than 50 percent RPD was established in the QAPP; a control limit of two times the RL was used to evaluate difference values. Several compounds exceeded the RPD or difference control limits for sample SD-314-0000 and field duplicate SD-325-0000. Because all RPD or difference values met the control limits for sample SD-321-0000 and field duplicate SD-324-0000, only samples SD-314-0000 and SD-325-0000 were qualified as estimated (flagged J) for compounds that exceeded the control limits, as shown in Table 6. It is probable that concentration differences were a result of matrix variability.

Surface sediment samples from all Phase 2 station locations and one subsurface sediment sample collected from 1 to 2 feet deep were analyzed for SVOCs. A comparison of SVOC concentrations found in project samples to Washington State Sediment Quality Standards (SQS) and Cleanup Screening Level (CSL) criteria (WAC 173-204) is presented in Table 7.

SQS criteria were exceeded in surface sediments for one or more SVOCs at eight of the 16 station locations. Stations with the most SQS exceedances were generally bank/outfall samples. The station with the most SQS exceedances was bank/outfall location SD-DUW-307. In general, this station also had the highest concentration of exceeded compounds.

The second stage of the analysis of each group of participants involved the calculation of the mean digit ratio (D) for each participant. The digit ratio was calculated as the ratio of the number of digits to the number of non-digits (i.e., the ratio of the number of fingers to the number of toes). The mean digit ratio for each participant was then calculated by averaging the digit ratios for all 10 fingers and 10 toes. The mean digit ratio for each participant was then used to calculate the D for each participant. The D for each participant was then used to calculate the D for each participant.

Example 11 shows that it is possible to implement the watermarking scheme using a public key cryptosystem based on elliptic curve scalar multiplication. The watermarking scheme can be extended to incorporate the elliptic curve standard [3] and [4] and the digital signature scheme [5].

the presence of a large amount of water-soluble organic material in the sample. The presence of such materials may have influenced the low apparent viscosity. It is also possible that the dependence of shear modulus on frequency (Fig. IV-30) might be due to the higher concentration of dissolved compounds.

**Table 6 Comparison of sample and field duplicate semivolatile organic compound (SVOC) results for the Lower Duwamish Triad project.**

Compound (results in µg/kg)	RL (µg/kg)	Sample SD-314-0000	Field Duplicate SD-325-0000	RPD or diff <sup>a</sup>	Sample SD-321-0000	Field Duplicate SD-324-0000	RPD or diff <sup>a</sup>
Phenol	20	160 J	560 J	111.1	130	140	7.4
4-Methylphenol	20	21	31	10.0	ND	ND	
Naphthalene	20	28 J	160 J	132.0	ND	ND	
2-Methylnaphthalene	20	27 J	100 J	73.0	ND	ND	
Acenaphthylene	20	20 U	22	2.0	ND	ND	
Acenaphthene	20	42 J	190 J	148.0	ND	ND	
Dibenzofuran	20	25	54	29.0	ND	ND	
Diethylphthalate	20	28	20 U	8.0	20 U	27	7.0
Fluorene	20	51 J	220 J	169.0	ND	ND	
Phenanthrene	20	400 J	1,400 J	111.1	81	68	13.0
Carbazole	20	69	67	2.0	ND	ND	
Anthracene	20	130 J	330 J	87.0	30	25	5.0
Di-n-Butylphthalate	20	30	42	12.0	ND	ND	
Fluoranthene	20	600 J	1,100 J	58.8	270	230	16.0
Pyrene	20	570 J	1,500 J	89.9	240	210	13.3
Butylbenzylphthalate	20	20 U	26	6.0	34	27	7.0
Benzo (a) anthracene	20	220 J	500 J	77.8	98	81	17.0
Bis (2-ethylhexyl) phthalate	20	150	140	6.9	240	200	18.2
Chrysene	20	290 J	550 J	61.9	140	110	24.0
Benzo (b) fluoranthene	20	260	430	49.3	180	150	18.2
Benzo (k) fluoranthene	20	210 J	420 J	66.7	140	130	7.4
Benzo (a) pyrene	20	230 J	510 J	75.7	110	97	13.0
Indeno (1,2,3-cd) pyrene	20	110 J	170 J	42.9	52	46	6.0
Dibenz (a,h) anthracene	20	32	72	40.0	ND	ND	
Benzo (g,h,i) perylene	20	88 J	150 J	62.0	44	39	5.0

µg/kg micrograms per kilogram

RL Laboratory reporting limit

RPD Relative percent difference

diff Difference between the sample result and field duplicate result

ND Not detected above the laboratory reporting limit; the RPD value or difference was not calculated.

<sup>a</sup> The relative percent difference was calculated if both the sample and field duplicate result were greater than five times the laboratory reporting limit (RL). If the sample or the field duplicate result was less than five times the RL, the difference between the two values was calculated. The control limit for RPD is 50 percent; the control limit for difference is two times the RL.

U The material was analyzed for, but was not detected. The associated numerical value is the reporting limit.

J The associated numerical value is considered an estimate.

Bold values indicate that the RPD or difference control limit was exceeded.

**Table 7. Comparison of Sediment Quality Standards and Cleanup Screening Levels to Semivolatile Organic Compounds.**  
Bold and shaded is > CSL, bold and unshaded is SQS<x<CSL, and values is < SQS and neither bolded nor shaded

Compound	Cleanup Screening Levels Criteria <sup>a</sup> (CSL)	Sediment Quality Standards <sup>a</sup> (SQS)	Results																
			SD-307-0000	SD-309-0000	SD-310-0000	SD-311-0000	SD-312-0000	SD-313-0000	SD-313-0001	SD-314-0000	SD-315-0000	SD-316-0000	SD-317-0000	SD-318-0000	SD-319-0000	SD-320-0000	SD-321-0000	SD-322-0000	SD-323-0000
<b>Low molecular weight PAH</b>			mg/kg carbon <sup>b</sup>	mg/kg carbon <sup>b</sup>										mg/kg carbon <sup>b</sup>					
LPAH	780	370	490	250	74	32	230	49	2.2	39 J	59	13	16	62	5.1	24	4.9	5.9	24
Naphthalene	170	99	0.91 U	22	1.6	3.4	6.6	9.9	1.1 U	1.7 J	1.3 U	0.84 U	0.78 U	2.2	1.1 U	1.5	0.89 U	0.99 U	0.88
Acenaphthylene	66	66	2.6	4.4	2.1	1.9	3.8	0.90 U	1.1 U	1.2 U	1.5	0.84 U	0.78 U	1.4	1.1 U	1.2 U	0.89 U	0.99 U	0.80 U
Acenaphthene	57	16	1.2	10	2.8	3.8	9.7	9.0	1.1 U	2.5 J	2.8	0.84 U	0.78	2.5	1.1 U	1.2	0.89 U	0.99 U	1.4
Fluorene	79	23	68	32	8.6	11	36	11	1.1 U	3.1 J	3.2	0.98	1.5	7.3	1.1 U	3.0	0.89 U	0.99 U	1.7
Phenanthrene	480	100	220	150	48	60	150	63	2.2	24 J	43	9.8	11	41	5.1	15	3.6	4.5	17
Anthracene	1,200	220	200	28	11	12	26	16	1.1 U	7.8 J	8.3	2.0	2.5	7.8	1.1 U	3.0	1.3	1.4	3.3
2-Methylnaphthalene	64	38	1.2	16	1.7	3.3	1.9	8.6	1.1 U	1.6 J	1.3 U	0.84 U	0.78 U	1.8	1.1 U	1.5	0.89 U	0.99 U	0.88
<b>High molecular weight PAH</b>			mg/kg carbon <sup>b</sup>	mg/kg carbon <sup>b</sup>										mg/kg carbon <sup>b</sup>					
HPAH	5,300	960	1,600	530	330	450	450	200	8.2	160 J	600	110	120	270	44	90	57	62	170
Fluoranthene	1,200	160	455	149	76	109	132	45	4.3	36 J	146	24	29	68	8.6	23	12	15	35
Pyrene	1,400	1,000	177	100	51	76	85	50	2.7	34 J	115	20	22	46	11	18	11	8.9	33
Benz(a)anthracene	270	110	214	50	29	38	43	20	1.1 U	13 J	42	8.9	9.3	24	3.4	7.4	4.4	4.9	14
Chrysene	460	110	300	54	37	46	47	22	1.2	17 J	61	12	16	31	4.3	9.9	6.2	7.4	18
Total benzofluoranthenes	450	230	320	92	73	100	75	33	1.1 U	28 J	130	27	28	53	9.9	17	14	14	41
Benzo(a)pyrene	210	99	91	45	31	42	38	19	1.1 U	14 J	50	10	10	25	4.0	8.0	4.9	5.4	16
Indeno(1,2,3-cd)pyrene	88	34	20	19	14	19	13	5.9	1.1 U	6.6 J	25	5.3	4.7	13	1.7	3.6	2.3	3.0	8.0
Dibenzo(a,h)anthracene	33	12	10	7.7	5.6	7.1	3.9	2.8	1.1 U	1.9 J	9.6	2.0	1.1	4.9	1.1 U	1.2 U	0.89 U	1.0	3.1
Benzo(g,h,i)perylene	78	31	12	13	11	14	9.7	5.4	1.1 U	5.3 J	19	4.3	3.5	9.3	1.4	2.6	2.0	2.2	6.0
<b>Phthalates</b>			mg/kg carbon <sup>b</sup>	mg/kg carbon <sup>b</sup>										mg/kg carbon <sup>b</sup>					
Dimethylphthalate	53	53	0.86 J	1.1	1.5	1.4	0.85	0.90 U	1.1 U	1.2 U	2.2	1.8	1.1	1.1 U	1.2 U	0.89 U	0.99 U	3.0	
Diethylphthalate	110	61	0.91 U	0.90 U	1.0	1.1 U	3.5	0.90 U	1.1 U	1.7	1.3 U	0.84 U	4.7	4.6	1.1 U	3.0	0.89 U	7.4	0.80 U
Di-n-Butylphthalate	1,700	220	3.0	0.90 U	0.96 J	1.1 U	0.89	1.5	1.1 U	1.8	1.3 U	0.93	1.8	1.0	1.6	2.3	0.89 U	0.99 U	1.2
Butylbenzylphthalate	64	4.9	1.5	1.5	2.1	2.2	1.4	1.9	1.1 U	8.9	2.8	3.9	2.0	1.1 U	2.3	1.5	1.5	4.0	
Bis(2-ethylhexyl)phthalate	78	47	64	10	14	21	8.5	4.5	1.1 U	9.0	39	13	14	14	5.4	6.8	11	11	16
<b>Other semivolatile organic compounds</b>			mg/kg carbon <sup>b</sup>	mg/kg carbon <sup>b</sup>										mg/kg carbon <sup>b</sup>					
Dibenzofuran	58	15	8.2	19	4.1	5.4	18	2.5	1.1 U	1.5	2.0	0.84 U	0.78	3.7	1.1 U	1.7	0.89 U	0.99 U	0.84
<b>Ionizable organic compounds</b>			µg/kg dry weight <sup>c</sup>	µg/kg dry weight <sup>c</sup>										µg/kg dry weight <sup>c</sup>					
Phenol	1,200	420	440	290	270	1,200	610	620	20 U	160	28	310	1,100	800	32	410	130	290	840
4-Methylphenol	670	52	19 J	20 U	27	22	22	20 U	21	20 U	19 U	34	28	20 U	20 U	20 U	20 U	83	

<sup>a</sup> Washington State Sediment Management Standards, Sediment Quality Standards (SQS) and Cleanup Screening Level (CSL), WAC Chapter 173-204.

<sup>b</sup> Units in mg/kg carbon represent concentrations in parts per million, normalized to organic carbon. To normalize to TOC, the dry weight concentration for each parameter is divided by the decimal fraction representing the percent TOC content of the sediment.

<sup>c</sup> µg/mg micrograms per kilogram.

*Italic and underlining* indicates the sample result is greater than the SQS value.

U The material was analyzed for, but was not detected. The associated numerical value is the reporting limit.

J The associated numerical value is considered an estimated concentration.

LPAH represents the sum of the following "low molecular weight polynuclear aromatic hydrocarbon" compounds: naphthalene, acenaphthylene, acenaphthene, fluorene, phenanthrene, and anthracene.

HPAH represents the sum of the following "high molecular weight polynuclear aromatic hydrocarbon" compounds: fluoranthene, pyrene, benzo(a)anthracene, chrysene, total benzofluoranthenes, benzo(a)pyrene, indeno(1,2,3-cd)pyrene, dibenzo(a,h)anthracene, and benzo(g,h,i)perylene.

Total benzofluoranthenes represents the sum of the concentrations of the "b" and "k" isomers.

## Grain Size Distribution

ARI analyzed 26 sediment samples for grain size distribution using the PSEP method (PSEP 1996). Analytical results are presented in Table E-5. USACE selected five additional sediment samples for grain size distribution analysis after the field sampling event was completed. These five samples were frozen prior to analysis and have been qualified as estimated (flagged J) since freezing may impact particle size distribution. No other data quality issues were associated with the grain size distribution analyses.

In general, station locations along the river bank had a greater percentage of sand and gravel in both surface and subsurface samples; station locations SD-DUW-321 and SD-DUW-322, which are located nearest to the navigation channel, contained a higher percentage of fine-grained material. The stations with a greater percentage of fine-grained material generally contained higher TOC concentrations and stations with a greater percentage of gravels and sands had lower TOC concentrations.

Two field duplicates were analyzed for grain size distribution (Table 7). The relative percent difference (RPD) was only calculated for the percent fines content (less than 62.5 microns) of the samples. The RPD values (0.8 and 5.4 percent) met the established control limit (less than 50 percent) established in the QAPP.

**Table 8. Comparison of sample and field duplicate percent fines results for the Lower Duwamish Triad project.**

Sample ID	Field Duplicate ID	Sample Result (percent fines <sup>a</sup> )	Field Duplicate Result (percent fines <sup>a</sup> )	RPD
SD-321-0000	SD-324-0000	77.7	78.3	0.8
SD-333-0000	SD-431	59.3	56.2	5.4

RPD = Percent Relative Difference:  $((A-B)/((A+B)) * 100\%)$

<sup>a</sup> The percent fines content (less than 62.5 microns) was used to compare sample and field duplicate results.

## PCBs

The Navy SPAWARSYSCEN Laboratory analyzed the samples using the EPA Method 4020 immunoassay. EPA sent Method 8082 (Aroclor) analyses to 3 EPA laboratories in Regions 6, 8, and 10. Table 9 shows the dry weight QC data for Method 4020. These results were judged acceptable in accordance with the QAPP. Table 10 shows results of field duplicate analyses using Method 8082, and indicates some heterogeneity in repeated sampling from the same grab. This shows that organic-carbon-normalized data. Tables 11 and 12 shows dry-weight results according to both methods. The organic-carbon normalized results are highlighted relative to the Washington State Sediment Management Standards (Sediment Quality Standard, SQS, and Cleanup Screening Level, CSL).

**Table 9. Method 4020 PCB and QC Results, Dry Weight.**

Field Station ID	SAMPLE LABEL	tPCB ( $\mu\text{g}/\text{Kg}$ )	Q	Stdev	%RSD
SD-307	SD-307-0000	2615		218	8.33%
SD-311	SD-311-0001	1200		324	27.04%
	SD-313-0002	33	U	6	17.35%
SD-316	SD-317-0000	349		7	2.12%
	SD-322-0002	1273		66	5.17%

Stdev: Standard Deviation from duplicate assay analyses ( $n=2$ );

% RSD: Percent Relative Standard Deviation [ $\{\text{stdev}/\text{mean}\} * 100\%$ ];

Q: Data Qualifiers where; U =Non-Detect;

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**Table 10. Method 8082 Total PCB Field Duplicate Results**

Field Station ID	SAMPLE LABEL	tPCB Dry Weight (mg/Kg)	Q	RPD	TOC%	tPCB mg/kg OC	RPD
SD-207	SD-207-0000	145.6	JE	20.0%	2.11	6.9	15.0%
	SD-207-0000(FD)	97	U		1.56	6.2	
SD-323	SD-323-0001	284.7		47.1%	1.65	17.3	2.4%
	SD-323-0001(FD)	792.3			1.73	45.8	
SD-309	SD-309-0002	538.6		1%	1.32	40.8	1%
	SD-309-0002(FD)	550.8			1.32	41.7	
SD-314	SD-314-0000	670		12%	1.66	40.4	10%
	SD-314-0000(FD)	850			1.73	49.1	
SD-333	SD-333-0000	1000		53%	2.24	44.6	54%
	SD-333-0000(FD)	303.4			2.24	13.5	

RPD =Percent Relative Difference:  $(|A-B|/(A+B)) * 100\%$

Table 11. Total PCB Results from Method 4020, Dry-weight and OC-normalized

*Italic and underlined* is > CSL. **Bolded and Shaded** is SQS<x<CSL, and Values < SQS are Normal Text

Field Station ID	Sample Label	tPCB (ug/Kg)	TOC Q (fraction)	PCB (mg/kgOC)	Field Station ID	Sample Label	tPCB (ug/Kg)	TOC Q (fraction)	PCB (mg/kgOC)	Field Station ID	Sample Label	tPCB (ug/Kg)	TOC Q (fraction)	PCB (mg/kgOC)			
SD-206	SD-206-0000	298	0.0234	<u>12.7</u>	SD-312	SD-312-0000	510	0.0258	19.8	SD-321	SD-321-0001	217	0.0225	9.6			
SD-207	SD-207-0000	259	0.0211	<u>12.3</u>		SD-312-0001	761	0.0109	<b>69.8</b>		SD-321-0002	425	0.0169	<u>25.1</u>			
SD-208	SD-208-0000	443	0.0146	<u>30.3</u>		SD-312-0002	66	U	0.00134	<u>49.3</u>		SD-321-0003	224	0.0196	<u>11.1</u>		
SD-209	SD-209-0000	210	0.0271	7.7		SD-312-0003	14	U	0.0013	<u>10.8</u>	SD-322	SD-321-0003	145	J	0.0174	8.3	
SD-210	SD-210-0000	187	0.0265	7.1		SD-312-0003(FD)	149	J	0.0172	8.7		SD-322-0000	186		0.0202	9.2	
SD-211	SD-211-0000	248	0.0216	11.5	SD-313	SD-313-0000	432		0.0222	<u>19.5</u>		SD-322-0001	583		0.019	<u>30.7</u>	
SD-212	SD-212-0000	204	0.0233	8.8		SD-313-0001	75	J	0.0181	4.1		SD-322-0002	1273		0.0263	<u>48.4</u>	
SD-213	SD-213-0000	291	0.0217	<u>15.4</u>		SD-313-0002	33	U	0.00947	3.5		SD-322-0003	514		0.0272	<u>18.9</u>	
SD-214	SD-214-0000	165	0.0278	5.9		SD-313-0003	17	U	0.0118	1.4		SD-322-0004	138	J	0.0144	9.6	
SD-215	SD-215-0000	515	0.0164	<u>31.4</u>	SD-314	SD-314-0000	505		0.0166	<u>30.4</u>		SD-322-0005	81	J	0.00852	9.5	
SD-216	SD-216-0000	195	0.0202	9.7		SD-314-0001	12	U	0.0244	0.5	SD-323	SD-323-0000	2043		0.0249	<u>62</u>	
SD-217	SD-217-0000	705	0.0184	<u>38.3</u>		SD-314-0002	6	U	0.00237	2.5		SD-323-0001	266		0.0165	<u>16.1</u>	
SD-307	SD-307-0000	2615	0.022	<b>118.9</b>		SD-314-0003	3	U	0.00428	0.7		SD-323-0002	63	U	0.0221	2.9	
SD-307-0001		56	U	0.008	7	SD-315	SD-315-0000	183		0.0157	11.7		SD-323-0003	43	U	0.0131	3.3
SD-307-0002		97	J	0.00226	<u>42.9</u>		SD-315-0001	0.3	U	0.00218	0.1	SD-330	SD-330-0000	359		0.0215	<u>16.2</u>
SD-307-0003		191	J	0.00317	<u>60.3</u>		SD-315-0002	3	U	0.00305	1	SD-331	SD-331-0000	446		0.0243	<u>18.4</u>
SD-309	SD-309-0000	420	0.0221	<u>19</u>		SD-315-0003	0.04	U	0.00183	0	SD-332	SD-332-0000	425		0.0224	<u>19</u>	
SD-309-0001		682		0.0177	<u>38.5</u>	SD-316	SD-316-0000	405		0.0225	<u>18</u>	SD-333	SD-333-0000	783		0.025	<u>31.3</u>
SD-309-0002		486		0.0132	<u>36.8</u>		SD-316-0001	564		0.0166	<u>34</u>	SD-334	SD-334-0000	305		0.0155	<u>19.2</u>
SD-309-0002(FD)		435		0.0166	26.2		SD-316-0002	671		0.0187	<u>35.9</u>	SD-335	SD-335-0000	238		0.0198	<u>12</u>
SD-309-0003		173		0.00941	<u>18.4</u>		SD-316-0003	3089		0.0153	<b>201.9</b>	SD-336	SD-336-0000	449		0.0155	<u>29</u>
SD-310	SD-310-0000	889		0.0198	<u>44.9</u>	SD-317	SD-317-0000	349		0.0258	<u>13.5</u>	SD-337	SD-337-0000	1200		0.0192	<u>62.5</u>
SD-310-0001		365		0.0154	<u>23.7</u>		SD-317-0001	823		0.02	<u>41.2</u>	SD-338	SD-338-0000	474		0.0181	<u>26.2</u>
SD-310-0002		2000		0.0157	<u>13.3</u>		SD-317-0002	6485		0.0211	<b>307.3</b>	SD-338	SD-338-0000FD	393		0.0181	<u>21.7</u>
SD-310-0003		731	U	0.00986	7.4		SD-317-0003	898		0.00816	<u>110</u>	SD-339	SD-339-0000	263		0.017	<u>15.5</u>
SD-311	SD-311-0000	461		0.0184	<u>25.1</u>	SD-318	SD-318-0000	408		0.0205	<u>19.9</u>	SD-340	SD-340-0000	415		0.0161	<u>25.8</u>
SD-311-0001		1200		0.0106	<b>113.2</b>		SD-318-0001	16	U	0.00824	1.9	SD-341	SD-341-0000	3186		0.0268	<b>118.9</b>
SD-311-0002		5072		0.0145	<u>349.8</u>	SD-319	SD-319-0000	1754		0.0174	<b>100.8</b>	SD-342	SD-342-0000	704		0.0145	<u>48.6</u>
SD-311-0003		2003		0.00725	<u>276.3</u>		SD-319-0001	167		0.018	9.3	SD-343	SD-343-0000	761		0.0196	<u>38.8</u>
							SD-319-0002	122	J	0.0222	5.5	SD-344	SD-344-0000	216		0.0258	8.4
							SD-319-0003	109	J	0.0181	6	SD-345	SD-345-0000	526		0.0124	<u>42.4</u>
						SD-320	SD-320-0000	2143		0.0162	<b>132.3</b>						
						SD-320-0001	745		0.0224	<u>33.3</u>							
						SD-320-0002	368		0.0239	<u>15.4</u>							
						SD-320-0003	116	J	0.0183	6.3							

Table 12. Total PCB Results from Method 8082, Dry-weight and OC-normalized

*Italic and Underlined* is SQS< x < CSL; **Bold** and **Shaded** is > CSL, and values < SQS are Normal Text; FD=Field Duplicate

Field Station	Sample ID	tPCB (ug/Kg dry)	tPCB (mg/kg OC)	Q	Field Station ID	Sample Label	tPCB (ug/Kg dry)	tPCB (mg/kg OC)	Q	Field Station ID	Sample Label	tPCB (ug/Kg dry)	tPCB (mg/kg OC)	Q					
SD-206	SD-206-0000	280	2.34	12		SD-313-0000	1150	<i>JE</i>	2.22	<b>51.8</b>	J	SD-322	SD-322-0000	110	2.02	5.4			
SD-207	SD-207-0000	145.6	E	2.11	6.9	SD-313-0001	64.2		1.81	3.5		SD-322	SD-322-0001	963.2	E	1.9	<b>50.7</b>	E	
	SD-207-0000(FD)	97	U	1.56	6.2	SD-313-0002	12	U	0.94	1.3	U	SD-322	SD-322-0002	2773		2.63	<b>103.4</b>		
SD-208	SD-208-0000	340.7	E	1.46	<u>23.3</u>	SD-313-0003	120	U	1.18	10.2	U	SD-322	SD-322-0003	1391		2.72	<u>51.1</u>		
SD-209	SD-209-0000	77.7	<u>JE</u>	2.71	2.9	SD-314-0000	670	E	1.66	<u>40.4</u>		SD-322	SD-322-0004	126.1		1.44	8.8		
SD-210	SD-210-0000	130		2.65	4.9	SD-314-0000(FD)	850		1.73	<u>49.1</u>		SD-322	SD-322-0005	231.8	E	0.85	<u>27.3</u>	E	
SD-211	SD-211-0000	610		2.16	<b>28.2</b>	SD-314-0001	20	U	2.44	0.8	U	SD-323	SD-323-0000	9400	J	2.49	<b>377.5</b>	J	
SD-212	SD-212-0000	48.9	E	2.33	2.1	SD-314-0002	580	U	0.23	<b>252.2</b>	U	SD-323	SD-323-0001	284.7		1.65	<u>17.3</u>		
SD-213	SD-213-0000	610	J	2.17	<u>28.1</u>	SD-314-0003	224	J	0.42	<u>53.3</u>	J	SD-323	SD-323-0003	792.3	E	1.73	<u>45.8</u>		
SD-214	SD-214-0000	8.7	J	2.78	0.3	SD-315-0000	260	J	1.57	<u>16.6</u>	J	SD-324	SD-324-0000	1.2	J	1.31	0.1	J	
SD-215	SD-215-0000	880		1.64	<u>53.7</u>	SD-315-0001	22.6	U	0.21	10.8	U	SD-325	SD-325-0000	850		2.23	<u>38.1</u>		
SD-216	SD-216-0000	360	J	2.02	<u>17.8</u>	SD-315-0002	.91	U	0.3	<u>30.3</u>	U	SD-326	SD-326-0002	550.8		1.58	<u>34.9</u>		
SD-217	SD-217-0000	292.7	<u>JE</u>	1.84	<u>15.9</u>	SD-315-0003	67	U	0.18	<u>37.2</u>	U	SD-330	SD-330-0000	680	J	1.72	<u>39.5</u>	J	
SD-307	SD-307-0000	2600		2.2	<b>118.2</b>	SD-316	SD-316-0000	940	2.25	<u>41.8</u>		SD-331	SD-331-0000	361.2	E	2.15	<u>16.8</u>	E	
SD-307	SD-307-0001	81.8	E	0.8	10.2	SD-316	SD-316-0001	735	1.66	<u>44.3</u>		SD-332	SD-332-0000	361.2	E	2.24	<u>16.1</u>	E	
SD-307	SD-307-0002	162.1	E	0.22	<b>73.7</b>	SD-316	SD-316-0002	779	E	1.87	<u>41.7</u>	E	SD-333	SD-333-0000	1000		2.24	<u>44.6</u>	
SD-307	SD-307-0003	70.4	E	0.31	<u>22.7</u>	SD-316	SD-316-0003	3065	1.53	<b>200.3</b>		SD-333	SD-333-0000(FD)	303.4	E	2.24	<u>13.5</u>		
SD-309	SD-309-0000	570		2.21	<u>25.8</u>	SD-317	SD-317-0000	800	2.58	<u>31</u>		SD-334	SD-334-0000	290		1.55	<u>18.7</u>		
SD-309	SD-309-0001	253.1	E	1.77	<u>14.3</u>	SD-317	SD-317-0001	1529	2	<b>76.5</b>		SD-335	SD-335-0000	400		1.98	<u>20.2</u>		
SD-309	SD-309-0002	538.6		1.32	<u>40.8</u>	SD-317	SD-317-0002	10438	2.11	<b>494.7</b>		SD-336	SD-336-0000	251	J	1.55	<u>16.2</u>	J	
SD-309	SD-309-0002(FD)	550.8		1.32	<u>41.7</u>	SD-317	SD-317-0003	1635	0.81	<b>201.9</b>		SD-337	SD-337-0000	1238	JE	1.92	<u>64.5</u>	E	
SD-309	SD-309-0003	127.2		0.94	<u>13.5</u>	SD-318	SD-318-0000	930	J	2.05	45.4	J	SD-338	SD-338-0000	430		1.81	<u>23.8</u>	
SD-310	SD-310-0000	560		1.98	<u>28.3</u>	SD-318	SD-318-0001	13	U	0.82	1.6	U	SD-339	SD-339-0000	480		1.7	<u>28.2</u>	
SD-310	SD-310-0001	274		1.54	<u>17.8</u>	SD-319	SD-319-0000	3100	1.74	<b>178.2</b>		SD-340	SD-340-0000	230		1.61	14.3		
SD-310	SD-310-0002	159.7		1.5	10.6	SD-319	SD-319-0001	120		1.8	6.7		SD-341	SD-341-0000	1398	E	2.68	<u>52.2</u>	E
SD-310	SD-310-0003	65.2	E	0.98	6.7	SD-319	SD-319-0002	132.9	E	2.22	6	E	SD-342	SD-342-0000	3639	JE	1.45	<u>251</u>	E
SD-311	SD-311-0000	3300	J	1.84	<u>179.3</u>	SD-319	SD-319-0003	13	U	1.81	0.7	U	SD-343	SD-343-0000	260	J	1.96	<u>13.3</u>	J
SD-311	SD-311-0001	1586	E	1.06	<u>149.6</u>	SD-320	SD-320-0000	8864	1.62	<b>547.2</b>		SD-344	SD-344-0000	11000		2.58	<b>426.4</b>		
SD-311	SD-311-0002	5789	E	1.45	<u>399.2</u>	SD-320	SD-320-0001	1481.2		2.24	<u>66.1</u>		SD-345	SD-345-0000	182.1	E	1.24	<u>14.7</u>	E
SD-311	SD-311-0003	4446	E	0.72	<u>617.5</u>	SD-320	SD-320-0002	1234.1		2.39	<u>51.6</u>								
SD-312	SD-312-0000	1200		2.58	46.5	SD-320	SD-320-0003	238.9		1.83	<u>13.1</u>								
SD-312	SD-312-0001	1869	E	1.09	<u>171.5</u>	SD-321	SD-321-0000 (FD)	510	J	2.25	<u>22.7</u>	J							
SD-312	SD-312-0002	6.3	J	0.13	4.8	SD-321	SD-321-0001	751.5		1.69	<u>44.5</u>								
SD-312	SD-312-0003	0.52	J	0.13	0.4	SD-321	SD-321-0002	358.4		1.96	<u>18.3</u>								
						SD-321	SD-321-0003	292	E	1.74	<u>16.8</u>	E							

The E qualifier notes that QC records were not available for inspection from EPA Region 8 Laboratory, so that the data are of unknown quality and should be used with caution. See text and Appendix G.

Ninety-nine PCB samples were measured by both methods. Seventeen samples in the 8082 results exceeded CSL; twelve samples in the 4020 results exceeded CSL. For a detailed comparison of data between Methods 4020 and 8082 (i.e., the Demonstration of Method Applicability), see the following URL:

[http://www.triadcentral.org/user/includes/dsp\\_profile.cfm?Project\\_ID=23](http://www.triadcentral.org/user/includes/dsp_profile.cfm?Project_ID=23)

Thoroughness of documentation for Method 8082 varied by laboratory, as described in Appendix G. For Region 10 Laboratory, good documentation was available, and the review was thorough and qualifiers were applied appropriately. A number of samples had elevated detection limits, and a number had surrogate recoveries that were somewhat out of range. The data evaluation concludes that the data are usable.

The Region 6 case narrative is also thorough, although the evaluation report states inaccurately that the samples were received past holding time, and mis-states that some sample delivery group results were outside Aroclor recovery advisory limits although they were not out of the limits stated in the QAPP. Some high bias from Aroclor 1248 was noted. The data are usable as qualified.

The Region 8 case narrative is lacking detail, and it was not possible to recover the records despite considerable help from the EPA Region 10 Manchester Laboratory staff. The narrative incorrectly reports surrogate recoveries for the samples as Aroclors. The surrogate advisory limits for the DCBP surrogate were not exceeded (see Appendix G) as stated in the narrative. These data are not of known quality due to the lack of documentation, and have been flagged "E" (a non-standard flag) suggesting that they should be used with extreme caution for that reason. The "J" qualifier has been removed, as discussed in Appendix G.

## References

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- Puget Sound Estuarine Program (PSEP). 1996. Recommended protocols for measuring selected environmental variables in Puget Sound. U.S. Environmental Protection Agency, Office of Coastal Waters.
- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C. December 1986.
- U.S. EPA. 2004. USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review. OSWER 9240.1-45. Office of Superfund Remediation and Technology Innovation (OSRTI), Washington DC. October 2004.
- USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.
- WAC 173-204. 1995. Sediment management standards. Washington Administrative Code. December 29, 1995.

## **Appendix A: Sediment Sample Records**



2200 Sixth Avenue, Suite 1100  
Seattle, Washington 98121  
(206) 441-9080  
FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad

DATES: 8/16/04 – 8/19/04, 8/26/04-8/27/04

LOCATION: Duwamish River – offshore of Boeing Plant 2

CREW: JW, RZ, RB, AS, GC, BC, VH GEAR:

Van Veen (surface sampling)  
Vibrocorer (subsurface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL * (ft)	CHARACTERISTICS (COLOR, TYPE, DEBRIS, ODOR)
SD-307	8/16/04	17:04	A	11.5	NA	0 – 0.33	Medium brown surface. Red/rusty brown fine to medium SAND with silt from 0 to 0.13 ft. Dark gray/black SILT with trace sand from 0.13 to 0.33 ft. Some twigs and roots throughout, concentrated in bottom layer.
	8/19/04	09:00	C	13.1	65 E	1 – 2	Very dark gray fine to medium SAND. Sample not solidly packed in core tube. Limited sample volume.
						2 – 3	Very dark gray fine SAND.
						3 – 4	Very dark gray fine SAND.
SD-309	8/16/04	17:23	A	11.8	NA	0 – 0.33	Slight sheen on water. Medium brown silty SAND with patches of rusty red from 0 to 0.07 ft. Dark gray/black sandy SILT with some angular rocky pieces, possibly slag, from 0.07 to 0.33 ft.
	8/19/04	11:15	A	10.5	100	1 – 2	Black-clayey SILT with trace gravel.
						2 – 3	Black clayey SILT with trace gravel to 2.75 ft. Black silty SAND with woody debris from 2.75 to 3 ft.
						3 – 4	Black silty SAND with woody debris.
SD-310	8/16/04	16:43	B	11.8	NA	0 – 0.33	Medium brown surface with algae. Dark gray SILT with some sand subsurface. Concrete or slag debris.
	8/19/04	11:45	A	8.17	81	1 – 2	Very dark gray/black alternating layers of fine-grained SAND, organic SILT, and clayey SILT.
						2 – 3	Very dark gray/black alternating layers of fine-grained SAND, organic SILT, and clayey SILT.
						3 – 4	Very dark gray/black alternating layers of fine-grained SAND, organic SILT, and clayey SILT to 3.3 ft. Very dark gray fine-grained SAND with some medium-grained sand with mica flakes from 3.3 to 4.0 ft.
SD-311	8/16/04	17:40	A	12.5	NA	0 – 0.33	Clam shell and mud shrimp on surface. Medium to light brown SILT from 0 to 0.13 ft. Dark gray/black SILT with some woody debris and rocks from 0.13 to 0.33 ft. Rock caught in sampler jaws.
	8/19/04	16:51	A	7.9	100	1 – 2	Black clayey SILT with gravel.
						2 – 3	Black clayey SILT with gravel. Brick fragment at 3 ft.
						3 – 4	Black silty SAND.
SD-312	8/16/04	18:23	D	13.5	NA	0 – 0.33	Algae layer on surface. Medium brown SILT from 0 to 0.13 ft. Gray to black silty fine SAND with woody debris (creosote rich) from 0.13 to 0.33 ft. Slight hydrogen sulfide odor and sheen present in bottom layer.
	8/19/04	17:00	A	7.7	64	1 – 2	Black sandy SILT with woody debris (4" and smaller).
						2 – 3	Black silty SAND with 1" dark black layer at 2.7 ft.
						3 – 4	Black silty SAND.



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Seattle, Washington 98121  
(206) 441-9080  
FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad

DATES: 8/16/04 – 8/19/04, 8/26/04-8/27/04

LOCATION: Duwamish River – offshore of Boeing Plant 2

CREW: JW, RZ, RB, AS, GC, BC, VH GEAR: Van Veen (surface sampling)  
Vibrocorer (subsurface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL <sup>a</sup> (ft)	CHARACTERISTICS (COLOR, TYPE, DEBRIS, ODOR)
SD-313	8/16/04	19:02	G	13.8	NA	0 – 0.33	Brick on surface. Medium brown SILT with iron granules and shell fragments from 0 to 0.13 ft. Dark gray to black SILT with some sand and gravel (possibly slag) from 0.13 to 0.33 ft.
	8/19/04	19:58	C	14.4	56	1 – 2	Black clayey SILT.
						2 – 3	Black clayey SILT to 2.3 ft. Black silty SAND from 2.3 to 3 ft.
						3 – 4	Black silty SAND.
SD-314	8/17/04	16:45	A	12.5	NA	0 – 0.33	Olive brown fine sand on surface. Dark gray clayey SILT with some small and large rocks to 0.33 ft. Chunks of asphalt present throughout sample.
	8/18/04	18:07	B	13.8	60	1 – 2	Brown silty CLAY from 1.0 to 1.1 ft. Dark brown silty SAND from 1.1 to 2.0 ft.
						2 – 3	Light gray clayey SILT from 2.0 to 2.1 ft. Dark gray/black fine to medium SAND from 2.1 to 3.0 ft.
						3 – 3.3	Dark brown silty SAND.
SD-315	8/17/04	17:46	B	11.8	NA	0 – 0.33	Light brown SAND on surface. Dark grayish brown SILT with some sand, clay and fine woody debris.
	8/19/04	18:52	A	10.9	71	1 – 2	Dark brownish gray sandy SILT with trace woody debris from 1.0 to 1.2 ft. Black silty SAND from 1.2 to 2.0 ft.
						2 – 3	Black silty SAND.
						3 – 4	Black silty SAND.
SD-316	8/16/04	15:57	B	9.8	NA	0 – 0.33	Light brown surface with algae cover. Dark gray SILT with trace fine sand and woodchips.
	8/19/04	10:40	A	11.25	100	1 – 2	Black silty CLAY.
						2 – 3	Black silty CLAY.
						3 – 4	Black silty CLAY from 3.0 to 3.2 ft. Black silty SAND with trace clay and 2" piece of asphalt-like material from 3.2 to 4.0 ft.
SD-317	8/16/04	16:26	A	11.5	NA	0 – 0.33	Brown surface with algae cover. Dark gray sandy SILT with trace wood chips. No odor or sheen.
	8/19/04	09:55	B	11.75	78 E	1 – 2	Black silty CLAY from 1.0 to 1.4 ft. Black sandy SILT with trace clay from 1.4 to 2.0 ft.
						2 – 3	Black sandy SILT with trace clay.
						3 – 4	Black sandy SILT with gravel.



2200 Sixth Avenue, Suite 1100  
 Seattle, Washington 98121  
 (206) 441-9080  
 FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad

DATES: 8/16/04 – 8/19/04, 8/26/04-8/27/04

LOCATION: Duwamish River – offshore of Boeing Plant 2

CREW: JW, RZ, RB, AS, GC, BC, VH

GEAR: Van Veen (surface sampling)  
 Vibrocorer (subsurface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL <sup>a</sup> (ft)	CHARACTERISTICS (COLOR, TYPE, DEBRIS, ODOR)
SD-318	8/16/04	16:12	A	10.2	NA	0 – 0.33	Light brown surface with algae. Dark gray SILT with trace sand and wood debris. No odor or sheen.
	8/18/04	17:00	B	9.2	67 E	0 – 1.5	Very dark gray SAND with mica. No debris.
SD-319	8/16/04	15:17	A	13.8	NA	0 – 0.33	Drab olive brown surface. Dark gray sandy (fine to medium) SILT with some small woodchips. No shells, odor, sheen, or debris.
	8/18/04	16:10	A	28.9	77 E	1 – 2	Black clayey SILT.
						2 – 3	Black clayey, sandy SILT with black sandy layer from 2.4 to 2.5 ft.
						3 – 4	Black clayey SILT from 3.0 to 3.5 ft. Black silty SAND from 3.5 to 4.0 ft.
SD-320	8/16/04	18:47	A	12.8	NA	0 – 0.33	Large, barnacle-covered rock (4") and algae on surface. Medium brown SILT with iron granules from 0 to 0.07 ft. Dark gray to black SILT with some sand, woody debris, and medium to large rocks from 0.07 to 0.33 ft. Isopod found in sediment.
	8/19/04	17:33	A	9.0	83 E	1 – 2	Black clayey SILT with brick, slag, and plastic wire debris to 1.2 ft. Black clayey SILT with some sand from 1.2 to 2.0 ft.
						2 – 3	Black clayey SILT with some sand.
						3 – 4	Black clayey SILT with some sand.
SD-321	8/16/04	14:23	A	19.0	NA	0 – 0.33	Drab olive brown surface. Dark gray SILT to 0.33 ft. Worm in sediment. No odor or sheen. Field duplicate collected (SD-324-0000).
	8/18/04	15:25	A	23.0	69 E	1 – 2	Black clayey SILT from 1.0 to 1.1 ft. Black silty SAND from 1.1 to 1.8 ft. Black clayey silt from 1.8 to 2.0 ft.
						2 – 3	Black clayey SILT with some sand.
						3 – 3.8	Black silty SAND.
SD-322	8/16/04	14:55	A	17.75	NA	0 – 0.33	Brown surface with dark gray SILT to 0.33 ft. Worms in sediment. No odor or sheen.
	8/18/04	11:20	C	21.25	73 E	1 – 2	Black silty CLAY.
						2 – 3	Black silty CLAY with woody debris.
						3 – 4	Black silty SAND with woody debris from 3 to 3.5 ft.
						4 – 5	Black silty SAND.
						5 – 6	Black silty SAND.



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(206) 441-9080  
FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad DATES: 8/16/04 – 8/19/04, 8/26/04-8/27/04  
LOCATION: Duwamish River – offshore of Boeing Plant 2 CREW: JW, RZ, RB, AS, GC, BC, VH GEAR: Van Veen (surface sampling)  
Vibrocorer (subsurface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL <sup>a</sup> (ft)	CHARACTERISTICE (COLOR, TYPE, DEBRIS, ODOR)
SD-323	8/17/04	16:30	B	7.8	NA	0 – 0.33	Medium brown on surface. Dark brown sandy silt with roots.
	8/19/04	18:27	A	11.35	60	1 – 2	Black sandy SILT with gravel.
						2 – 3	Black sandy SILT from 2.0 to 2.1 ft. Black silty CLAY with trace amounts of organic debris from 2.1 to 2.5 ft. Black silty CLAY with large pieces of woody debris from 2.5 to 3.0 ft.
						3 – 4	Black silty CLAY with large pieces of woody debris from 3.0 to 2.1 ft. Black silty SAND from 3.1 to 4.0 ft.

REP. NO.: Replicate number represents the sequential attempt at each station (Rep B is the second attempt).

% REC: Percent recovery of subsurface core. Value was determined by dividing the total depth of sediment in core tube by the total depth of core tube penetration and multiplying by 100.

NA: Not applicable.

Percent recovery should be considered an estimate. Depth of core tube penetration was not based on actual measurement but was estimated based on physical indications on the core tube.

<sup>a</sup>: Sample intervals were not adjusted based on percent recovery of the cores. The top of the sediment in each core tube was considered zero in measuring sample interval depth.



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FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad

DATES: 8/26/04 and 8/27/040

LOCATION: Duwamish River - offshore of Boeing Plant 2

CREW: BC, AS

GEAR: Van Veen (surface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL (ft)	CHARACTERISTICE (COLOR, TYPE, DEBRIS, ODOR)
SD-206	8/26/04	12:30	A	13.8	NA	0 - 0.33	Light brown sandy SILT on surface (top 0.5 in). Black clayey SILT with small wood chips to 0.33 ft.
SD-207	8/26/04	13:12	A	17.7	NA	0 - 0.33	Light brown sandy SILT on surface (top 0.5 in). Black clayey SILT with organic material to 0.33 ft.
SD-208	8/26/04	13:35	A	18.4	NA	0 - 0.33	Light brown sandy SILT (top 1 in). Black clayey SILT with organic material to 0.33 ft.
SD-209	8/26/04	13:55	B	24.6	NA	0 - 0.33	Light brown sandy SILT (top 0.5 in). Black clayey SILT to 0.33 ft. Red worm found in sample.
SD-210	8/27/04	10:00	A	11.8	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray clayey SILT with some worms to 0.33 ft.
SD-211	8/27/04	10:20	A	9.8	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray clayey SILT with some worms and benthic shrimp to 0.33 ft.
SD-212	8/27/04	10:45	A	11.8	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray clayey SILT with worms to 0.33 ft.
SD-213	8/27/04	11:00	A	9.2	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray sandy SILT with woody debris and worms to 0.33 ft.
SD-214	8/27/04	11:20	A	14.4	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray clayey SILT with some woody debris and worms to 0.33 ft.
SD-215	8/27/04	12:35	A	15.7	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Black clayey SILT with organic material to 0.33 ft.
SD-216	8/26/04	18:25	A	23.6	NA	0 - 0.33	Light brown silty SAND (top 0.5 in). Dark brown clayey SILT to 0.33 ft.
SD-217	8/26/04	18:10	A	22.3	NA	0 - 0.33	Light brown silty SAND (top 0.5 in). Dark brown sandy SILT with woody debris to 0.33 ft.
SD-330	8/27/04	13:08	A	11.8	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Black clayey SILT with worms to 0.33 ft.
SD-331	8/27/04	13:25	A	12.5	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Black clayey SILT with woody debris to 0.33 ft.
SD-332	8/26/04	18:55	A	14.4	NA	0 - 0.33	Light brown silty SAND (top 0.5 in). Black clayey SILT with organic matter to 0.33 ft.
SD-333	8/27/04	15:45	C	14.6	NA	0 - 0.33	Brown silty SAND (top 1 in). Gray sandy SILT with organic matter and some gravel to 0.33 ft.
SD-334	8/26/04	17:11	A	16.4	ND	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray silty SAND with trace gravel and trace organic debris to 0.33 ft.
SD-335	8/27/04	12:50	A	15.7	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Black clayey SILT with organic material to 0.33 ft.
SD-336	8/27/04	13:40	A	11.2	NA	0 - 0.33	Brown silty SAND (top 1 in). Dark gray sandy SILT some woody debris and gravel to 0.33 ft.
SD-337	8/27/04	14:07	C	11.8	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Black sandy SILT with minor gravel to 0.33 ft.
SD-338	8/26/04	16:50	F	13.8	NA	0 - 0.33	Brown silty SAND (top 1 in). Black clayey SILT with worms, some organic matter and small fragments of woody debris to 0.33 ft.
SD-339	8/26/04	16:15	F	14.4	NA	0 - 0.33	Brown silty SAND (top 0.5 in). Dark gray silty CLAY to 0.33 ft.
SD-340	8/26/04	18:38	A	15.7	NA	0 - 0.33	Light brown silty SAND (top 0.5 in). Dark brown clayey SILT to 0.33 ft.
SD-341	8/26/04	15:15	C	13.1	NA	0 - 0.33	Brown gravelly SAND with silt, woody debris. Macrophytic algae on surface.



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(206) 441-9080  
FAX (206) 441-9108

## SEDIMENT SAMPLE RECORD

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PROJECT NAME: Lower Duwamish Triad

DATES: 8/26/04 and 8/27/04

LOCATION: Duwamish River – offshore of Boeing Plant 2

CREW: BC and AS

GEAR: Van Veen (surface sampling)

STATION LOCATION	DATE	TIME	REP. NO.	WATER DEPTH (ft)	% REC	SAMPLE INTERVAL (ft)	CHARACTERISTICS (COLOR, TYPE, DEBRIS, ODOR)
SD-342	8/27/04	14:36	A	2.6	NA	0 – 0.33	Brown silty SAND (top 0.5 in). Black clayey SILT with some woody debris to 0.33 ft.
SD-343	8/27/04	14:57	A	13.1	NA	0 – 0.33	Brown silty SAND (top 1 in). Black sandy SILT with large woody debris fragments and some gravel to 0.33 ft. Sulfur odor present.
SD-344	8/26/04	15:00	D	13.8	NA	0 – 0.33	Homogenous brown-orange gravelly SAND with rust mottling; silt; sticks, woody debris metal fork.
SD-345	8/26/04	17:30	B	14.4	NA	0 – 0.33	Brown silty SAND (top 0.5 in). Dark gray silty SAND to 0.33 ft.

REP. NO.: Replicate number represents the sequential attempt at each station (Rep B is the second attempt).

% REC: Percent recovery of subsurface core. Value was determined by dividing the total depth of sediment in core tube by the total depth of core tube penetration and multiplying by 100.

NA: Not applicable.

## **Appendix B: Summary Photographic Documentation**

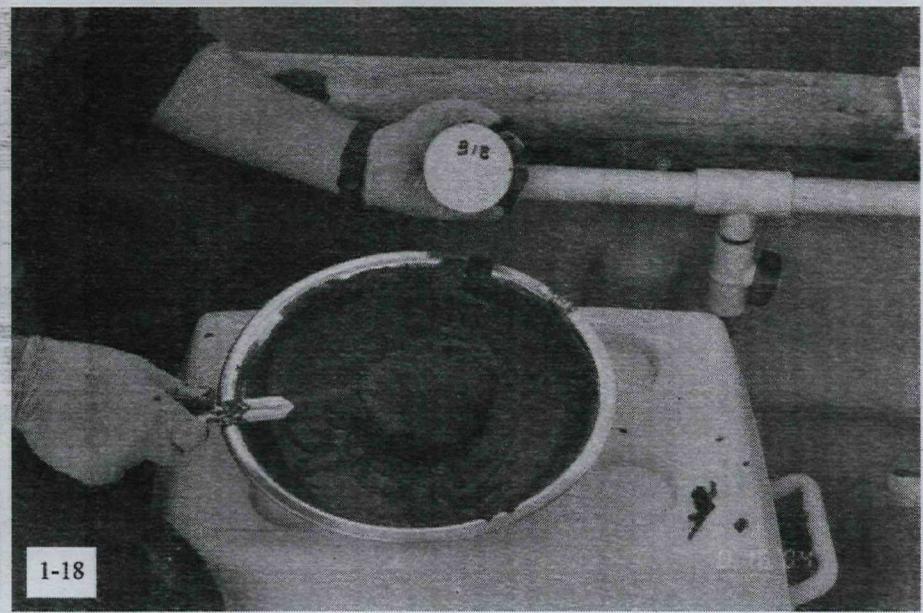
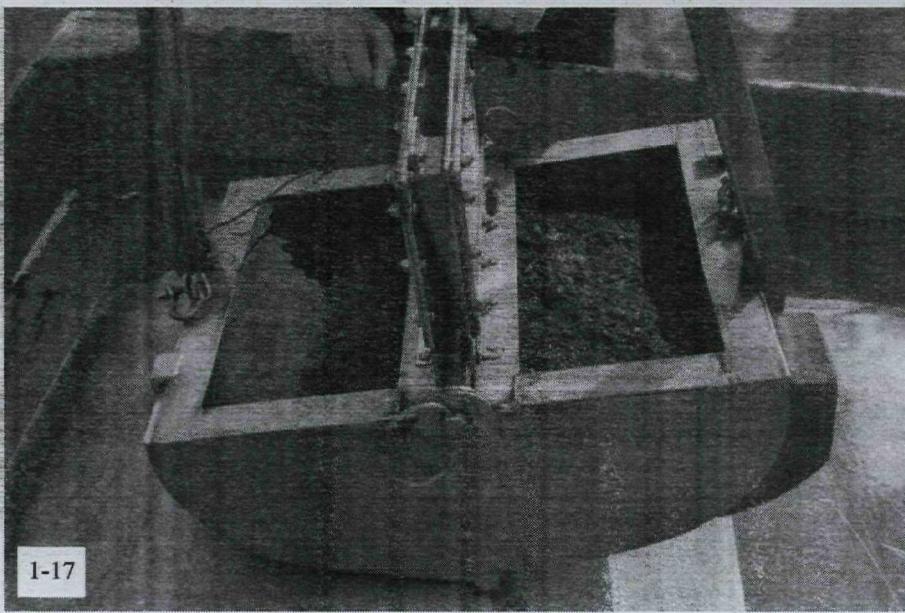
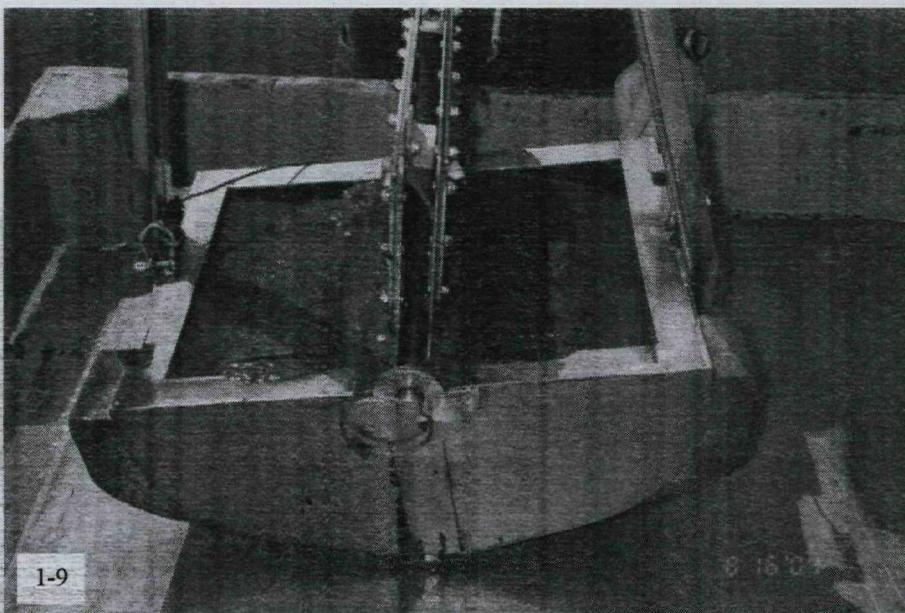
**Lower Duwamish Triad Sampling Event  
Seattle, Washington  
Summary Photographic Log**

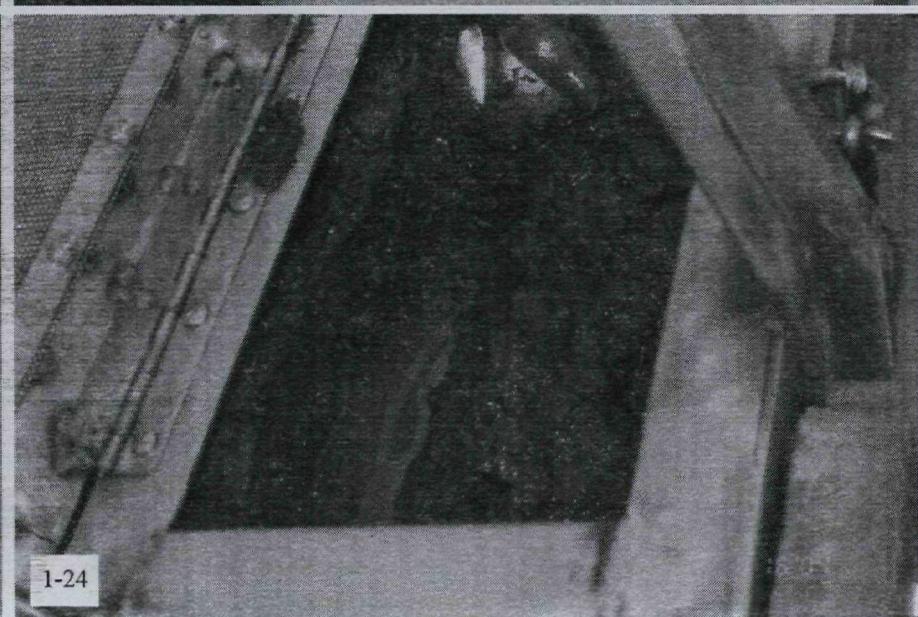
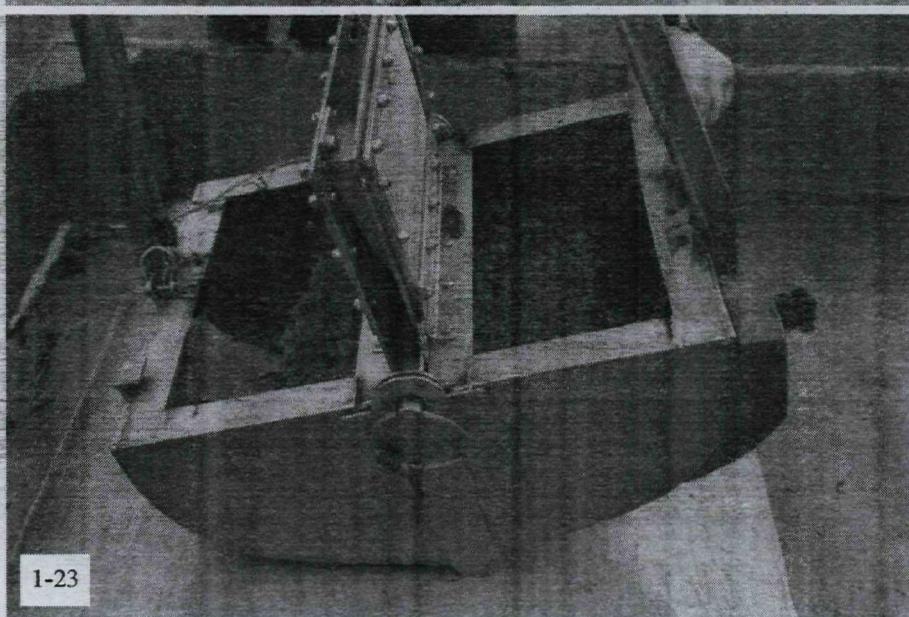
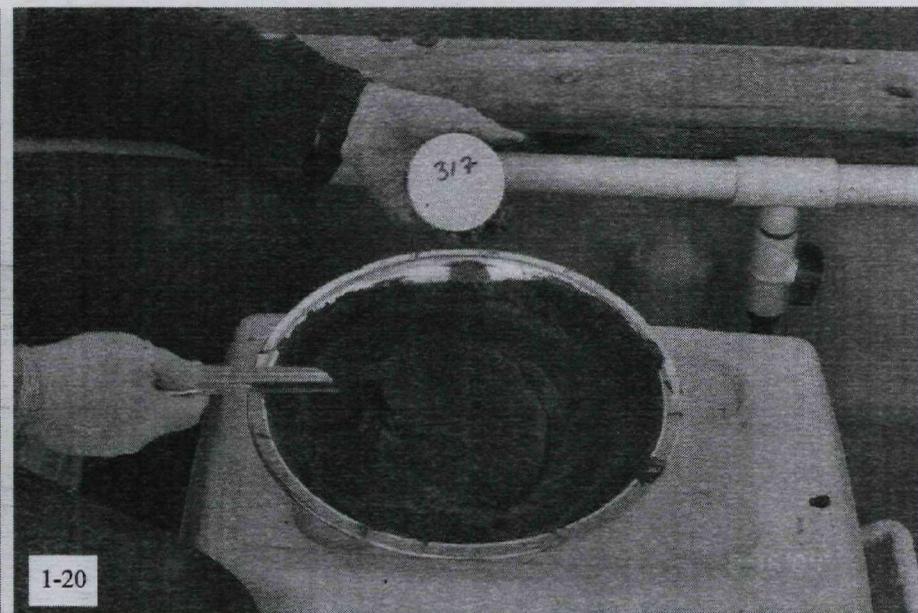
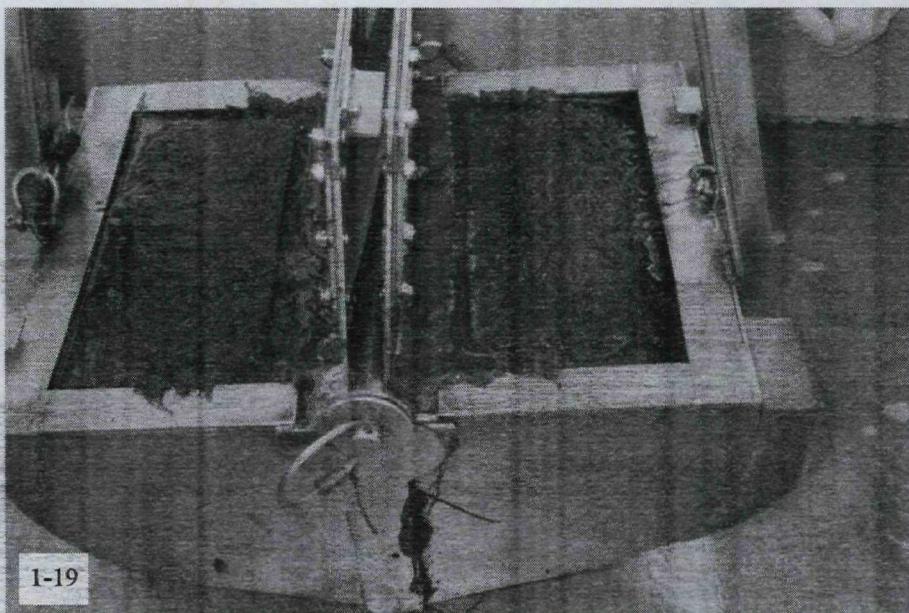
Photo Number	Photo Description
1-9	Van Veen surface sample grab at station SD-322, replicate A.
1-10	Homogenized sample SD-322-0000 in stainless steel bowl.
1-17	Van Veen surface sample at station SD-318, replicate A, with both access doors open.
1-18	Homogenized sample SD-318-0000 in stainless steel bowl.
1-19	Van Veen surface sample at station SD-317, replicate A.
1-20	Homogenized sample SD-317-0000 in stainless steel bowl.
1-23	Van Veen surface sample at station SD-307, replicate A.
1-24	Sediment layers in sample SD-307-0000.
2-1	Van Veen surface sample at station SD-309, replicate A.
2-2	Homogenized sample SD-309-0000 in stainless steel bowl.
2-13	Van Veen surface sample at station SD-314, replicate F.
2-14	Homogenized sample SD-314-0000 (Rep F) in stainless steel bowl.
2-18	Sediment core from station SD-322- depths as shown.
2-19	Sediment core from station SD-322- depths as shown.
2-20	Observed sheen on ship deck from bottom of sediment core at station SD-313 (rep A).
2-33	Processed sediment core from location SD-314. Photos 2-33 to 2-34 taken from top to bottom of core.
2-34	Processed sediment core from location SD-314. Photos 2-33 to 2-34 taken from top to bottom of core.
2-35	Sample SD-314-0001 homogenized in stainless steel bowl.
2-36	Sample SD-314-0002 homogenized in stainless steel bowl.
2-37	Sample SD-314-0003 homogenized in stainless steel bowl.
4-5	Processed sediment core from location SD-309. Photos 4-5 through 4-7 taken from top to bottom of core.
4-6	Processed sediment core from location SD-309. Photos 4-5 through 4-7 taken from top to bottom of core.
4-7	Processed sediment core from location SD-309. Photos 4-5 through 4-7 taken from top to bottom of core.
4-8	Sample SD-309-0001 homogenized in stainless steel bowl.
4-9	Sample SD-309-0002 homogenized in stainless steel bowl.

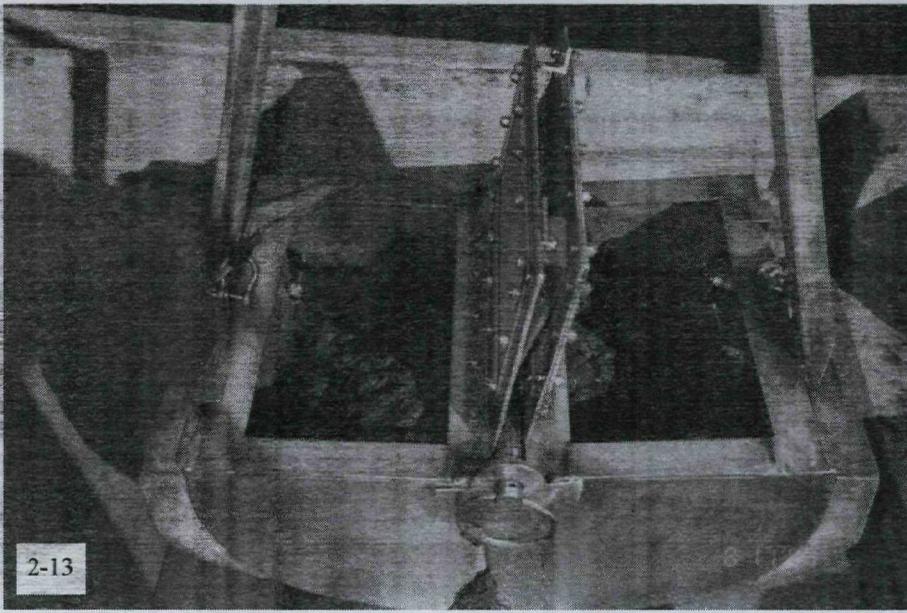
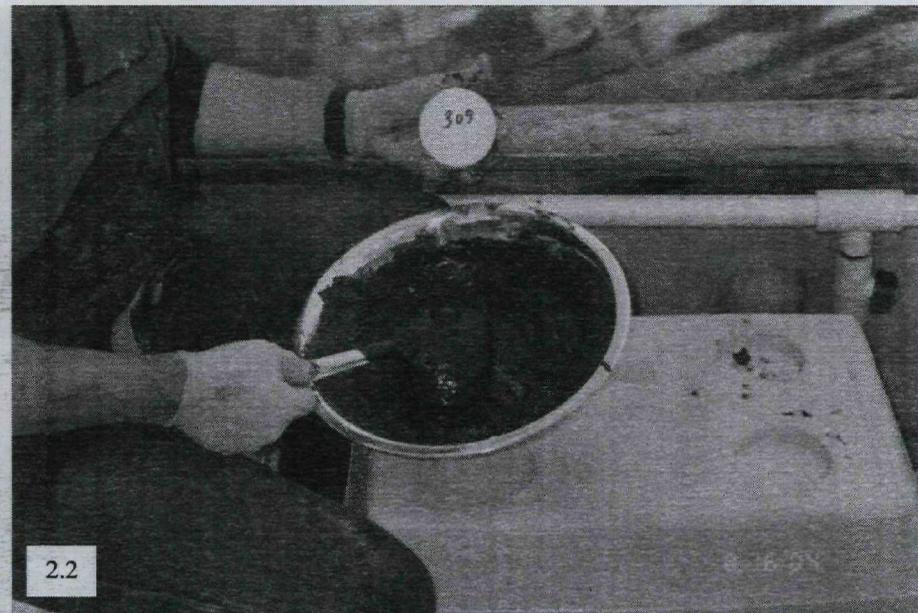
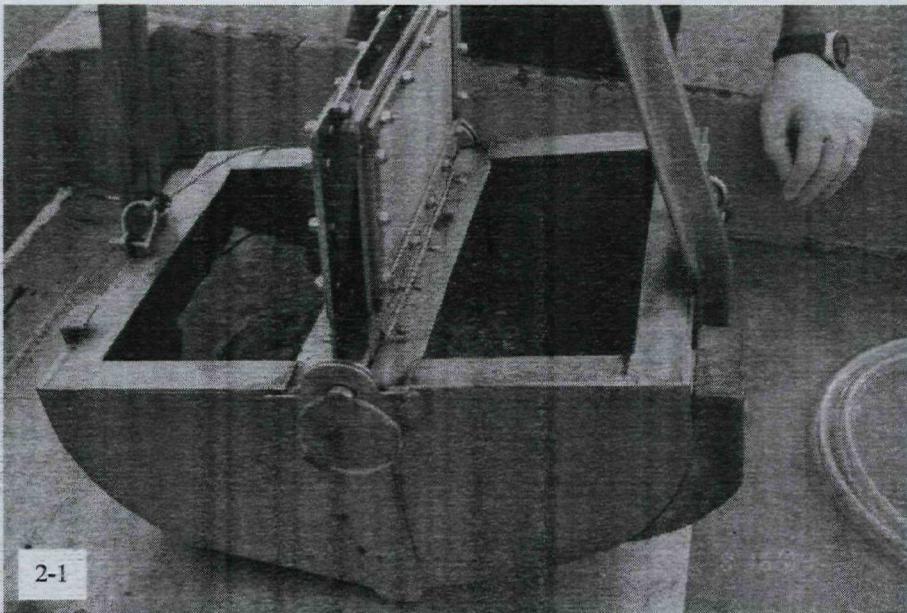
*Lower Duwamish Triad Sampling Event—Technical Report*

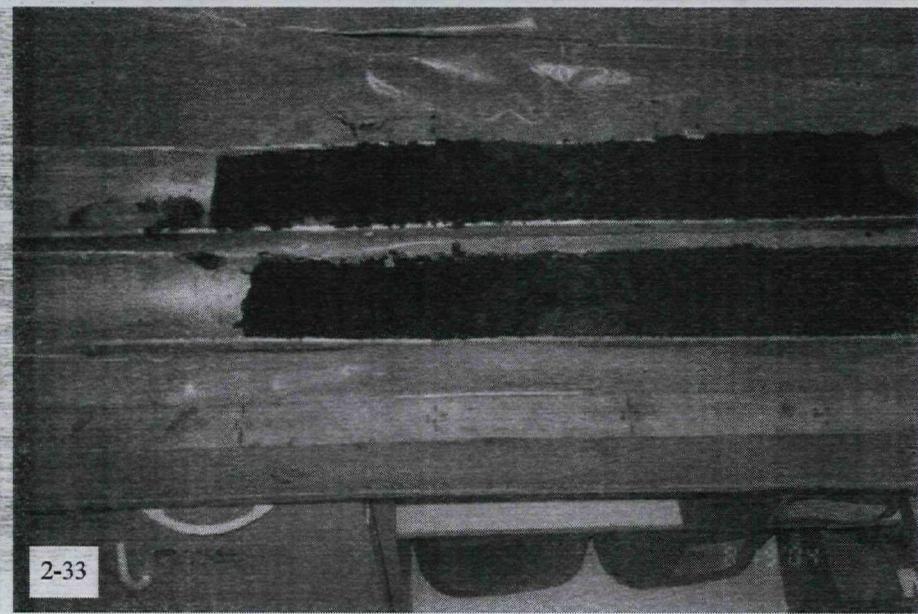
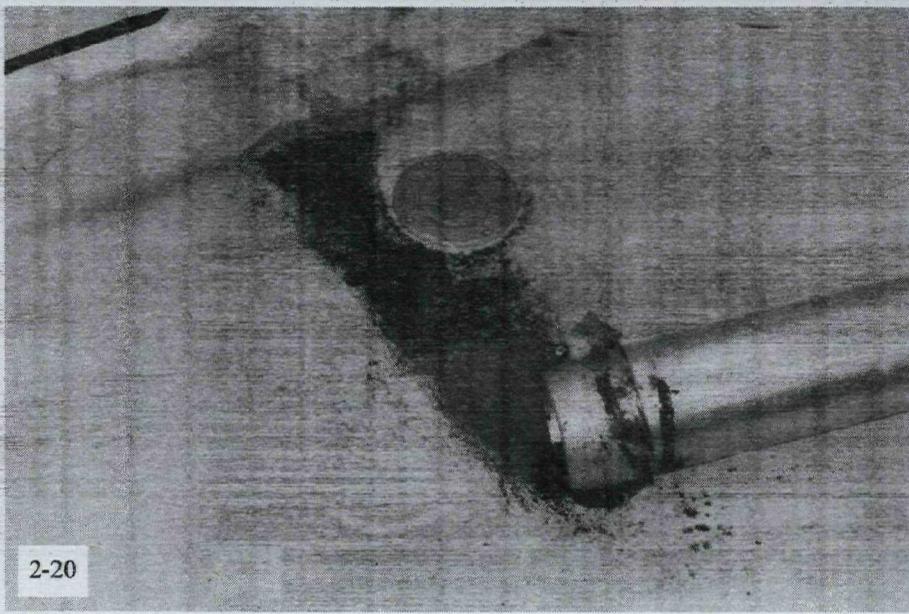
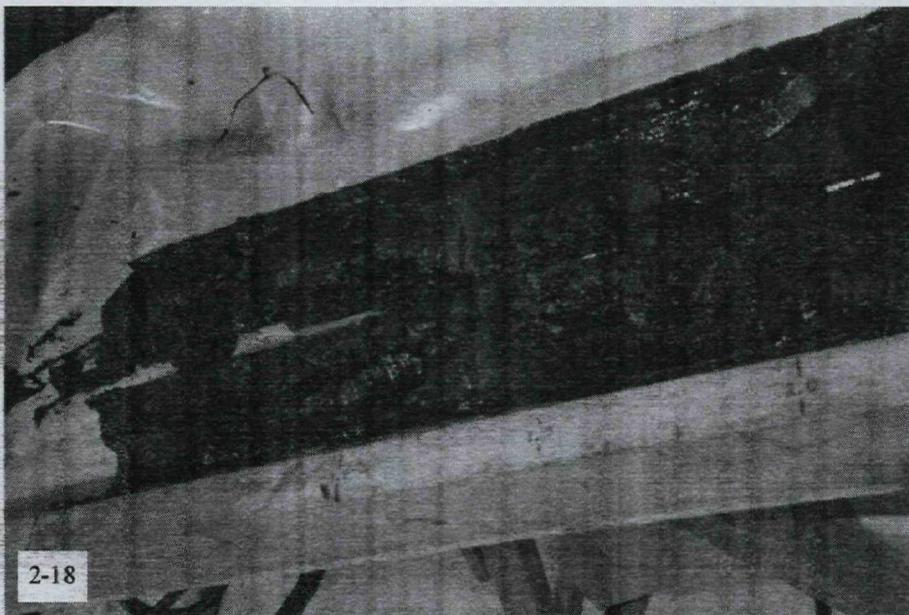
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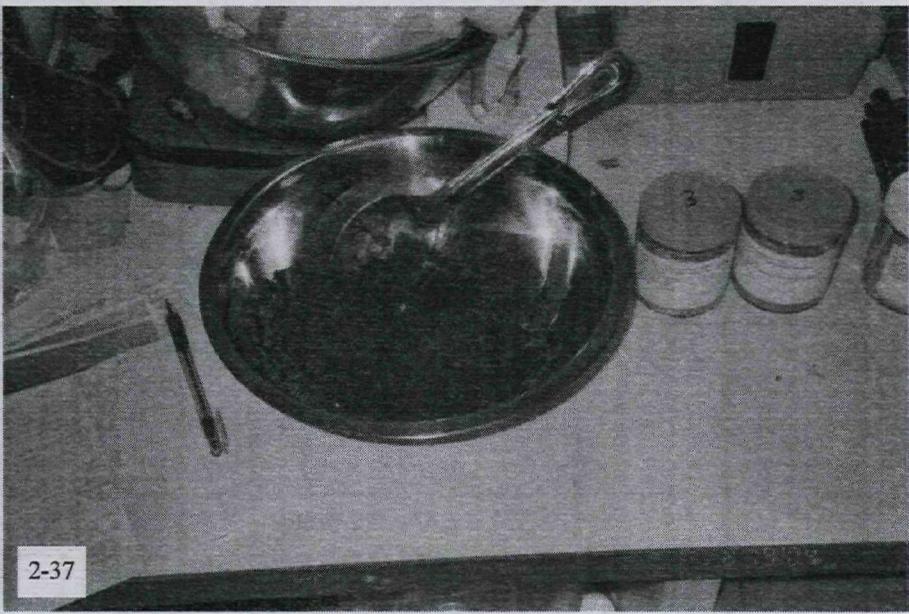
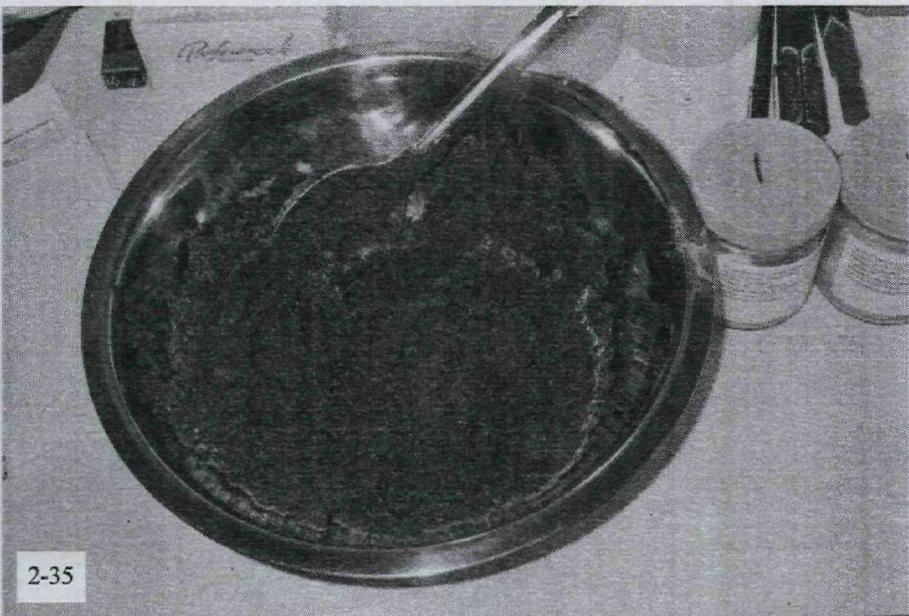
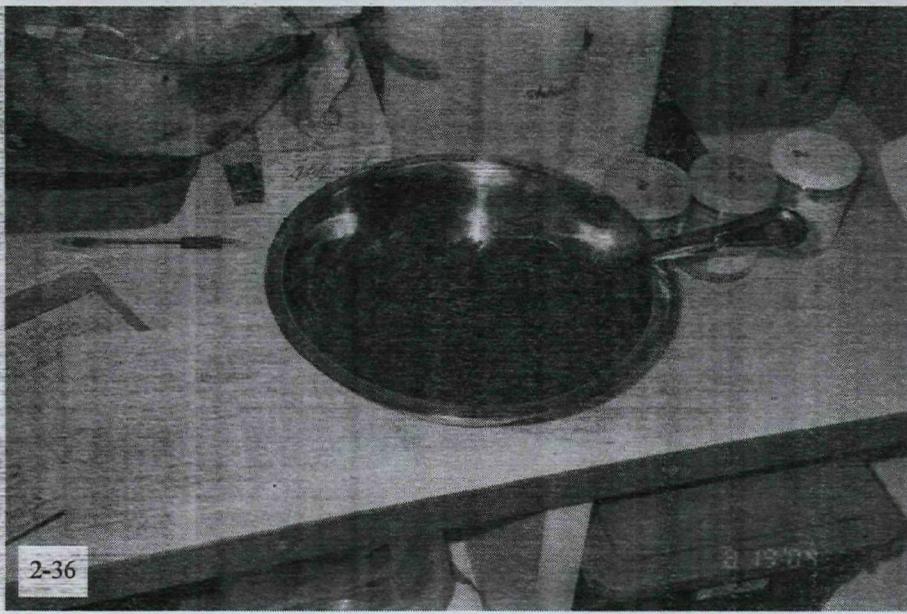
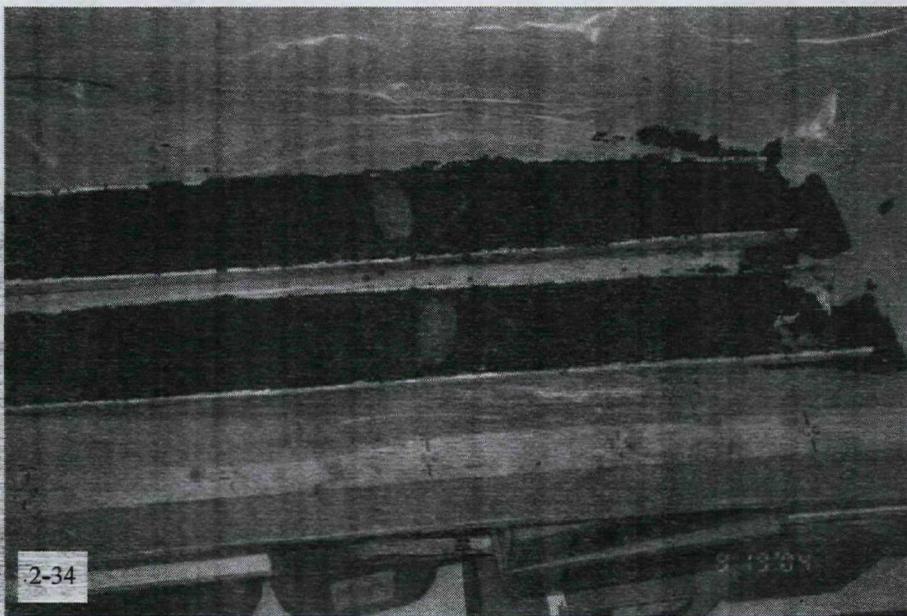
Photo Number	Photo Description
4-10	Sample SD-309-0003 homogenized in stainless steel bowl.
4-15	Processed sediment core from location SD-317. Photos 4-15 through 4-17 taken from top to bottom of core.
4-16	Processed sediment core from location SD-317. Photos 4-15 through 4-17 taken from top to bottom of core.
4-17	Processed sediment core from location SD-317. Photos 4-15 through 4-17 taken from top to bottom of core.
4-18	Sample SD-317-0001 homogenized in stainless steel bowl.
4-19	Sample SD-317-0002 homogenized in stainless steel bowl.
4-20	Sample SD-317-0003 homogenized in stainless steel bowl.
6-3	Van Veen surface sample at station SD-344.
6-4	Homogenized sample SD-344-0000 in stainless steel bowl.
7-6	Van Veen surface sample at station SD-211.
7-7	Homogenized sample SD-211-0000 in stainless steel bowl.

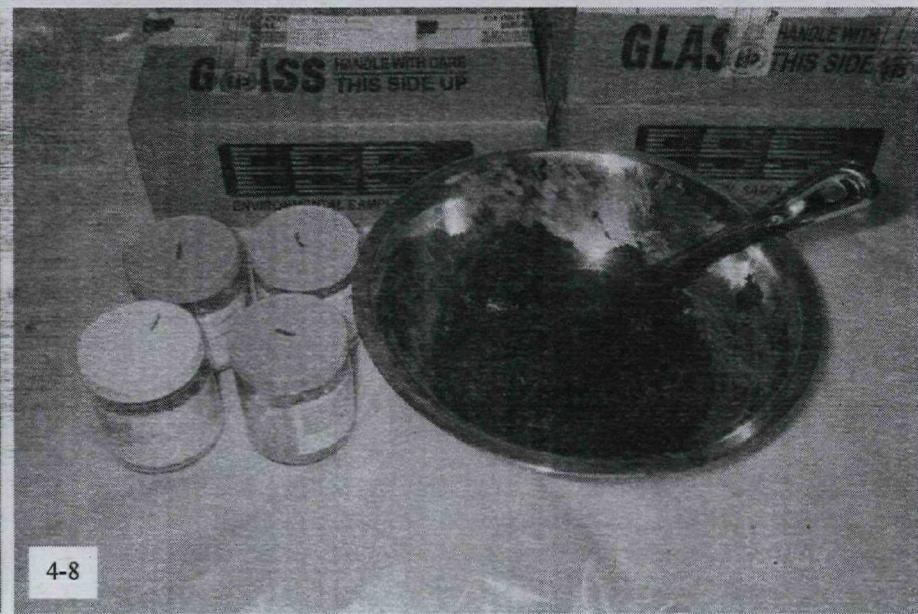
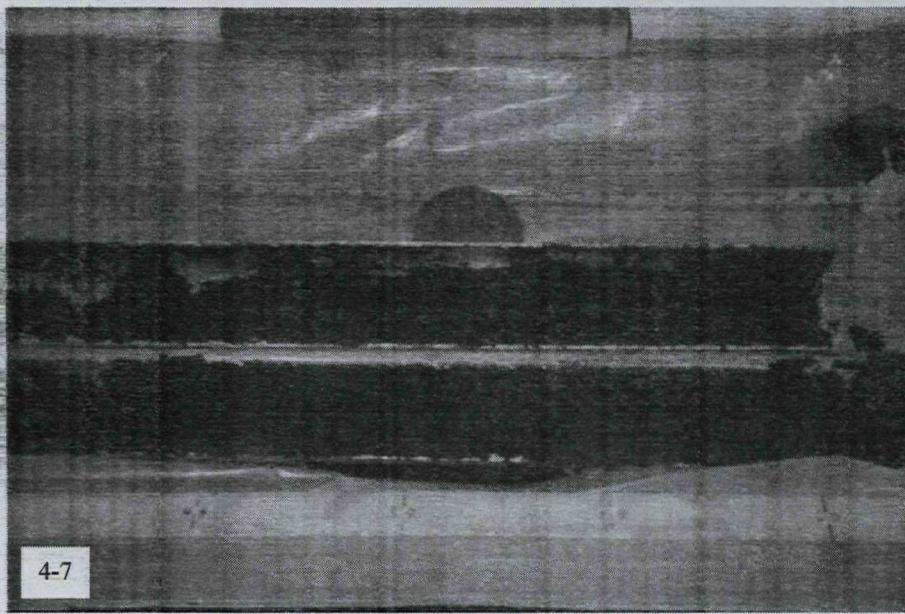
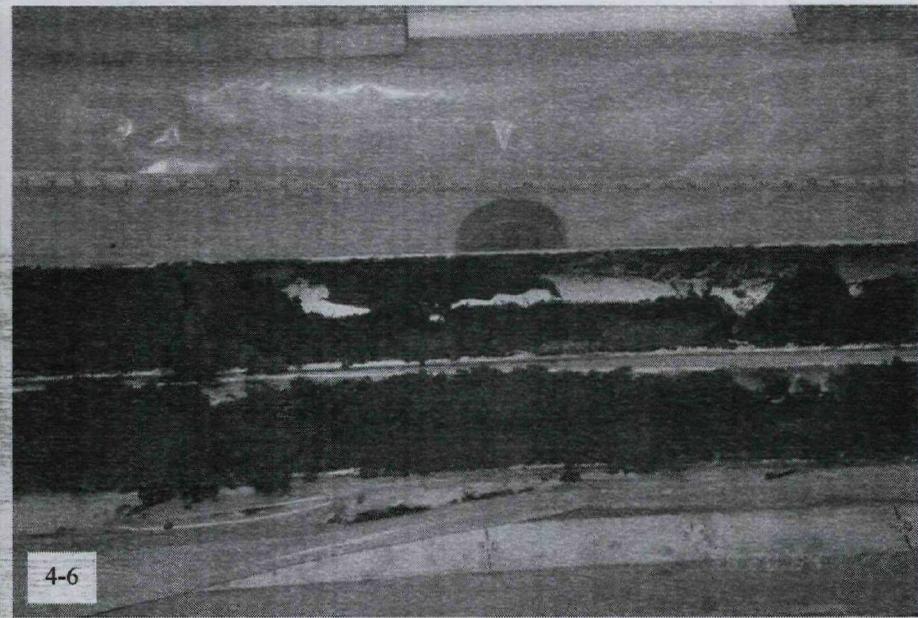
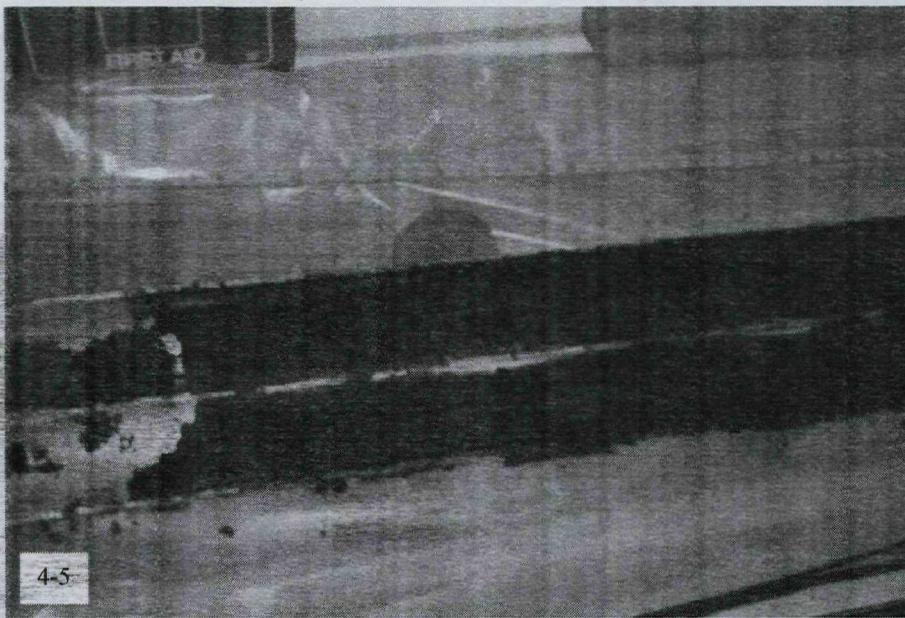


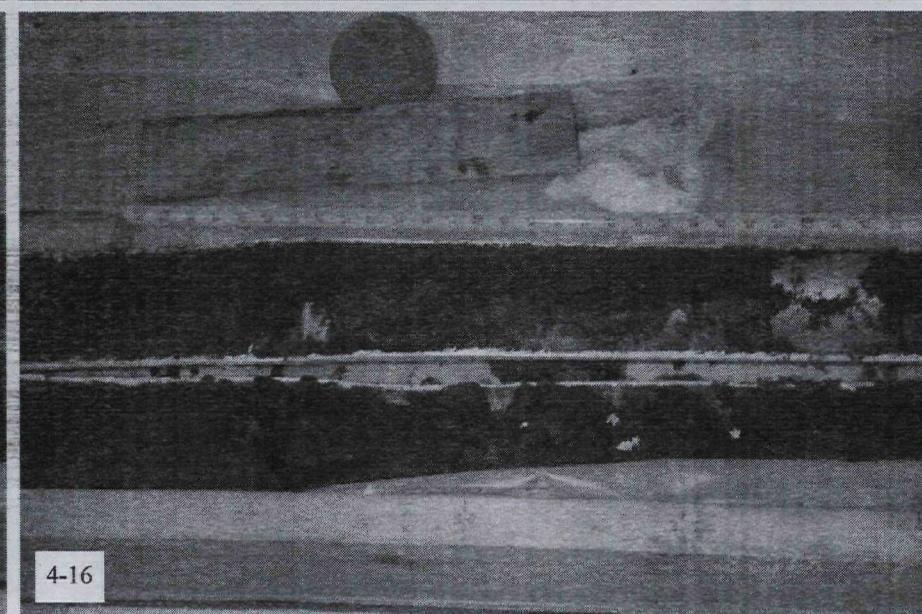
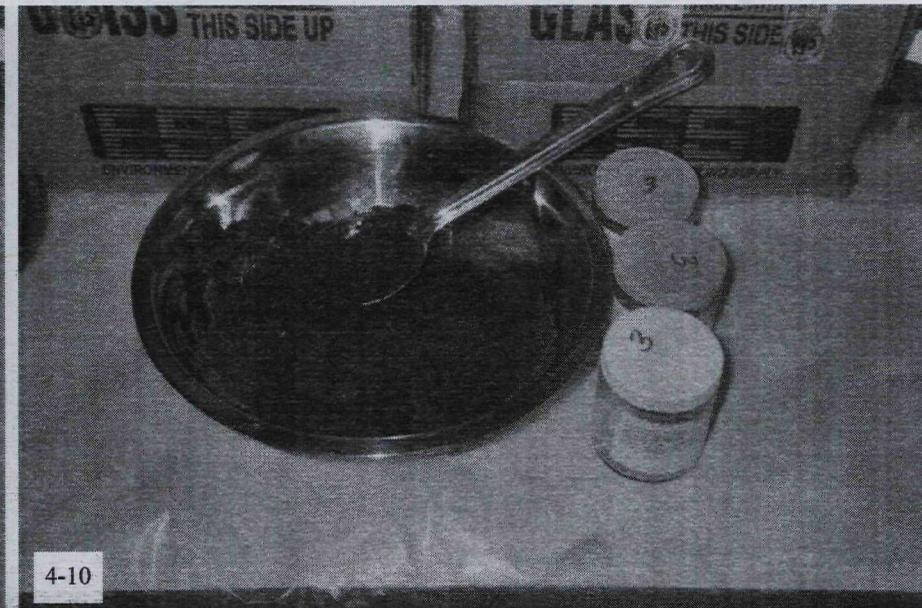
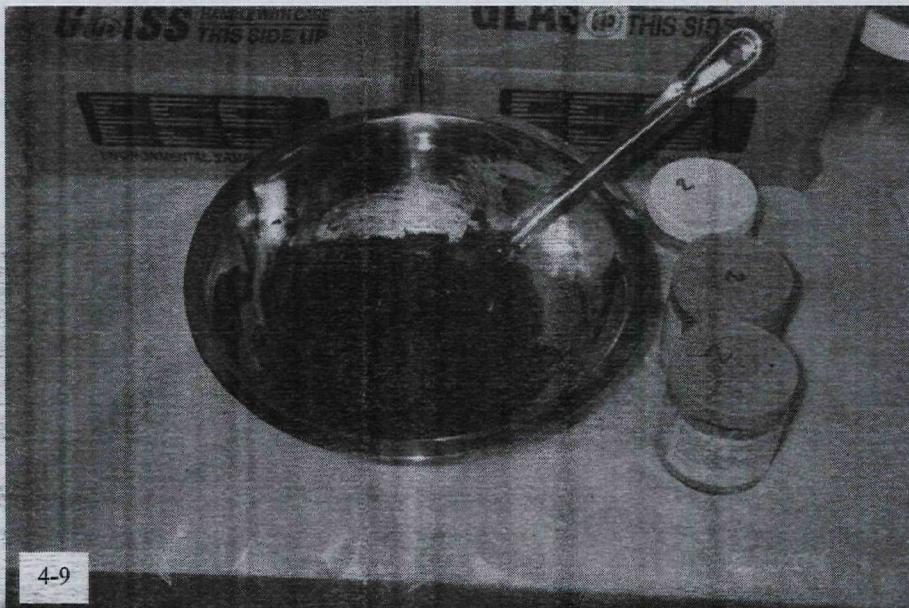


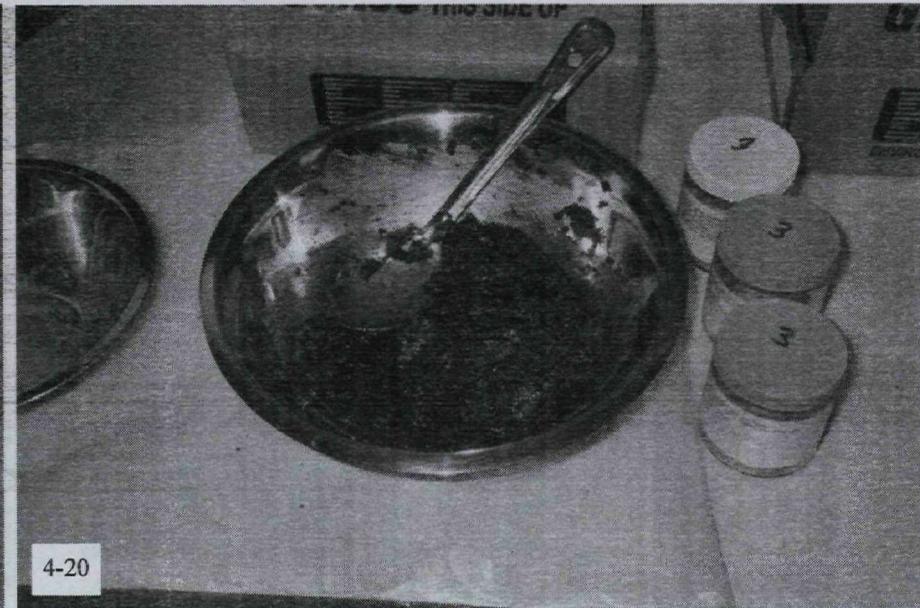
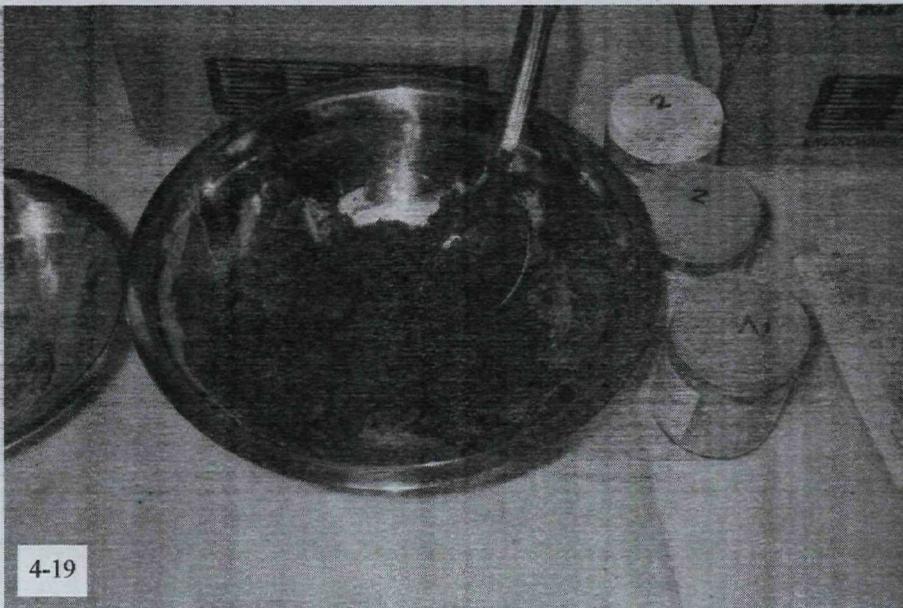
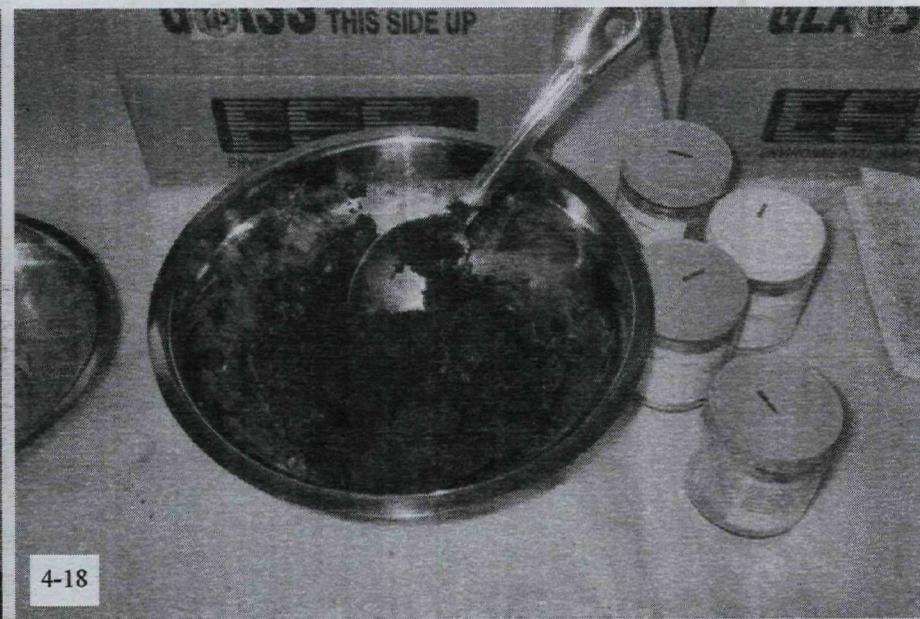
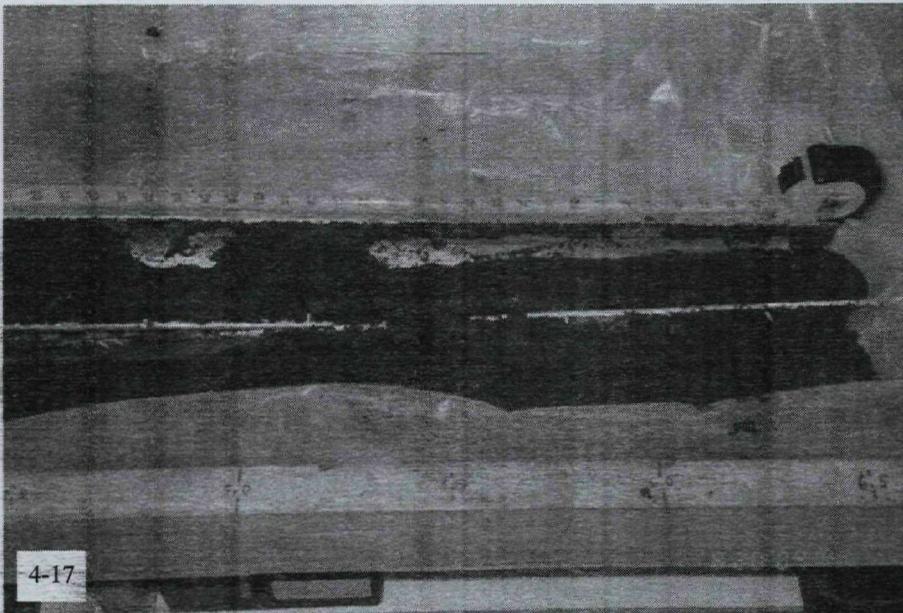


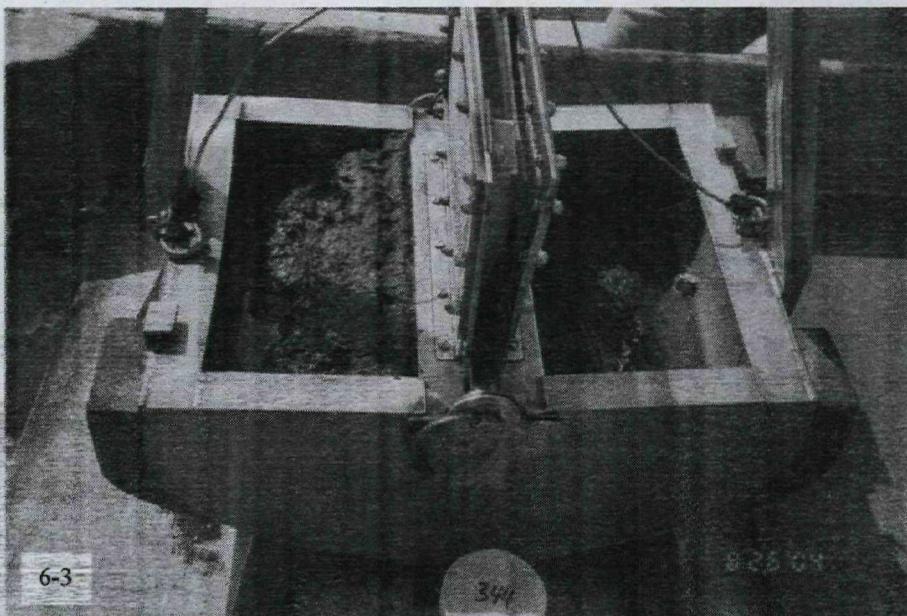




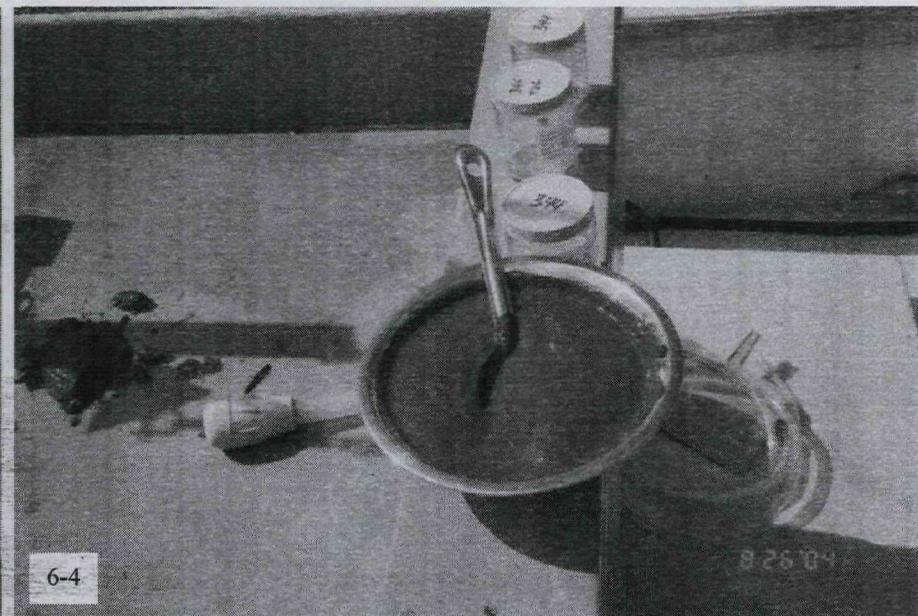




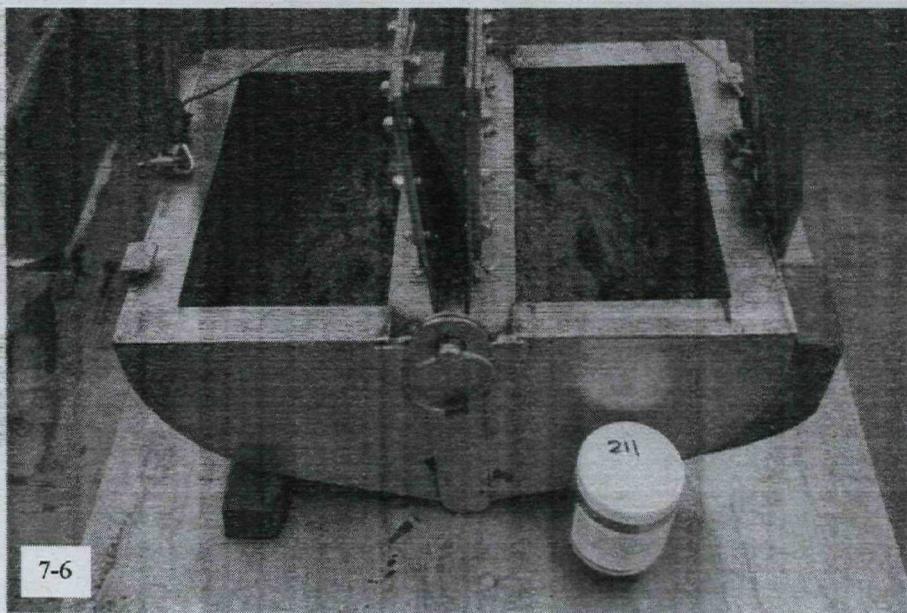




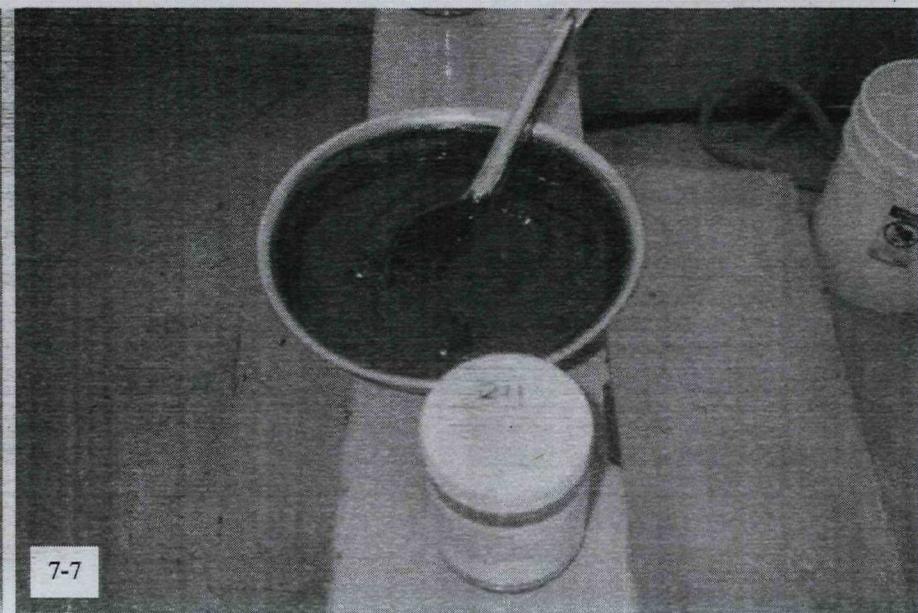
6-3



6-4



7-6



7-7

## **Appendix C: Station Location Coordinates**

**Table C-1. Planned and final field station location coordinates for the Lower Duwamish Triad sampling event.**

Station ID	Station Coordinates <sup>a</sup>					
	Planned Station Location		Final Surface Grab Location		Final Subsurface Core Location	
	Northing	Easting	Northing	Easting	Northing	Easting
SD-DUW-307	195686	1275819	195684	1275819	195673	1275804
SD-DUW-309	195579	1275858	195580	1275858	195576	1275839
SD-DUW-310	195518	1275879	195517	1275880	195514	1275867
SD-DUW-311	195458	1275906	195457	1275909	195460	1275899
SD-DUW-312	195430	1275928	195427	1275912	195427	1275919
SD-DUW-313	195308	1275972	195311	1275946	195309	1275932
SD-DUW-314	195231	1275992	195235	1275968	195237	1275967
SD-DUW-315	195158	1276004	195159	1276003	195159	1275999
SD-DUW-316	195652	1275820	195650	1275820	195647	1275802
SD-DUW-317	195624	1275838	195624	1275837	195632	1275821
SD-DUW-318	195542	1275864	195543	1275863	195550	1275868
SD-DUW-319	195438	1275874	195433	1275871	195435	1275867
SD-DUW-320	195386	1275930	195387	1275927	195382	1275925
SD-DUW-321	195768	1275591	195765	1275593	195768	1275600
SD-DUW-322	195315	1275871	195314	1275870	195311	1275864
SD-DUW-323	195355	1275948	195348	1275946	195347	1275938
SD-DUW-206	195606	1275811	195606	1275814	NA	NA
SD-DUW-207	195555	1275827	195554	1275827	NA	NA
SD-DUW-208	195511	1275851	195512	1275852	NA	NA
SD-DUW-209	195418	1275852	195410	1275860	NA	NA
SD-DUW-210	195366	1275875	195365	1275875	NA	NA
SD-DUW-211	195322	1275905	195319	1275903	NA	NA
SD-DUW-212	195278	1275903	195278	1275905	NA	NA
SD-DUW-213	195577	1275816	195579	1275818	NA	NA
SD-DUW-214	195231	1275902	195230	1275903	NA	NA
SD-DUW-215	195467	1275849	195464	1275851	NA	NA
SD-DUW-216	195184	1275932	195185	1275934	NA	NA
SD-DUW-217	195130	1275950	195131	1275949	NA	NA
SD-DUW-330	195628	1275810	195629	1275811	NA	NA
SD-DUW-331	195610	1275830	195608	1275828	NA	NA
SD-DUW-332	195567	1275853	195566	1275858	NA	NA
SD-DUW-333	195596	1275856	195589	1275849	NA	NA
SD-DUW-334	195473	1275893	195473	1275890	NA	NA
SD-DUW-335	195444	1275852	195446	1275854	NA	NA
SD-DUW-336	194948	1276033	194947	1276034	NA	NA

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*Lower Duwamish Triad Sampling Event—Technical Report***Table C-1 (continued). Planned and final field station location coordinates for the Lower Duwamish Triad sampling event.**

Station ID	Station Coordinates <sup>a</sup>				Final Subsurface Core Location	
	Planned Station Location		Final Surface Grab Location			
	Northing	Easting	Northing	Easting	Northing	Easting
SD-DUW-337	195270	1275948	195270	1275948	NA	NA
SD-DUW-338	195297	1275991	195306	1275944	NA	NA
SD-DUW-339	195210	1276004	195214	1275963	NA	NA
SD-DUW-340	195383	1275907	195381	1275908	NA	NA
SD-DUW-341	195672	1275843	195676	1275827	NA	NA
SD-DUW-342	195406	1275922	195406	1275922	NA	NA
SD-DUW-343	195528	1275875	195527	1275875	NA	NA
SD-DUW-344	195705	1275820	195707	1275799	NA	NA
SD-DUW-345	195135	1275986	195135	1275985	NA	NA

<sup>a</sup> Coordinates are North American Datum (NAD) of 1983 State Plane Washington North.  
NA Not applicable.

Lower Duwamish Triad Sampling Event—Technical Report

Station ID	Planned Station Location Northing	Planned Station Location Easting	Final Surface Grab Location Northing	Final Surface Grab Location Easting	Final Subsurface Core Location Northing	Final Subsurface Core Location Easting
SD-DUW-337	195270	1275948	195270	1275948	NA	NA
SD-DUW-338	195297	1275991	195306	1275944	NA	NA
SD-DUW-339	195210	1276004	195214	1275963	NA	NA
SD-DUW-340	195383	1275907	195381	1275908	NA	NA
SD-DUW-341	195672	1275843	195676	1275827	NA	NA
SD-DUW-342	195406	1275922	195406	1275922	NA	NA
SD-DUW-343	195528	1275875	195527	1275875	NA	NA
SD-DUW-344	195705	1275820	195707	1275799	NA	NA
SD-DUW-345	195135	1275986	195135	1275985	NA	NA

Coordinates are North American Datum (NAD) of 1983 State Plane Washington North.

## **Appendix D: Samples Submitted for Laboratory Analysis**

*See Appendix D for a listing of all samples submitted for laboratory analysis.*

**Table D-1. Phase 2 sediment samples collected and analyzed for the Lower Duwamish Triad sampling event.**

Sample ID	Regional Sample ID	SPAWAR	Analytical Resources, Inc					EPA Region 10 Environmental Laboratory			
		PCBs (4020)	TOC	GS	Hg	SVOC	TCLP Pb	PCBs (8082)	Metals	SVOC/P CB <sup>a</sup>	SVOC
SD-307-0000	04344100	X	X	X	O	O			X	X	
SD-307-0001	04344101	X	X					X	X		
SD-307-0002	04344102	X	X					X			
SD-307-0003	04344103	X	X					X			
SD-309-0000	04344104	X	X	X	O	O			X	X	
SD-309-0001	04344105	X	X					X	X		
SD-309-0002	04344106	X	X					X			
SD-309-0003	04344107	X	X					X			
SD-310-0000	04344108	X	X	X	O	O			X	X	
SD-310-0001	04344109	X	X					X	X		
SD-310-0002	04344110	X	X					X			
SD-310-0003	04344111	X	X					X			
SD-311-0000	04344112	X	X	X	O	O			X	X	
SD-311-0001	04344113	X	X					X	X		
SD-311-0002	04344114	X	X					X			
SD-311-0003	04344115	X	X					X			
SD-312-0000	04344116	X	X	X	O	O			X	X	
SD-312-0001	04344117	X	X					X	X		
SD-312-0002	04344118	X	X					X			
SD-312-0003	04344119	X	X					X			
SD-313-0000	04344120	X	X	X	O	O			X	X	
SD-313-0001	04344121	X	X		O	O		X	X		
SD-313A-0001 <sup>b</sup>	04344174									X	
SD-313-0002	04344122	X	X					X			
SD-313-0003	04344123	X	X					X			
SD-314-0000	04344124	X	X	X	O	O			X	X	
SD-314-0001	04344125	X	X					X	X		
SD-314-0002	04344126	X	X					X			
SD-314-0003	04344127	X	X					X			
SD-315-0000	04344128	X	X	X	O	O			X	X	
SD-315-0001	04344129	X	X					X	X		
SD-315-0002	04344130	X	X					X			
SD-315-0003	04344131	X	X					X			
SD-316-0000	04344132	X	X	X	O	O			X	X	
SD-316-0001	04344133	X	X					X	X		
SD-316-0002	04344134	X	X					X			
SD-316-0003	04344135	X	X					X			
SD-317-0000	04344136	X	X	X	O	O			X	X	
SD-317-0001	04344137	X	X					X	X		
SD-317-0002	04344138	X	X					X			
SD-317-0003	04344139	X	X					X			
SD-318-0000	04344140	X	X	X	O	O			X	X	
SD-318-0001	04344141	X	X					X	X		
SD-319-0000	04344144	X	X	X	O	O			X	X	
SD-319-0001	04344145	X	X					X	X		
SD-319-0002	04344146	X	X					X			

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June 6, 2008

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*Lower Duwamish Triad Sampling Event—Technical Report*

**Table D-1 (continued): Phase 2 sediment samples collected and analyzed for the Lower Duwamish Triad sampling event.**

Sample ID	Regional Sample ID	SPAWAR Analytical Resources, Inc					EPA Region 10 Environmental Laboratory			
		PCBs (4020)	TOC	GS	Hg	SVOC	TCLP Pb	PCBs (8082)	Metals	SVOC/PCB <sup>a</sup>
SD-319-0003	04344147	X	X					X		
SD-320-0000	04344148	X	X	X	O	O			X	X
SD-320-0001	04344149	X	X					X	X	
SD-320-0002	04344150	X	X					X		
SD-320-0003	04344151	X	X					X		
SD-321-0000	04344152	X	X	X	O	O			X	X
SD-321-0001	04344153	X	X					X	X	
SD-321-0002	04344154	X	X					X		
SD-321-0003	04344155	X	X					X		
SD-322-0000	04344158	X	X	X	O	O			X	X
SD-322-0001	04344159	X	X					X	X	
SD-322-0002	04344160	X	X					X		
SD-322-0003	04344161	X	X					X		
SD-322-0004	04344162	X	X					X		
SD-322-0005	04344163	X	X					X		
SD-323-0000	04344164	X	X	X	O	O			X	X
SD-323-0001	04344165	X	X					X	X	
SD-323-0002	04344166	X	X					X		
SD-323-0003	04344167	X	X					X		
SD-324-0000 <sup>c</sup>	04344168	X	X	X	O	O			X	X
SD-325-0000 <sup>c</sup>	04344169	X	X		O	O			X	X
SD-326-0001 <sup>c</sup>	04344170	X	X					X		
SD-327-0002 <sup>c</sup>	04344171	X						X		
SD-328-0003 <sup>c</sup>	04344172	X	X							
SD-434 <sup>d</sup>	04344173							X	X	
WD01 <sup>e</sup>	NA						X			X

O      Sample analysis added after sampling event was complete. Analysis performed using sample jar submitted for TOC.

PCBs (4020) polychlorinated biphenyls (PCBs) by EPA method 4020 (immunoassay).

TOC Total organic carbon by modified EPA method 9060.

GS Grain size by Puget Sound Estuary Program (PSEP) protocols.

Hg Mercury by EPA method 7471A.

PCBs (8082) PCBs by EPA method 8082.

SVOCs Semivolatile organic compounds by EPA method 8260B.

TCLP Pb Lead by the toxicity characteristic leaching procedure (TCLP) (EPA methods 1311 and 6010B).

Metals Metals by EPA method 6010B.

<sup>a</sup> SVOC/PCB = Analyzed for PCBs by EPA method 8082 and then archived (frozen) for SVOC analysis at a later date, if needed.

<sup>b</sup> Sample SD-313A-0001 collected from a rejected core attempt at location SD-321 because a sheen was observed.

<sup>c</sup> Field duplicate of primary sample.

<sup>d</sup> Sample SD-434 is a rinsate sample.

<sup>e</sup> Sample WD01 collected from investigation derived waste (IDW) buckets to determine disposal requirement.

Shaded cells indicate that sample was submitted as a "contingency" sample; sample jars were archived (frozen) upon laboratory receipt (samples may be analyzed at a future date based on project decisions).

NA Not applicable

SPAWAR U.S. Navy Space and Naval Warfare Systems Center laboratory in San Diego, California.

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**Table D-2. Phase 3 sediment samples collected and analyzed for the Lower Duwamish Triad sampling event.**

Sample ID	Regional Sample ID	SPAWAR	Analytical Resources, Inc				EPA Region X Environmental Laboratory	
		PCBs (4020)	TOC	GS	Hg	TCLP Pb	PCBs (8082)	Metals
SD-206-0000	04354000	X	X				X	
SD-207-0000	04354001	X	X				X	
SD-208-0000	04354002	X	X				X	
SD-209-0000	04354003	X	X				X	
SD-210-0000	04354004	X	X				X	
SD-211-0000	04354005	X	X				X	
SD-212-0000	04354006	X	X				X	
SD-213-0000	04354007	X	X				X	
SD-214-0000	04354008	X	X				X	
SD-215-0000	04354009	X	X				X	
SD-216-0000	04354010	X	X				X	
SD-217-0000	04354011	X	X				X	
SD-330-0000	04354012	X	X				X	
SD-331-0000	04354013	X	X				X	
SD-332-0000	04354014	X	X				X	
SD-333-0000	04354015	X	X	X	O		X	X
SD-334-0000	04354016	X	X				X	
SD-335-0000	04354017	X	X				X	
SD-336-0000	04354018	X	X				X	
SD-337-0000	04354019	X	X	X			X	X
SD-338-0000	04354020	X	X				X	
SD-339-0000	04354021	X	X				X	
SD-340-0000	04354022	X	X				X	
SD-341-0000	04354023	X	X		O		X	
SD-342-0000	04354024	X	X	X			X	X
SD-343-0000	04354025	X	X				X	
SD-344-0000	04354026	X	X				X	
SD-345-0000	04354027	X	X				X	
SD-431 <sup>a</sup>	04354031	X	X	X			X	X
SD-432 <sup>a</sup>	04354032	X	X				X	
SD-433 <sup>a</sup>	04354033	X	X				X	
WD02 <sup>b</sup>	NA					X		

O Sample analysis added after sampling event was complete. Analysis performed using sample jar submitted for TOC.

PCBs (4020) Polychlorinated biphenyls (PCBs) by immunoassay.

TOC Total organic carbon by EPA method 9060.

GS Grain size by PSEP protocols.

Hg Mercury by EPA method 7471A.

TCLP Pb Lead by the toxicity characteristic leaching procedure (TCLP) (EPA methods 1311 and 6010B).

PCBs (8082) PCBs by EPA method 8082.

<sup>a</sup> Field duplicate of primary sample.

<sup>b</sup> Sample WD02 collected from investigation derived waste (IDW) buckets to determine disposal requirements. Shaded cells indicate that sample was submitted as a "contingency" sample; sample jars were archived (frozen) upon laboratory receipt (samples may be analyzed at a future date based on project decisions).

NA Not applicable.

SPAWAR U.S. Navy Space and Naval Warfare Systems Center laboratory in San Diego, California.

## **Appendix E: Analytical Results from ARI**

**Table E-1. Phase 2 total organic carbon analytical results for the Lower Duwamish Triad sampling event.**

Sample ID	TOC (percent)
SD-307-0000	2.20
SD-307-0001	0.800
SD-307-0002	0.226
SD-307-0003	0.317
SD-309-0000	2.21
SD-309-0001	1.77
SD-309-0002	1.32
SD-309-0003	0.941
SD-310-0000	1.98
SD-310-0001	1.54
SD-310-0002	1.50
SD-310-0003	0.986
SD-311-0000	1.84
SD-311-0001	1.06
SD-311-0002	1.45
SD-311-0003	0.725
SD-312-0000	2.58
SD-312-0001	1.09
SD-312-0002	0.134
SD-312-0003	0.130
SD-313-0000	2.22
SD-313-0001	1.81
SD-313-0002	0.947
SD-313-0003	1.18
SD-314-0000	1.66
SD-314-0001	2.44
SD-314-0002	0.237
SD-314-0003	0.428
SD-315-0000	1.57
SD-315-0001	0.218
SD-315-0002	0.305
SD-315-0003	0.183
SD-316-0000	2.25
SD-316-0001	1.66
SD-316-0002	1.87
SD-316-0003	1.53
SD-317-0000	2.58
SD-317-0001	2.00
SD-317-0002	1.14
SD-317-0003	1.76

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**Table E-1 (continued). Phase 2 total organic carbon analytical results for the Lower Duwamish Triad sampling event.**

Sample ID	TOC (percent)
SD-317-0002	2.11
SD-317-0003	0.816
SD-318-0000	2.05
SD-318-0001	0.824
SD-319-0000	1.74
SD-319-0001	1.80
SD-319-0002	2.22
SD-319-0003	1.81
SD-320-0000	1.62
SD-320-0001	2.24
SD-320-0002	2.39
SD-320-0003	1.83
SD-321-0000	2.25
SD-321-0001	1.69
SD-321-0002	1.96
SD-321-0003	1.74
SD-322-0000	2.02
SD-322-0001	1.90
SD-322-0002	2.63
SD-322-0003	2.72
SD-322-0004	1.44
SD-322-0005	0.852
SD-323-0000	2.49
SD-323-0001	1.65
SD-323-0002	2.21
SD-323-0003	1.31
SD-324-0000 <sup>a</sup>	2.23
SD-325-0000 <sup>a</sup>	1.73
SD-326-0001 <sup>a</sup>	1.58
SD-328-0003 <sup>a</sup>	1.72

TOC total organic carbon analyzed by EPA method 9060.  
field duplicate samples.

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**Table E-2. Phase 3 total organic carbon analytical results for the Lower Duwamish Triad sampling event.**

Sample ID	TOC (percent)
SD-206-0000	2.34
SD-207-0000	2.11
SD-208-0000	1.46
SD-209-0000	2.71
SD-210-0000	2.65
SD-211-0000	2.16
SD-212-0000	2.33
SD-213-0000	2.17
SD-214-0000	2.78
SD-215-0000	1.64
SD-216-0000	2.02
SD-217-0000	1.84
SD-330-0000	2.15
SD-331-0000	2.43
SD-332-0000	2.24
SD-333-0000	2.50
SD-334-0000	1.55
SD-335-0000	1.98
SD-336-0000	1.55
SD-337-0000	1.92
SD-338-0000	1.81
SD-339-0000	1.7
SD-340-0000	1.61
SD-341-0000	2.68
SD-342-0000	1.45
SD-343-0000	1.96
SD-344-0000	2.58
SD-345-0000	1.24
SD-431 <sup>a</sup>	2.24
SD-432 <sup>a</sup>	1.92
SD-433 <sup>a</sup>	1.56

TOC = total organic carbon analyzed by EPA method 9060.  
field duplicate samples.

Table E-3. Semivolatile organic compound sediment analytical results for the Lower Duwamish Triad sampling event.

Compound	Sample results ( $\mu\text{g/kg}$ )																		
	SD-307-0000	SD-309-0000	SD-310-0000	SD-311-0000	SD-312-0000	SD-313-0000	SD-313-0001	SD-314-0000	SD-315-0000	SD-316-0000	SD-317-0000	SD-318-0000	SD-319-0000	SD-320-0000	SD-321-0000	SD-322-0000	SD-323-0000	SD-324-0000 <sup>a</sup>	SD-325-0000 <sup>a</sup>
Phenol	440	290	270	1,200	610	620	20 U	160 J	28	310	1,100	800	32	410	130	290	840	140	560 J
Bis-(2-chloroethyl) ether	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2-Chlorophenol	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
1,3-Dichlorobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
1,4-Dichlorobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Benzyl alcohol	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
1,2-Dichlorobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2-Methylphenol	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2,2'-Oxybis (1-Chloropropane)	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
4-Methylphenol	52	19 J	20 U	27	22	22	20 U	21	20 U	19 U	34	28	20 U	83	20 U				
N-Nitroso-di-N-propylamine	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Hexachloroethane	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Nitrobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Isopropene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2-Nitrophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2,4-Dimethylphenol	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Benzoic acid	200 U	200 U	200 U	200 U	200 U	200 U	200 U	200 U	190 U	200 U	190 U	200 U	200 U						
Bis (2-chloroethoxy) methane	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2,4-Dichlorophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
1,2,4-Trichlorobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Naphthalene	20 U	490	31	62	170	220	20 U	28 J	20 U	19 U	20 U	46	20 U	24	20 U	20 U	22	20 U	160 J
4-Chloroaniline	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Hexachlorobutadiene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
4-Chloro-3-methylphenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2-Methylnaphthalene	27 U	350	34	60	148	190	20 U	27 J	20 U	19 U	20 U	37	20 U	24	20 U	20 U	22	20 U	100 J
Hexachlorocyclopentadiene	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2,4,6-Trichlorophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2,4,5-Trichlorophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2-Chloronaphthalene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
2-Nitroaniline	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Dimethylphthalate	19 J	24	29	26	22	20 U	20 U	20 U	35	40	29	23	20 U	74	20 U				
Acenaphthylene	57	98	42	35	98	20 U	20 U	20 U	23	19 U	20 U	28	20 U	22					
3-Nitroaniline	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Acenaphthene	26	220	56	70	250	200	20 U	42 J	44	19 U	20	52	20 U	35	20 U				
2,4-Dinitrophenol	200 U	200 U	200 U	200 U	200 U	200 U	200 U	200 U	190 U	200 U	190 U	200 U	200 U						
4-Nitrophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U

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Table E-3 (continued). Semivolatile organic compound sediment analytical results for the Lower Duwamish Triad sampling event.

Compound	Sample results ( $\mu\text{g}/\text{kg}$ )																		
	SD-307-0000	SD-309-0000	SD-310-0000	SD-311-0000	SD-312-0000	SD-313-0000	SD-313-0001	SD-314-0000	SD-315-0000	SD-316-0000	SD-317-0000	SD-318-0000	SD-319-0000	SD-320-0000	SD-321-0000	SD-322-0000	SD-323-0000	SD-324-0000 <sup>a</sup>	SD-325-0000 <sup>a</sup>
Dibenzofuran	180	410	82	100	460	55	20 U	25	31	19 U	20	75	20 U	28	20 U	20 U	21	20 U	54
2,6-Dinitrotoluene	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
2,4-Dinitrotoluene	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Diethylphthalate	20 U	20 U	20	20 U	91	20 U	20 U	28	20 U	19 U	120	94	20 U	48	20 U	150	20 U	27	20 U
4-Chlorophenyl-phenylether	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Fluorene	1,500	810	170	210	940	240	20 U	51 J	51	22	38	150	20 U	48	20 U	20 U	42	20 U	220 J
4-Nitroaniline	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
4,6-Dinitro-2-methylphenol	200 U	200 U	200 U	200 U	200 U	200 U	200 U	200 U	190 U	200 U	190 U	200 U	200 U						
N-Nitrosodiphenylamine	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
4-Bromophenyl-phenylether	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Hexachlorobenzene	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U						
Pentachlorophenol	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Phenanthrene	4,900	3,400	960	1,100	3,800	1,400	39	400 J	680	220	280	840	89	250	81	90	420	68	1,400 J
Carbazole	290	360	110	140	230	75	20 U	69	150	43	56	100	20 U	27	20 U	20	64	20 U	67
Anthracene	4,400	620	220	220	680	360	20 U	130 J	130	45	64	160	20 U	48	30	28	82	25	330 J
Di-n-Butylphthalate	65	20	19 J	20 U	23	34	20 U	30	20 U	21	47	21	28	37	20 U	20 U	30	20 U	42
Fluoranthene	10,000	3,300	1,500	2,000 <sup>b</sup>	3,400	1,000	77	600 J	2,300	530	760	1,400	150	380	270	300	880	230	1,100 J
Pyrene	3,900	2,200	1,000	1,400	2,200	1,100	49	570 J	1,800	450	570	950	190	290	240	180	830	210	1,500 J
Butylbenzylphthalate	33	34	41	40	36	43	20 U	20 U	140	64	100	42	20 U	38	34	31	100	27	26
3,3'-Dichlorobenzidine	98 U	99 U	98 U	98 U	99 U	98 U	99 U	98 U	100 U	97 U	98 U	97 U	99 U	97 U	99 U	100 U	98 U	98 U	98 U
Benzo (a) anthracene	4,700	1,100	580	700 <sup>b</sup>	1,100	440	20 U	220 J	660	200	240	490	60	120	98	98	340	81	500 J
Bis (2-ethylhexyl) phthalate	140	220	270	380	220	100	20 U	150	610	290	360	290	94	110	240	230	410	200	140
Chrysene	6,600	1,200	730	840	1,200	490	22	290 J	950	280	420	630	75	160	140	150	450	110	550 J
Di-n-Octyl phthalate	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	20 U	19 U	20 U	19 U	20 U	20 U					
Benzo (b) fluoranthene	4,500	1,200	760	1,100	1,100	370	20 U	260	1,100	340	430	640	83	130	180	160	550	150	430
Benzo (k) fluoranthene	2,500	830	690	760	840	360	20 U	210 J	950	270	280	440	90	150	140	130	460	130	420 J
Benzo (a) pyrene	2,000	1,000	620	770	970	430	20 U	230 J	780	230	260	510	69	130	110	110	400	97	510 J
Indeno (1,2,3-cd) pyrene	440	410	270	350	330	130	20 U	110 J	390	120	120	270	30	58	52	60	200	46	170 J
Dibenzo (a,h) anthracene	230	170	110	130	100	63	20 U	32	150	45	28	100	20 U	20 U	21	77	20 U	72	
Benzo (g,h,i) perylene	270	280	210	250	120	20 U	88 J	300	97	90	190	24	42	44	45	150	39	150 J	

<sup>a</sup>  $\mu\text{g}/\text{kg}$  – microgram per kilogram.

field duplicate sample.

<sup>b</sup> Bold type indicates the sample result is greater than the reporting limit.

Values reported on a dry-weight basis.

U – The material was analyzed for, but was not detected. The associated numerical value is the reporting limit.

J – The associated numerical value is considered an estimated concentration.

**Table E-4. Total mercury analytical results for the Lower Duwamish Triad sampling event.**

Sample ID	Hg (mg/kg)	Holding time exceedance (days)
SD-307-0000	0.09 J	22
SD-309-0000	0.24 J	22
SD-310-0000	0.14 J	22
SD-311-0000	0.11 J	22
SD-312-0000	0.20 J	22
SD-313-0000	0.39 J	22
SD-313-0001	0.16 J	19
SD-314-0000	0.23 J	21
SD-315-0000	0.09 J	21
SD-316-0000	0.15 J	22
SD-317-0000	0.10 J	22
SD-318-0000	0.10 J	22
SD-319-0000	0.16 J	22
SD-320-0000	0.21 J	22
SD-321-0000	0.18 J	22
SD-322-0000	0.14 J	22
SD-323-0000	0.20 J	21
SD-324-0000 <sup>a</sup>	0.20 J	22
SD-325-0000 <sup>a</sup>	0.20 J	21
SD-333-0000	0.10 J	12
SD-341-0000	0.09 R	11

Hg total mercury analyzed by EPA method 7471A.  
mg/kg milligrams per kilogram.

J The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.

R The sample result is rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

<sup>a</sup> field duplicate sample.

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**Table E-5. Sediment sample grain size distribution summary (percent retained in each grain size fraction) for the Lower Duwamish Triad sampling event.**

Sample ID	Gravel	Sand					Silt				Clay		
		Very Coarse	Coarse	Medium	Fine	Very Fine	Coarse	Medium	Fine	Very Fine	<10	9 to 10	8 to 9
Phi size	>-1	-1 to 0	0 to 1	1 to 2	2 to 3	3 to 4	4 to 5	5 to 6	6 to 7	7 to 8	8 to 9	9 to 10	<10
Sieve Size (microns)		10 to 18 (2000-1000)	18 to 35 (1000-500)	35 to 60 (500-250)	60 to 120 (250-125)	120 to 230 (125-62)	62.5-31.0	31.0-15.6	15.6-7.8	7.8-3.9	3.9-2.0	2.0-1.0	<1.0
SD-322-0002	0.0 J	0.2 J	1.7 J	2.9 J	8.9 J	14.3 J	6.2 J	24.4 J	20.8 J	7.1 J	3.6 J	3.2 J	6.7 J
SD-311-0002	24.6 J	4.2 J	7.0 J	14.5 J	22.6 J	11.7 J	2.2 J	4.1 J	3.9 J	3.1 J	1.1 J	0.4 J	0.6 J
SD-311-0003	-5.7 J	3.8 J	9.0 J	21.0 J	29.4 J	16.8 J	6.9 J	3.0 J	2.3 J	1.2 J	0.4 J	0.1 J	0.5 J
SD-317-0002	2.2 J	1.7 J	6.0 J	13.6 J	17.9 J	14.3 J	7.7 J	11.1 J	10.2 J	6.7 J	3.8 J	2.2 J	2.5 J
SD-317-0003	3.4 J	3.6 J	11.6 J	25.9 J	25.6 J	14.0 J	6.1 J	2.9 J	2.5 J	1.6 J	1.0 J	0.7 J	1.0 J
SD-315-0000	1.4	1.7	3.7	17.0	18.4	10.4	10.7	12.8	9.9	5.2	2.5	1.7	4.6
SD-307-0000	0.6	1.4	3.1	21.3	21.3	7.3	10.4	12.1	8.7	5.2	2.9	1.8	4.0
SD-309-0000	3.3	2.3	3.3	5.5	6.7	8.1	11.6	26.1	15.9	5.8	3.6	2.3	5.6
SD-310-0000	0.4	1.0	2.4	4.0	7.5	10.6	17.5	26.7	14.3	5.4	2.9	1.9	5.2
SD-311-0000	2.6	1.6	2.8	5.0	11.3	8.8	14.2	23.2	15.3	5.7	2.9	2.0	4.7
SD-312-0000	1.5	2.2	2.8	4.7	9.3	11.5	12.8	21.2	15.4	6.6	4.0	2.6	5.4
SD-313-0000	13.9	3.1	4.9	7.7	8.8	9.8	8.4	15.2	12.0	6.6	3.1	1.7	4.8
SD-314-0000	10.5	2.0	4.6	11.2	10.9	11.4	14.5	13.4	9.1	4.8	2.3	1.4	3.8
SD-316-0000	2.9	1.6	1.7	4.6	6.6	9.4	18.6	23.0	15.2	6.0	2.7	2.1	5.6
SD-317-0000	0.3	1.9	4.0	6.1	7.6	9.5	19.6	18.5	13.4	6.2	3.4	2.4	7.3
SD-318-0000	0.2	1.5	2.5	3.7	6.7	10.7	19.0	20.0	15.3	7.1	3.6	2.2	7.5
SD-319-0000	0.3	0.8	1.7	3.4	11.4	22.4	15.8	17.9	11.9	4.4	2.5	1.7	5.9
SD-320-0000	3.0	1.8	3.3	6.7	9.4	10.9	13.3	19.2	14.4	7.0	2.8	2.2	6.1
SD-321-0000	0.2	1.0	1.0	1.8	6.5	11.9	13.3	20.8	15.7	8.7	5.2	3.6	10.4
SD-322-0000	0.0	1.9	1.2	1.6	3.8	11.7	15.8	18.9	17.5	8.4	4.8	4.1	10.3
SD-323-0000	6.4	5.7	10.1	14.2	13.7	8.6	10.7	11.9	7.2	3.3	2.0	1.4	4.8
SD-324-0000*	0.2	1.0	0.9	1.7	6.1	11.9	14.1	19.1	16.9	8.8	5.2	3.8	10.4
SD-333-0000	1.1	2.4	6.8	10.0	12.2	8.3	16.2	15.7	10.0	5.7	4.0	1.8	5.9
SD-337-0000	2.1	1.8	7.0	13.6	15.4	19.8	11.2	12.3	7.5	3.0	2.0	1.3	3.0
SD-342-0000	4.8	1.0	3.8	9.0	13.5	15.1	13.7	15.6	10.5	5.0	2.6	1.8	3.6
SD-431 <sup>a</sup>	3.0	2.5	6.5	10.7	10.4	10.7	18.3	15.0	8.1	5.0	3.1	2.0	4.7

<sup>a</sup> field duplicate sample.

## Herrera Environmental Consultants, Inc.

### Memorandum

**To:** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc:** Peter Jowise, Herrera Environmental Consultants  
**From:** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date:** January 21, 2005  
**Subject:** Grain Size Distribution Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of grain size distribution data collected from the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 26 sediment samples, including 2 field duplicates between August 16 and 27, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples for grain size distribution using the PSEP method (PSEP 1996).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004) and the *Recommended Protocols for Measuring Selected Environmental Variables in Puget Sound (PSEP 1996)*.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. Data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

### Data Validation

#### Custody, Preservation, Holding Times, and Completeness—Acceptable with Qualification

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. The reported samples were analyzed within the required holding time of 180 days from the date of collection. The laboratory data package is complete and contains test results for all samples listed on the chain of custody forms.

The U.S Army Corps of Engineers selected five additional sediment samples (SD-311-0002, SD-311-0003, SD-317-0002, SD-317-0003, and SD-322-0002) for grain size distribution analysis after the field sampling event was completed. These five samples were frozen prior to analysis.

and have been qualified as estimated (flagged J) because freezing may affect the particle size distribution.

### **TriPLICATE ANALYSIS—ACCEPTABLE**

TriPLICATE analyses were performed at the required frequency. TriPLICATE results were reported for samples SD-315-0000, SD-333-0000, and a batch sample. The relative standard deviation (RSD) values for each phi size (ranging from 0.10 to 7.39 percent) met the control limits (less than 25 percent) established by the QAPP.

### **FIELD DUPLICATE—ACCEPTABLE**

Two field duplicate samples were analyzed for grain size distribution by the laboratory. As shown in the following table, the relative percent difference (RPD) values for percent fines (0.8 and 5.4 percent) met the control limits (less than 50 percent) established by the QAPP.

Sample ID	Field Duplicate ID	Sample Result (percent fines <sup>a</sup> )	Field Duplicate Result (percent fines <sup>a</sup> )	RPD
SD-321-0000	SD-324-0000	77.7	78.3	0.8
SD-333-0000	SD-431	59.3	56.2	5.4

### **Laboratory Reporting Limits—Acceptable**

The laboratory reported grain size distribution as specified in the method (PSEP 1996).

### **Overall Assessment of Data Quality**

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable as qualified.

### **Laboratory Communications**

The laboratory was not contacted regarding the grain size distribution analyses.

### **Definition of Data Qualifiers**

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U     The analyte was analyzed for but was not detected above the reporting limit.
- J     The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ    The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R     The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

## References

- Puget Sound Estuarine Program (PSEP). 1996. Recommended protocols for measuring selected environmental variables in Puget Sound. U.S. Environmental Protection Agency, Office of Coastal Waters.
- USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

## Herrera Environmental Consultants, Inc.

### Memorandum

**To:** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc:** Peter Jowise, Herrera Environmental Consultants  
**From:** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date:** June 6, 2008  
**Subject:** Mercury Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of mercury data collected from the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 21 sediment samples, including 2 field duplicates between August 16 and 27, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples for mercury using EPA Method 7471A (US EPA 1986).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004), the *Recommended Protocols for Measuring Selected Environmental Variables in Puget Sound* (PSEP 1996), the *Contract Laboratory Program National Functional Guidelines for Inorganic Data Review (Functional Guidelines)* (US EPA 2004), and the specified analytical method.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. Data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

### Data Validation

#### Custody, Preservation, Holding Times, and Completeness—Acceptable with Qualification

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. As discussed below, all reported samples were not analyzed within the required holding time of 28 days from the date of collection. The laboratory data package is complete and contains test results for all samples listed on the chain of custody forms.

The U.S Army Corps of Engineers selected the 21 sediment samples for mercury analysis after the field sampling event was completed. All samples were analyzed between 11 and 22 days

beyond the recommended holding time of 28 days. All samples that had a detected concentration of mercury above the laboratory reporting limit were qualified as estimated (flagged J) to indicate that sample results may be biased low. Mercury was not detected in sediment sample SD-341-0000 at the laboratory reporting limit and the result was rejected (flagged R), in accordance with Functional Guidelines. Because the reported concentration is expected to be lower than the actual concentration (associated with the extended holding time prior to analysis), the presence or absence of the analyte cannot be verified.

#### **Initial Calibration Verification—Acceptable**

An initial calibration verification (ICV) standard was reported with the analytical run. The percent recovery value (108.3 percent) met the control limits (90 to 110 percent) established by the QAPP.

#### **Continuing Calibration Verification—Acceptable**

Continuing calibration verification (CCV) standards were analyzed at the required frequency. The percent recovery values (ranging from 100.8 to 107.3 percent) met the control limits (80 to 120 percent) established by the QAPP.

#### **Blank Analysis—Acceptable**

Method blanks and instrument blanks were analyzed at the required frequency. Blanks did not contain levels of mercury above the laboratory reporting limit.

#### **Matrix Spike Analysis—Acceptable with Qualification**

Matrix spike (MS) analyses were performed at the required frequency. Matrix spike results were reported for samples SD-307-0000 and SD-333-0000. The percent recovery value for sample SD-307-0000 (111 percent) met the control limits (75 to 125 percent) established by the QAPP, but the MS percent recovery for sample SD-333-0000 (131 percent) exceeded the control limits. The only other sample (SD-341-0000) associated with the MS analysis of sample SD-333-0000 was undetected. In accordance with Functional Guidelines sample SD-333-0000 was qualified as estimated (flagged J), as shown in the following table.

Sample ID	Criteria	Qualifier
SD-333-0000	Associated matrix spike recovery > 125 percent and sample reported above detection limit.	J

### Duplicate Analysis—Acceptable

Duplicate analyses were performed at the required frequency. Duplicate results were reported for samples SD-307-0000 and SD-333-0000. Both samples had reported values of mercury that were less than five times the reporting limit. In accordance with Functional Guidelines, the difference between the sample and duplicate results must be less than a value equal to the reporting limit. Both duplicate analyses met this criteria and no data were qualified.

### Field Duplicate—Acceptable

Two field duplicate samples were analyzed by the laboratory. As shown in the following table, all relative percent difference (RPD) values (10.5 and 14.0 percent) met the control limits (less than 50 percent) established by the QAPP.

Sample ID	Field Duplicate ID	Sample Result (mg/kg)	Field Duplicate Result (mg/kg)	RPD
SD-321-0000	SD-324-0000	0.18 J	0.20 J	10.5
SD-314-0000	SD-325-0000	0.23 J	0.20 J	14.0

### Laboratory Control Sample Analysis—Acceptable

Laboratory control samples (LCS) were analyzed with each batch. The percent recovery values (116 and 116 percent) met the control limits (80 to 120 percent) established by the QAPP.

### Laboratory Reporting Limits—Acceptable

The laboratory met the detection limit (DL) of 0.1 mg/kg specified in the QAPP.

### Overall Assessment of Data Quality

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable as qualified with the exception of one rejected (flagged R) sample result value for holding time exceedance.

### Laboratory Communications

The laboratory was not contacted regarding the mercury analyses.

## Definition of Data Qualifiers

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U The analyte was analyzed for but was not detected above the reporting limit.
- J The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

## References

- Puget Sound Estuarine Program (PSEP). 1996. Recommended protocols for measuring selected environmental variables in Puget Sound. U.S. Environmental Protection Agency, Office of Coastal Waters.
- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C. December 1986.
- U.S. EPA. 2004. USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review. OSWER 9240.1-45. Office of Superfund Remediation and Technology Innovation (OSRTI), Washington DC. October 2004.
- USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

## Herrera Environmental Consultants, Inc.

### Memorandum

**To** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc** Peter Jowise, Herrera Environmental Consultants  
**From** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date** June 6, 2008  
**Subject** Lead Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of lead data collected from excess sediment waste buckets for the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 2 sediment samples from sediment waste buckets on September 1, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples by the toxicity characteristic leaching procedure (TCLP) for lead using EPA Methods 1311 and 6010B (US EPA 1986).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Contract Laboratory Program National Functional Guidelines for Inorganic Data Review (Functional Guidelines)* (US EPA 2004) and the specified analytical method.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. Data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

### Data Validation

#### Custody, Preservation, Holding Times, and Completeness—Acceptable

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. All reported samples were analyzed within the required holding time of 180 days from the date of collection. The laboratory data package is complete and contains test results for all samples listed on the chain of custody forms.

### **Initial Calibration Verification—Acceptable**

An initial calibration verification (ICV) standard was reported with the analytical run. The percent recovery value (98.5 percent) met the control limits (90 to 110 percent) established by Functional Guidelines.

### **Continuing Calibration Verification—Acceptable**

Continuing calibration verification (CCV) standards were analyzed at the required frequency. The percent recovery values (ranging from 97.1 to 104.0 percent) met the control limits (80 to 120 percent) established by Functional Guidelines.

### **ICP Interference Check Sample Analysis—Acceptable**

An ICP interference check samples (ICP-ICS) was analyzed with the sample batch. ICP interference check sample criteria (i.e., percent recovery value from 80 to 120 percent) established by Functional Guidelines were met for all compounds.

### **Blank Analysis—Acceptable**

Method blanks and instrument blanks were analyzed at the required frequency. Blanks did not contain levels of lead above the laboratory reporting limit.

### **Matrix Spike Analysis—Acceptable**

Matrix spike (MS) analyses were performed at the required frequency. Matrix spike results were reported for sample WD01. The percent recovery value (99.0 percent) met the control limits (75 to 125 percent) established by Functional Guidelines.

### **Duplicate Analysis—Acceptable**

Duplicate analyses were performed at the required frequency. Duplicate results were reported for sample WD01. The relative percent difference (RPD) value (0 percent) met the control limits (less than 20 percent) established by Functional Guidelines.

No field duplicates were collected.

### **Laboratory Control Sample Analysis—Not Analyzed**

Laboratory control samples (LCS) were not reported with the TCLP lead samples.

### **Laboratory Reporting Limits—Acceptable**

The QAPP did not specify detection limits for the TCLP lead analysis. The detection limits reported by the laboratory (0.1 ug/L) were reasonable for the method.

### **Overall Assessment of Data Quality**

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable.

### **Laboratory Communications**

The laboratory was not contacted regarding the lead analyses.

### **Definition of Data Qualifiers**

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U     The analyte was analyzed for but was not detected above the reporting limit.
- J     The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ    The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R     The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

### **References**

- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C. December 1986.

U.S. EPA. 2004. USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review. OSWER 9240.1-45. Office of Superfund Remediation and Technology Innovation (OSRTI), Washington DC. October 2004.

USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

# Herrera Environmental Consultants, Inc.

## Memorandum

**To** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc** Peter Jowise, Herrera Environmental Consultants  
**From** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date** June 6, 2008  
**Subject** Lead Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of lead data collected from excess sediment waste buckets for the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 2 sediment samples from sediment waste buckets on September 1, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples by the toxicity characteristic leaching procedure (TCLP) for lead using EPA Methods 1311 and 6010B (US EPA 1986).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Contract Laboratory Program National Functional Guidelines for Inorganic Data Review (Functional Guidelines)* (US EPA 2004) and the specified analytical method.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. Data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

## Data Validation

### Custody, Preservation, Holding Times, and Completeness—Acceptable

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. All reported samples were analyzed within the required holding time of 180 days from the date of collection. The laboratory data package is complete and contains test results for all samples listed on the chain of custody forms.

### **Initial Calibration Verification—Acceptable**

An initial calibration verification (ICV) standard was reported with the analytical run. The percent recovery value (98.5 percent) met the control limits (90 to 110 percent) established by Functional Guidelines.

### **Continuing Calibration Verification—Acceptable**

Continuing calibration verification (CCV) standards were analyzed at the required frequency. The percent recovery values (ranging from 97.1 to 104.0 percent) met the control limits (80 to 120 percent) established by Functional Guidelines.

### **ICP Interference Check Sample Analysis—Acceptable**

An ICP interference check samples (ICP-ICS) was analyzed with the sample batch. ICP interference check sample criteria (i.e., percent recovery value from 80 to 120 percent) established by Functional Guidelines were met for all compounds.

### **Blank Analysis—Acceptable**

Method blanks and instrument blanks were analyzed at the required frequency. Blanks did not contain levels of lead above the laboratory reporting limit.

### **Matrix Spike Analysis—Acceptable**

Matrix spike (MS) analyses were performed at the required frequency. Matrix spike results were reported for sample WD01. The percent recovery value (99.0 percent) met the control limits (75 to 125 percent) established by Functional Guidelines.

### **Duplicate Analysis—Acceptable**

Duplicate analyses were performed at the required frequency. Duplicate results were reported for sample WD01. The relative percent difference (RPD) value (0 percent) met the control limits (less than 20 percent) established by Functional Guidelines.

No field duplicates were collected.

### **Laboratory Control Sample Analysis—Not Analyzed**

Laboratory control samples (LCS) were not reported with the TCLP lead samples.

### Laboratory Reporting Limits—Acceptable

The QAPP did not specify detection limits for the TCLP lead analysis. The detection limits reported by the laboratory (0.1 ug/L) were reasonable for the method.

### Overall Assessment of Data Quality

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable.

### Laboratory Communications

The laboratory was not contacted regarding the lead analyses.

### Definition of Data Qualifiers

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U      The analyte was analyzed for but was not detected above the reporting limit.
- J      The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ     The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R      The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

### References

- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C. December 1986.

U.S. EPA. 2004. USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review. OSWER 9240.1-45. Office of Superfund Remediation and Technology Innovation (OSRTI), Washington DC, October 2004.

USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

## Herrera Environmental Consultants, Inc.

### Memorandum

**To** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc** Peter Jowise, Herrera Environmental Consultants  
**From** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date** June 6, 2008  
**Subject** Semivolatile Organic Compound Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of semivolatile organic compound (SVOC) data collected from the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 19 sediment samples, including 2 field duplicates between August 16 and 27, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples for SVOCs using EPA Method 8270C (US EPA 1986).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Contract Laboratory Program National Functional Guidelines for Organic Data Review (Functional Guidelines)* (US EPA 1999), the *Recommended Protocols for Measuring Selected Environmental Variables in Puget Sound (PSEP 1996)*, and the specified analytical method.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. Data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

### Data Validation

#### Custody, Preservation, Holding Times, and Completeness—Acceptable

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. Samples were archived (frozen) by the laboratory within 14 days from sample collection to extend the acceptable holding time. All reported samples were analyzed within the PSEP recommended holding time for archived samples of one year from collection to extraction and 40 days from extraction to analysis. The laboratory data package is complete and contains test results for all samples listed on the chain of custody form.

### **Instrument Tuning and Mass Calibration—Acceptable**

The tuning compound decafluorotriphenylphosphine was analyzed at the required frequency and all relative abundance values met the criteria established in Functional Guidelines.

### **Initial Calibration—Acceptable**

Initial calibrations were analyzed at the required frequency. Initial calibration criteria (i.e., percent relative standard deviation [RSD] values less than or equal to 30 percent and relative response factors [RRFs] greater than 0.05) established by Functional Guidelines were met for all target compounds.

### **Continuing Calibration Verification—Acceptable with Discussion**

Continuing calibration verification (CCV) standards were analyzed at the required frequency. Continuing calibration criteria (i.e., relative percent difference values less than or equal to 25 percent and RRFs greater than 0.05) established by Functional Guidelines with the exceptions noted below.

The percent difference (%D) values exceeded the 25 percent criterion for benzyl alcohol (37 percent), 2,4-dinitrophenol (29 percent), 4-nitrophenol (32 percent), and 4-nitroaniline (26 percent) in the CCV standard analyzed on 10/11/04, and for benzyl alcohol (29 percent) in the CCV standard analyzed on 10/13/04. No data were qualified because method criteria for an acceptable CCV specifies that up to any four SVOCs may fail to meet %D criteria as long as they have a %D value of less than or equal to 40.0 percent.

### **Blank Analysis—Acceptable**

Method blanks were analyzed at the required frequency. Blanks did not contain levels of SVOCs above the laboratory reporting limit.

### **Surrogate Analysis—Acceptable with Discussion**

Six surrogate compounds were analyzed with each sample and blank in accordance with the requirements in the method. All surrogate recovery values were within the control limits (50 to 130 percent) established by the QAPP with the following exceptions.

As shown in the table below, several samples had a surrogate percent recovery for d4-1,2-dichlorobenzene (ranging from 41.2 to 49.6 percent) that did not meet the control limit (50 to 130 percent) established in the QAPP. However, all surrogate recovery values met both the laboratory control limits and the EPA CLP control limits. No data were qualified because Functional Guidelines allows any one surrogate from each fraction (base/neutral or acid) to exceed percent recovery criteria.

Percent recoveries for surrogates d4-1,2-dichlorobenzene (41.2 percent) and d-5-nitrobenzene (48.8 percent) exceeded control limits established by the QAPP for sample SD-313-0001. No data were qualified because the percent recovery failures were marginal and both surrogate recoveries met the laboratory control limits and the EPA CLP control limits.

Sample	Surrogate	Percent Recovery	QAPP Control Limits (%)	Laboratory Control Limits (%)	CLP Control Limits (%)
SD-309-0000	d4-1,2-Dichlorobenzene	48.8	50-130	30-84	20-130
SD-310-0000	d4-1,2-Dichlorobenzene	47.2	50-130	30-84	20-130
SD-311-0000	d4-1,2-Dichlorobenzene	48.8	50-130	30-84	20-130
SD-312-0000	d4-1,2-Dichlorobenzene	46.8	50-130	30-84	20-130
SD-313-0001	d4-1,2-Dichlorobenzene	41.2	50-130	30-84	20-130
SD-313-0001	d-5-Nitrobenzene	48.8	50-130	28-103	23-120
SD-314-0000	d4-1,2-Dichlorobenzene	49.6	50-130	30-84	20-130
SD-316-0000	d4-1,2-Dichlorobenzene	47.6	50-130	30-84	20-130
SD-318-0000	d4-1,2-Dichlorobenzene	44.0	50-130	30-84	20-130
SD-319-0000	d4-1,2-Dichlorobenzene	48.4	50-130	30-84	20-130
SD-324-0000	d4-1,2-Dichlorobenzene	49.6	50-130	30-84	20-130

### Internal Standard Evaluation—Acceptable with Discussion

Internal standards were added to all samples, blanks, and QC samples as required. The response and retention time criteria established by Functional Guidelines were met with the exceptions noted below.

Response area for perylene-d12 in the matrix spike (46 percent) and matrix spike duplicate (40 percent) analysis for sample SD-315-0000 exceeded the 50 to 150 percent criteria established by Functional Guidelines. No data were qualified because the response area for perylene-d12 in sample SD-315-0000 (80 percent) met the control limits.

### Matrix Spike Analysis—Acceptable with Discussion

Matrix spike/matrix spike duplicate (MS/MSD) analyses were performed at the required frequency. MS/MSD results were reported for sample SD-315-0000. The percent recovery values (ranging from 54.7 to 103 percent) met the control limits (40 to 140 percent) established by the QAPP, with the following exceptions:

The percent recovery for pyrene (20.2 percent) in the matrix spike did not meet the control limits established by the QAPP. No data were qualified because the percent recovery for pyrene in the matrix spike duplicate (42.3 percent) met the control limits and all other data met the control limits.

### **Duplicate Analysis—Acceptable**

Matrix spike/matrix spike duplicate (MS/MSD) analyses were performed at the required frequency. MS/MSD results were reported for sample SD-315-0000. The relative percent difference (RPD) values (0 to 5.8 percent) met the control limits (less than 50 percent) established by the QAPP.

### **Field Duplicate Analysis—Acceptable with Qualification**

Two field duplicate samples were analyzed for SVOCs. The relative percent difference (RPD) was calculated for each compound where both duplicate values were greater than five times the reporting limit (RL). The difference between duplicate values was calculated if the detected compound concentration was less than five times the RL in either the sample or the field duplicate. A control limit of less than 50 percent RPD was established in the QAPP and a control limit of two times the RL was used to evaluate difference values. As shown in the following table, 15 compounds exceeded the RPD or difference control limits for field duplicate samples SD-314-0000 and SD-325-0000. Because all RPD or difference values met the control limits for sample SD-321-0000 and field duplicate SD-324-0000, only samples SD-314-0000 and SD-325-0000 were qualified as estimated (flagged J) for compounds that exceeded the control limits, as shown in the following table.

### **Laboratory Control Sample Analysis—Acceptable**

Laboratory control samples (LCS) were analyzed at the required frequency. All percent recovery values (ranging from 58.4 to 100 percent) met the control limits (50 to 130 percent) established in the QAPP.

### **Laboratory Reporting Limits—Acceptable**

The QAPP specified reporting limits ranging from 32 to 160 mg/kg for selected compounds. The detection limits reported by the laboratory met the reporting limits specified in the QAPP.

### **Overall Assessment of Data Quality**

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable as qualified.

### **Laboratory Communications**

The laboratory was not contacted regarding the SVOC analyses.

Compound (results in µg/kg)	RL (µg/kg)	Sample SD-314- 0000	Field Duplicate SD-325- 0000	RPD / Difference	Sample SD-321- 0000	Field Duplicate SD-324- 0000	RPD / Difference
Phenol	20	160 J	560 J	111 / NA	130	140	7 / NA
4-Methylphenol	20	21	31	NA / 10.0	ND	ND	
Naphthalene	20	28 J	160 J	NA / 132	ND	ND	
2-Methylnaphthalene	20	27 J	100 J	NA / 73	ND	ND	
Acenaphthylene	20	20 U	22	NA / 2	ND	ND	
Acenaphthene	20	42 J	190 J	NA / 148	ND	ND	
Dibenzofuran	20	25	54	NA / 29	ND	ND	
Diethylphthalate	20	28	20 U	NA / 8	20 U	27	NA / 7
Fluorene	20	51 J	220 J	NA / 169	ND	ND	
Phenanthrene	20	400 J	1,400 J	111 / NA	81	68	NA / 13
Carbazole	20	69	67	NA / 2	ND	ND	
Anthracene	20	130 J	330 J	87 / NA	30	25	NA / 5
Di-n-Butylphthalate	20	30	42	NA / 12	ND	ND	
Fluoranthene	20	600 J	1,100 J	59 / NA	270	230	16 / NA
Pyrene	20	570 J	1,500 J	90 / NA	240	210	13 / NA
Butylbenzylphthalate	20	20 U	26	NA / 6	34	27	NA / 7
Benzo (a) anthracene	20	220 J	500 J	78 / NA	98	81	NA / 17
Bis (2-ethylhexyl) phthalate	20	150	140	7 / NA	240	200	18 / NA
Chrysene	20	290 J	550 J	62 / NA	140	110	24 / NA
Benzo (b) fluoranthene	20	260	430	49 / NA	180	150	18 / NA
Benzo (k) fluoranthene	20	210 J	420 J	67 / NA	140	130	7 / NA
Benzo (a) pyrene	20	230 J	510 J	76 / NA	110	97	NA / 13
Indeno (1,2,3-cd) pyrene	20	110 J	170 J	43 / NA	52	46	NA / 6
Dibenz (a,h) anthracene	20	32	72	NA / 40	ND	ND	
Benzo (g,h,i) perylene	20	88 J	150 J	NA / 62	44	39	NA / 50

µg/kg micrograms per kilogram

RL Laboratory reporting limit

RPD Relative percent difference

Difference Difference between the sample result and field duplicate result

NA Not applicable

ND Not detected above the laboratory reporting limit; the RPD value or difference was not calculated.

<sup>a</sup> The relative percent difference was calculated if both the sample and field duplicate result were greater than five times the laboratory reporting limit (RL). If the sample or the field duplicate result was less than five times the RL, the difference between the two values was calculated. The control limit for RPD is 50 percent; the control limit for difference is two times the RL

U The material was analyzed for, but was not detected. The associated numerical value is the reporting limit.

J The associated numerical value is considered an estimate.

Bold values indicate that the RPD or difference control limit was exceeded.

# Herrera Environmental Consultants, Inc.

## Memorandum

**To:** John Wakeman, U.S. Army Corps of Engineers, Seattle District  
**cc:** Peter Jowise, Herrera Environmental Consultants  
**From:** Gina Catarra and Rob Zisette, Herrera Environmental Consultants  
**Date:** June 6, 2008  
**Subject:** Total Organic Carbon Data Validation for Lower Duwamish Triad Sampling Event

This memorandum presents a review of total organic carbon (TOC) data collected from the Lower Duwamish Triad Sampling Event site located in Seattle, Washington. Herrera Environmental Consultants collected 99 sediment samples, including 7 field duplicates between August 16 and 27, 2004. Analytical Resources, Inc. of Tukwila, Washington analyzed the samples for TOC using EPA Method 9060 modified by Puget Sound Estuarine Program (PSEP) for sediments (PSEP 1996).

The laboratory's performance was reviewed in accordance with quality control (QC) criteria outlined in the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004), the *Recommended Protocols for Measuring Selected Environmental Variables in Puget Sound (PSEP 1996)*, and the specified analytical method.

Quality control data submitted by the laboratory were reviewed; raw laboratory data was provided but was not reviewed. No data qualifiers (flags) were added to the sample results in the laboratory reports. Data validation results are summarized below, followed by a summary of laboratory communications and definitions of data qualifiers.

## Data Validation

### Custody, Preservation, Holding Times, and Completeness—Acceptable

Sample custody was properly maintained from sample collection to receipt at the laboratory. The samples were properly preserved and were received intact at the laboratory. The reported samples were analyzed within the required holding time of 28 days from the date of collection. The laboratory data package is complete and contains test results for all samples listed on the chain of custody forms.

### **Initial Calibration Verification—Acceptable**

Initial calibration verification (ICV) standards were analyzed with each batch. All percent recovery values (ranging from 94.4 to 103 percent) met the control limits (75 to 125 percent) established by the QAPP.

### **Blank Analysis—Acceptable**

Method blanks were analyzed at the required frequency. The method blanks did not contain reportable levels of TOC above the laboratory reporting limit.

### **Matrix Spike Analysis—Acceptable**

Matrix spike (MS) analyses were performed at the required frequency. MS results were reported for samples SD-206-0000, SD-310-0002, SD-315-0000, SD-315-0001, SD-323-0003, and SD-338-0000. The percent recovery values (ranging from 99.5 to 110 percent) met the control limits (75 to 125 percent) established by the QAPP.

### **TriPLICATE ANALYSIS—ACCEPTABLE**

TriPLICATE analyses were performed at the required frequency. TriPLICATE results were reported for samples SD-206-0000, SD-310-0002, SD-315-0000, SD-315-0001, SD-323-0003, and SD-338-0000. The relative standard deviation (RSD) values (ranging from 5.4 to 14.5 percent) met the control limits (less than 25 percent) established by the QAPP.

### **Field Duplicate—Acceptable**

Seven field duplicate samples were analyzed by the laboratory. As shown in the following table, all relative percent difference (RPD) values (ranging from 0.9 to 30 percent) met the control limits (less than 50 percent) established by the QAPP.

Sample ID	Field Duplicate ID	Sample Result (percent)	Field Duplicate Result (percent)	RPD
SD-314-0000	SD-325-0000	1.66	1.73	4.1
SD-320-0003	SD-328-0003	1.83	1.72	6.2
SD-321-0000	SD-324-0000	2.25	2.23	0.9
SD-323-0001	SD-326-0001	1.65	1.58	4.3
SD-207-0000	SD-433	2.11	1.56	30.0
SD-333-0000	SD-431	2.50	2.24	11.0
SD-338-0000	SD-432	1.81	1.92	5.9

### Laboratory Control Sample Analysis—Acceptable

Standard Reference Material (SRM) analyses were performed with each batch. The SRM (NIST # 8704) percent recovery values (ranging from 79.7 to 102.1 percent) met the control limits (75 to 125 percent) established by the QAPP.

### Laboratory Reporting Limits—Acceptable

The laboratory met the detection limit (DL) specified in the QAPP.

### Overall Assessment of Data Quality

The usability of the data is based on the guidance documents listed above. Upon consideration of the information presented here, the data are acceptable.

### Laboratory Communications

The laboratory was not contacted regarding the TOC analyses.

### Definition of Data Qualifiers

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U     The analyte was analyzed for but was not detected above the reporting limit.
- J     The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ    The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R     The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

## References

- Puget Sound Estuarine Program (PSEP). 1996. Recommended protocols for measuring selected environmental variables in Puget Sound. U.S. Environmental Protection Agency, Office of Coastal Waters.
- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C. December 1986.
- U.S. EPA. 2004. USEPA Contract Laboratory Program National Functional Guidelines for Inorganic Data Review. OSWER 9240.1-45. Office of Superfund Remediation and Technology Innovation (OSRTI), Washington DC. October 2004.
- USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

## Definition of Data Qualifiers

The following data validation qualifiers were used in the review of this data set. These qualifiers are from the *Lower Duwamish Triad Sampling Event Quality Assurance Project Plan (QAPP)* (USACE 2004).

- U     The analyte was analyzed for but was not detected above the reporting limit.
- J     The analyte was positively identified; the associated numerical value is an estimate of the concentration of the analyte in the sample.
- UJ    The analyte was not detected above the sample reporting limit. However, the reporting limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.
- R     The sample results are rejected due to serious deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

## References

- Puget Sound Estuarine Program (PSEP). 1996. Recommended protocols for measuring selected environmental variables in Puget Sound. U.S. Environmental Protection Agency, Office of Coastal Waters.
- U.S. EPA. 1986. Test Methods for Evaluating Solid Waste, Physical/Chemical Methods SW-846 Third Edition, Updates I, II, IIA, IIB, and III. Office of Solid Waste and Emergency Response, U.S. Environmental Protection Agency, Washington, D.C., December 1986.
- U.S. EPA. 1999. USEPA Contract Laboratory Program National Functional Guidelines for Organic Data Review. U.S. Environmental Protection Agency, Washington, D.C., EPA 540/R-99/008. October 1999.
- USACE. 2004. Lower Duwamish Triad Sampling Event Quality Assurance Project Plan. Prepared for U.S. Environmental Protection Agency, Region 10, by U.S. Army Corps of Engineers, Seattle District.

## **Appendix G: Evaluation of PCB Results from Method 8082 at Three EPA Laboratories**

**LOWER DUWAMISH TRIAD SAMPLING EVENT**

**PCB SEDIMENT SAMPLING**

**QUALITY CONTROL SUMMARY REPORT**

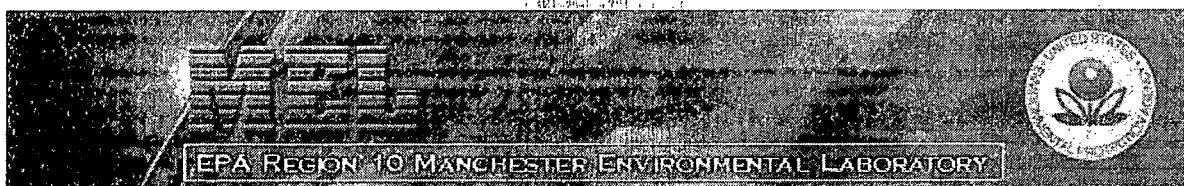
**Prepared By**

**SEATTLE DISTRICT CORPS OF ENGEERS  
Engineering and Technology Section**



**In Cooperation With**

**USEPA REGION X LABORATORY**



Contents:

**EPA Region 10 PCB Data Review**

**EPA Region 8 PCB Data Review**

**EPA Region 6 PCB Data Review**

## CHECKLIST FOR SEMIVOLATILE ORGANIC COMPOUNDS IN SEDIMENT

USACE DATA REVIEW - LOWER DUWAMISH TRIAD SAMPLING EVENT (PCB DATA)	
<b>GENERAL INFORMATION</b>	
<b>EPA Region 10</b> <b>Manchester Laboratory</b> 7411 Beach Drive E. Port Orchard, WA 98366 Barry V. Pepich, PhD, Director (360) 871-8701	
<b>Data Report prepared by</b> Cathy Martin (Seattle District USACE Chemist)	<b>Report Prepared:</b> 26 September 2008
<b>Date Sampled</b>	16-27 August 2004
<b>Date Received by Lab</b>	Acceptable. However, documentation not available.
<b>Date Analysis Began</b>	Acceptable. However, documentation not available. Analysis was complete by 12 January 2005
<b>Problems Noted (e.g., deviations from prescribed methods, analytical procedures):</b>	
USACE reviewer was unable to associate all samples with method QC data, based on information available at this time. Region 10 data review was assumed to incorporate the necessary evaluation. It is unknown what impact this has on data usability.	
<b>All required documents submitted?</b>	
USACE reviewer assumed that all necessary documentation was submitted. Data packages were not available at the time of USACE review.	
<b>Analytical Method</b>	
John V. Pepich, PhD, Director SW-846 8082	
<b>COMPLETENESS AND HOLDING CONDITIONS</b>	
<b>Samples Submitted = 51</b>	<b>Samples Analyzed = 51</b>
<b>Holding Conditions Acceptable? (One year for frozen sediments)</b>	Acceptable. Documentation of holding conditions was not available in the laboratory narratives. However, samples were frozen and the time from sampling to final documentation of results did not exceed 4.5 months. (Information above provided by John Wakeman.)
<b>If no, identify samples</b>	
NA	

(Signature) \_\_\_\_\_  
 Date: \_\_\_\_\_  
 (Print Name) \_\_\_\_\_  
 (Title) \_\_\_\_\_

(Signature) \_\_\_\_\_  
 Date: \_\_\_\_\_  
 (Print Name) \_\_\_\_\_  
 (Title) \_\_\_\_\_

<b>FORMAT</b>	
<b>Standard Report Sheets</b>	
Concentration in proper units and significant figures?	Yes
Qualifiers defined (e.g., U = undetected)	Yes (see EPA Region 10 QC Review pages 6, 7)
Samples detection limits (DL) provided for each analyte?	No. However, for data qualified as "U" have an implied DL.
<b>QA/QC SAMPLES</b>	
<b>Method Blanks</b>	
Total Number	12
Frequency	Minimum frequency of 1 batch ( $\leq$ 20 samples)
Aroclors detected outside < 20% of any reported value or <RL	None
<b>Certified Reference Materials<sup>a</sup></b>	
Total #	2
Frequency	2/51 samples
CRM Used	Sequim
Aroclors outside acceptance limit	None
<b>Field Duplicates</b>	
Total #	3
Frequency	5 duplicates/51 samples
Samples/Aroclors with >100% RPD or CV	None
<b>Matrix Spikes</b>	
Total #	4 MS/MSD pairs
Frequency	4/51 samples
Aroclors with <50% Recovery	None
<b>Detection Limits</b>	
Did any DL exceed SLK?	Yes
If yes, detection limits exceeding SL (identify samples)	Most sample detection limits exceed the project-specific reporting limits due to dilution. The dilution was required due to high concentrations of Aroclor-1254.

Surrogate Recovery	
Were surrogates added to all samples?	Yes
Identify samples with DCBP outside 40-140% TCMX outside 50-130%	SD-310-0000 = 353% DCBP SD-311-0000 = 353% DCBP; 263% TCMX SD-322-0000 = 416% DCBP; 297% TCMX SD-324-0000 = 154% DCBP SQ-1 = 141% DCBP OBS4342A1 (blank) = 44% TCMX SD-319-0003 = 33% TCMX SD-216-0000 = 50% DCBP SD-313-0003 = 31% TCMX SD-314-0002 = 49% TCMX SD-315-0002 = 150% DCBP SD-330-0000 = 150% DCBP SD-340-0000 = 161% DCBP SD-336-0000 = 144% DCBP SD-433 = 30%, TCMX OBS4344A1 (blank) = 151%, DCBP SD-315-0002 (MS) = 10%, TCMX SD-315-0002 (MSD) = 3% TCMX; 42% DCBP OBS4349A1 (blank) = 0%. TCMX SD-315-0003 = 49% TCMX
<b>See Region 10 Data Review (following) for assessment.</b>	
Laboratory Control Samples	
Total #	9
Frequency	9/51 samples
Aroclors with outside 50-130% Recovery	None

<sup>a</sup> According to EPA Region 10, SQ-1 had a spiked value of 170 ug/kg when created; early historical results averaged 120 ug/kg. No control limits have been set in this laboratory.



UNITED STATES ENVIRONMENTAL PROTECTION AGENCY  
REGION 10 LABORATORY  
7411 Beach Dr. East  
Port Orchard, Washington 98366

January 12, 2005

**MEMORANDUM**

**SUBJECT:** Data Review for the PCB Aroclor Analysis of Sediment and Water Samples from the Lower Duwamish Triad Project

Project Code: HWD-188A

Account Code: 040510P302D99C

**FROM:** Gerald Dodo, USEPA

**TO:** Howard Orlean, USEPA

**CC:** John Wakeman, Herrera Environmental Consultants  
Ravi Sanga, USEPA Allison Hiltner, USEPA Martha  
Lentz, USEPA Julius Nwosu, USEPA

The data review of the PCB Aroclor analyses of sediment and water samples collected from the above referenced site has been completed. Sediment sample extractions were performed using a modified EPA method 3541. The water samples were extracted using a modified draft EPA method 3511. Extract cleanups were performed using modified EPA methods 3660B, 3620B and 3665A. Analyses of the samples were performed using a modified EPA Method 8082 (PCB Aroclors).

The following samples were reviewed in this report:

04344100	04344104	04344108	04344112	04344116
04344118	04344119	04344120	04344122	04344123
04344124	04344126	04344127	04344128	04344130
04344131	04344132	04344136	04344140	04344141
04344144	04344145	04344147	04344148	04344152
04344158	04344164	04344166	04344167	04344168
04344169	04344173	04354000	04354004	04354005

04354007	04354008	04354009	04354010	04354012
04354015	04354016	04354017	04354018	04354020
04354021	04354022	04354025	04354026	04354032
04354033				

## DATA QUALIFICATIONS

The following comments refer to the laboratory performance in meeting the quality control specifications outlined in the analytical methods, the project's quality assurance plan, the Manchester Laboratory Quality Assurance Manual, Standard Operating Procedures, and professional judgement.

The conclusions presented herein are based on the information provided for the review.

### Holding Time

The sediment samples were stored frozen upon receipt by the laboratory with a one year holding time criterion applied for extractions. The sediment samples were extracted within four months from time of collection. The water sample, 04344173, was extracted within seven days from time of collection.

Extracts were analyzed within 40 days from preparation except for a diluted re-analysis of sample 04344112 which exceeded this criterion by six days. This re-analysis was performed for acquiring a more accurate measurement of PCB 1254 only. Due to the chemical stability of PCBs, this small exceedance was not judged to be significant therefore no qualifiers were applied based on holding times.

### Initial Calibrations (ICAL) -Acceptable

Single or multi-level initial calibrations were performed for the PCB Aroclors. Multi-level initial calibrations were performed for detected PCB Aroclors. The multi-level initial calibrations met the technical acceptance criteria for correlation coefficients ( $\geq 0.99$ ) and percent difference as compared to the curve ( $\pm 20\%$ ).

A multi-level initial calibration was performed for PCB 1248 but after the sample analyses. The detected PCB 1248 concentrations for the samples could only be estimated due to the presence of PCB 1254. This initial calibration was used as an attempt to get a better estimated value instead of use of single level calibration.

### Continuing Calibration Verification (CCV)

The PCB Aroclor CCV checks met the criteria for frequency of analysis and percent difference

*Data Review Lower Duwamish Triad -  
PCB Aroclors*

from the true value ( $\pm 15\%$  for target compounds and  $\pm 30\%$  for surrogate compounds) except as follows.

The CCV for the diluted extract re-analyses for samples 04344148 and 04344164 resulted with a percent difference of 17% for PCB 1254. These re-analyses were performed for acquiring a more accurate measurement of PCB 1254 only. A calibration standard was analyzed just before this CCV and a CCV was analyzed just after these two sample analyses which resulted with acceptable percent difference values. Therefore, no qualifiers were applied since the first CCV was only slightly out of criterion and these other calibration standard analyses were acceptable.

The CCV for the diluted extract re-analysis for sample 04344112 resulted with a percent difference of  $>15\%$  for PCB 1254. This re-analysis was performed for acquiring a more accurate measurement of PCB 1254 only. The reported PCB 1254 for this sample was qualified 'J'.

**Blanks - Acceptable**

Method blanks were prepared with each extraction batch. PCB Aroclors were not detected in the blanks.

**Surrogate Spikes**

The criteria for surrogate recoveries are 50-130% for tetrachlorometaxylene (TCMX) and 40-140% for decachlorobiphenyl (DCB). Several samples resulted with TCMX recoveries  $<50\%$  including the matrix spike analyses 04344130S1 and 04344130S2. The spike recoveries for PCB 1016 which is the earliest eluting Aroclor were still acceptable at 79% and 89%. For this reason, qualifiers were not applied toward the PCB Aroclors based on low TCMX recoveries.

Samples 04344112, 04344130, 04344164, 04344168, 04354010, 04354012, 04354018, and 04354022 resulted with DCB recoveries  $>140\%$ . Detected PCB Aroclor results for these samples were qualified 'J'. Non-detected PCB Aroclor results were not qualified since the high recoveries do not indicate a problem with the reported quantitation limits.

**Laboratory Fortified Blanks (LFB) - Acceptable**

LFBs were prepared using PCBs 1016 and 1260. The recoveries met the criterion of 50-130%.

**Matrix Spike/Matrix Spike Duplicate (MS/MSD) - Acceptable**

Samples 04344119, 04344130, and 04344168 were used for MS/MSD analyses (S1/S2). PCBs 1016 and 1260 were spiked into the MS/MSD aliquots. The recoveries met the criterion of 40-140%.

**Laboratory Duplicate**

Samples 04344169, 04354015, and 04354025 were analyzed in duplicate. Only PCB 1254 was

detected in these samples. The PCB 1254 relative percent differences (RPD) for samples 04344169, 04354015 and duplicates were <50.

The PCB 1254 RPD for sample 04354025 and duplicate was >50, therefore, the reported results for this Aroclor, sample and duplicate were qualified 'J'. This duplicate analysis was performed by taking aliquots out of separate jars. The duplicate analyses for samples 04344169 and 04354015 were performed by taking aliquots out of the same jar.

#### Reference Sample -Acceptable

The Sequim I reference sample was analyzed three times with the sediment samples. Only PCB 1254 was detected in this sample with results of 134, 174, and 190 ug/kg. These results were judged acceptable as the expected concentration for this sample is 170 ug/kg with an historical average of 112 ug/kg and a standard deviation of 30 ug/kg.

#### Compound Quantitation

The initial calibration functions were used for calculations. Reported quantitation limits were based on the initial calibration standards and sample size used for the analysis. Sediment sample results are on a dry weight basis. Detected PCB Aroclors below the quantitation limits were qualified 'J'.

All detected PCB 1248 results were qualified 'J' due to the potential interference from PCB 1254. In addition, the PCB 1260 detected results for sample 04344120, 04344127, and 04344128 were qualified 'J' for the same reason.

#### Compound Identification

Target compounds were identified based on chromatographic retention times of two dissimilar gas chromatography columns as determined from the initial calibration and peak pattern recognition.

Although detected PCB 1248 results are being reported, when considering the presence of PCB 1254 and weathering effects, it is possible that PCB 1242 may have been present instead.

The GC/ECD chromatograms for samples 04344126, 04344130, 04344131, and 04344133 displayed several early eluting peaks that did not match a PCB Aroclor pattern. Analysis of the extract for sample 04344130 by a GC/AED in a halogen specific mode determined that these compounds were not chlorinated or brominated.

Chemical Abstract Service (CAS) numbers with a "\*" indicates that the number was created at the Region 10 Laboratory due to lack of an existing one.

#### Overall Assessment

*Data Review Lower Duwamish Triad -  
PCB Aroclors*

All requirements for data qualifiers from the preceding sections were accumulated. Each sample data summary sheet and each compound was checked for positive or negative results. From this overall need for data qualifiers for each analysis was determined. In cases where more than one of the preceding sections required data qualifiers, the most restrictive qualifier has been added to the data.

**In general, all unqualified data can be used without restriction. The usefulness of qualified data should be treated according to the severity of the qualifier.** Should questions arise regarding the qualification of data and its relation to the usefulness, the reader is encouraged to contact Gerald Dodo at the Region 10 laboratory, phone number (360)871-8728.

## LABORATORY QUALIFIER/REMARK CODE DEFINITIONS

<b>Qualifier/ Remark Code</b>	<b>Definition (Codes Assigned to Values)</b>
<	<b>Microbiology</b> – Level of target organism present in the sample is less than detection limit. The reported value is the detection limit.
	<b>Flash Point</b> – The expected flash point temperature is less than the reported value.
>	<b>Microbiology</b> – Level of target organism exceeds upper limit for acceptable range of countable colonies (MF only) or exceeds MPN indices based on number of positive tubes (MPN only). The reported value is the upper limit.
	<b>Flash Point</b> – If the sample has a flashpoint, it is greater than the reported value.
J	The identification of the analyte is acceptable; the reported value is an estimate.
JK	The identification of the analyte is acceptable; the reported value is an estimate and may be biased high. The actual value is expected to be less than the reported value.
JL	The identification of the analyte is acceptable; the reported value is an estimate and may be biased low. The actual value is expected to be greater than the reported value.
K	The identification of the analyte is acceptable; the reported value may be biased high. The actual value is expected to be less than the reported value.
L	The identification of the analyte is acceptable; the reported value may be biased low. The actual value is expected to be greater than the reported value.
N	There is presumptive evidence that the analyte is present; the analyte is reported as a tentative identification.
NJ	There is presumptive evidence that the analyte is present; the analyte is reported as a tentative identification. The reported value is an estimate.
U	The analyte was not detected at or above the reported value.
UJ	The analyte was not detected at or above the reported value. The reported value is an estimate.

<b>Qualifier/ Remark Code</b>	<b>Definition (Codes With No Reported Values)</b>
A	Absent – The target parameter was analyzed for but was not present or was undetected. No value is reported with this qualification.
NA	Not Applicable, the parameter was not analyzed for, or there is no analytical result for this parameter. No value is reported with this qualification.
P	Present at an undetermined level – The target parameter is present but not quantifiable or no quantifiable result was determined. No value is reported with this qualification.
R	The presence or absence of the analyte can not be determined from the data due to severe quality control problems. The data are rejected and considered unusable. No value is reported with this qualification.

- T** A trace of the subject parameter was present. For asbestos analysis the subject parameter was identified but at a low level that a quantifiable percentage of content is unreliable. No value is reported with this qualification.
- TNTC** Too Numerous To Count – Any membrane where the total number of bacterial colonies exceeds 200 per membrane, or if the colonies are not distinct enough for accurate counting (i.e. confluent growth).
- F** The associated datum was generated using field methods and/or screening methods. The identification of the analyte is acceptable and the reported value has been found to be acceptable for use.
- JF** The associated datum was generated using field methods and/or screening methods. The identification of the analyte is acceptable and the reported value is an estimate.
- JKF** The associated datum was generated using field methods and/or screening methods. The identification of the analyte is acceptable; the reported value is an estimate and may be biased high. The actual value is expected to be less than the reported value.
- JLF** The associated datum was generated using field methods and/or screening methods. The identification of the analyte is acceptable; the reported value is an estimate and may be biased low. The actual value is expected to be greater than the reported value.
- UF** The associated datum was generated using field methods and/or screening methods. The analyte was not detected at or above the reported value.
- UJF** The associated datum was generated using field methods and/or screening methods. The analyte was not detected at or above the reported value. The reported value is an estimate.

**Qualifier/  
Remark Code)**

**Cross Reference to Older Codes**

**A** UND, ND – Undetected, Not detected

**NA** NAR, NAF – No analytical result, Not analyzed for

**P** PΝQ – Present but not quantified

**R** REJ - Rejected

**T** TRACE

**NOTE:** For any qualifier code see the QA memo or case narrative for a more detailed description of its use

<b>USACE DATA REVIEW- LOWER DUWAMISH TRIAD SAMPLING EVENT (PCB DATA)</b>	
<b>GENERAL INFORMATION</b>	
<b>Data Report prepared by</b> Cathy Martin (Seattle District USACE Chemist)	<b>Report Prepared</b> 26 September 2008
<b>Date Sampled</b>	16-27 August 2004
<b>Date Received by Lab</b>	Acceptable. 12 Dec 2004
<b>Date Analysis Began</b>	Acceptable. However, documentation not available. Results were initially reported on 31 Dec 2004
<b>Problems Noted (e.g., deviations from prescribed methods, analytical procedures):</b>	
The "Analyst Notes" provided in the narrative by EPA Region 8 incorrectly reports the surrogate recoveries for the samples and does not provide a project-specific assessment. Region 8 applied surrogate acceptance criteria of $\pm 30\%$ . However, the project-specific surrogate recovery limits for DCBP are 40-140%. Therefore, the samples qualified as "J" by Region 8 have been revised to not qualified.	
It is unknown why the analyte list and surrogates for Region 8 are different from Region 10, nor why method QC was not reported.	
<b>All required documents submitted?</b> Data packages not available for this review.	
<b>Analytical Method:</b> SW-846 8082	
<b>COMPLETENESS AND HOLDING CONDITIONS</b>	
<b>Samples Submitted = 30</b>	<b>Samples Analyzed = 29</b> Sample SD-331-0000 was lost during preparation at the laboratory.
<b>Holding Conditions Acceptable? one year for frozen sediments)</b>	Acceptable - received on 11 Dec 2004 at 2° C
<b>If no, identify samples</b>	NA
<b>FORMAT</b>	
<b>Standard Report Sheets</b>	
<b>Concentration in proper units and significant figures?</b>	Data packages unavailable
<b>Qualifiers defined (e.g., U = undetected)</b>	No - USACE reviewer assumed that CLP National Functional Guidelines qualifiers were applied.
<b>Samples detection limits (DL) provided for each analyte?</b>	No - USACE reviewer assumed DL only where U qualifier is applied by the laboratory.

<b>QA/QC Samples</b>	
<b>Method Blanks</b>	
Total Number	Data not available
Frequency	
Aroclors detected outside < 20% of any reported value or <RL	
<b>Certified Reference Materials</b>	
Total #	2
Frequency	2/29 samples
CRM Used	Sequim SQ-1
Aroclors outside acceptance limit	All are outside limits. No explanation was provided by EPA Region 8 in the case narrative.
<b>Field Duplicates</b>	
Total #	2 collected but not analyzed in this batch
Frequency	NA
Samples/Aroclors with >100% RPD or CV	NA
<b>Laboratory Control Samples</b>	
Total #	Data not available
Frequency	
Aroclors with outside 50-130% Recovery	
<b>Matrix Spikes</b>	
Total #	Data not available
Frequency	
Aroclors with <50% Recovery	
<b>Detection Limits</b>	
Did any DL exceed SLK?	Data not available
If yes, detection limits exceeding SL (identify samples)	
<b>Surrogate Recovery</b>	
Were surrogates added to all samples?	Yes. However, TCMX was not added as a surrogate.
Identify samples with DCBP outside 40-140% TCMX outside 50-130%	None
<b>Method Blanks (relative blank contamination)</b>	
For target compounds, was blank contamination >5% of a sample concentration?	Data not available.
If yes, identify Aroclor and samples	
<b>Laboratory Control Samples</b>	
Total #	Data not available.
Frequency	
Aroclors with outside 50-130% Recovery	

<sup>a</sup> According to EPA Region 10, SQ-1 had a spiked value of 170 ug/kg when created, early historical results averaged 120 ug/kg. No control limits have been set in this laboratory.

**PCBs in Soil  
From R10 Lower Duwamish Project**

**Analyst:** Mark A. Murphy

**Introduction:**

Thirty soil samples from the Lower Duwamish project were submitted to the EPA Region 8 laboratory for PCB analysis on 12/01/2004. The cooler was delivered by FedEx and the samples were received properly preserved and at 2°C. All holding times were met.

**Analytical Methods:**

EPA method 3545A, "Pressurized Fluid Extraction (PFE)," revision 1, January 1998. This extraction procedure was used for the PCB soil samples.

EPA method 3660B, "Sulfur Cleanup," CD-ROM revision 2, 1996.

EPA method 8082, "Polychlorinated Biphenyls (PCBs) By Gas Chromatography," revision 0, December 1996 was used for the PCB analysis.

**Analyst Notes:**

Surrogate recoveries reported on certificates of analysis are for Aroclor 1260. Aroclor recoveries in six samples were "J" flagged due to surrogate recoveries below the +/-30% acceptance range.

<u>EPA Region 8 sample number</u>	<u>EPA Sample Number</u>	<u>Location number</u>	<u>Surrogate recovery</u>
1.) AF00513	04-35-4001	SD-207-0000	Aroclor 1248 = 52.5% Aroclor 1254 = 47.4% Aroclor 1260 = 62.2
2.) AF00515	04-35-4003	SD-209-0000	Aroclor 1248 = 62.3% Aroclor 1254 = 56.3%
3.) AF00516	04-35-4006	SD-212-0000	Aroclor 1248 = 53.1% Aroclor 1254 = 48.0% Aroclor 1260 = 62.9%
4.) AF00517	04-35-4011	SD-217-0000	Aroclor 1248 = 65.7% Aroclor 1254 = 59.4%
5.) AF00520	04-35-4019	SD-337-0000	Aroclor 1248 = 64.6% Aroclor 1254 = 58.3%
6.) AF00522	04-35-4024	SD-342-0000	Aroclor 1248 = 64.2% Aroclor 1254 = 58.0%

EPA Region 8 sample AF00518, Location number SD-331-0000, EPA Region 10 sample 04-35-4013 was not reported (DNR) because the sample was lost during sample preparation. No further problems were encountered during the analysis.

**USACE DATA REVIEW- LOWER DUWAMISH TRIAD SAMPLING EVENT  
(PCB DATA)**

GENERAL INFORMATION	
<b>EPA Region 6</b> 10625 Fallstone Road, Houston, TX 77099 (281) 983-2100, phone; (281) 983-2248, fax	
<b>Data Report prepared by</b> Cathy Martin (Seattle District USACE Chemist)	<b>Report Prepared:</b> 26 September 2008
<b>Date Sampled</b>	16-27 August 2004
<b>Date Received by Lab</b>	Unknown
<b>Date Analysis Began</b>	Unknown;
<b>Problems Noted (e.g., deviations from prescribed methods, analytical procedures):</b> <p>The Region 6 Report Narrative incorrectly Reports that the samples were received past the storage hold time. The sample were stored at 0°C and shipped frozen to Region 6. Therefore, the results should not be qualified due to hold time exceedances.</p> <p>Region 6 Report Narrative states that B4L0908 BS1 Aroclor recoveries were below advisory limits of 70-130%. However, the project-specific control limits are 50-130 %. Therefore, the associated samples should not be qualified due low bias.</p> <p>The USACE reviewer concurs with the following case narrative that accuracy/bias cannot be assessed for Aroclors not included in the LCS. In addition, USACE concurs that high bias in matrix spike B4L1001-MS1/MSD1 should be qualified due to contributions from Aroclor 1248.</p>	
<b>All required documents submitted?</b> Yes	
<b>Chain-of-Custody and Sample Receipt Form</b> not included in data package.	
<b>Analytical Method</b>	Region 6 Laboratory Data
COMPLETENESS AND HOLDING CONDITIONS	
<b>Samples Submitted</b> = 21	<b>Samples Analyzed</b> = 21
<b>Holding Conditions Acceptable? one year for frozen sediments)</b>	Yes (see "Problems Noted" above)
<b>If no, identify samples</b>	NA
FORMAT	
<b>Standard Report Sheets</b>	Yes
<b>Concentration in proper units and significant figures?</b>	Yes
<b>Qualifiers defined (e.g., U = undetected)</b>	Yes
<b>Samples detection limits (DL) provided for each analyte?</b>	Yes

Comments on the following questions affect the quality of the data package. If the answer is No, the data package is not acceptable. If the answer is Yes, the data package is acceptable.

1. The data package includes all the required documents (e.g., Chain-of-Custody and Sample Receipt Form).

2. The data package includes all the required forms (e.g., Qualifiers defined, Detection limits).

Comments on the following questions affect the quality of the data package. If the answer is No, the data package is not acceptable. If the answer is Yes, the data package is acceptable.

1. The data package includes all the required documents (e.g., Chain-of-Custody and Sample Receipt Form).

2. The data package includes all the required forms (e.g., Qualifiers defined, Detection limits).

<b>Method Blanks</b>		
Total Number	2	
Frequency	2/21 samples (2 batches)	
Aroclors detected above 2x MDL ( $\mu\text{g}/\text{kg}$ )	None	
<b>Certified Reference Materials</b>		
Total #	1	
Frequency	1/21 samples	
CRM Used	Sequim	
Aroclors outside acceptance limit	Aroclor 1254 = 82.3 ppb, 48.4% recovery	
<b>Field Duplicates</b>		
Total #	Unknown	
Frequency	Unknown	
Samples/Aroclors with >100% RPD or CV	Unknown	
<b>Matrix Spikes</b>		
Total #	2	
Frequency	2/21 samples (2 batches)	
Aroclors with <50% Recovery	None	
<b>Detection Limits</b>		
Did any DL exceed SLK?	No	
If yes, detection limits exceeding SL (identify samples)	NA	
<b>Surrogate Recovery</b>		
Were surrogates added to all samples?	No	
Identify samples with DCBP outside 40-140% TCMX outside 50-130%	NA	
<b>Method Blanks (relative blank contamination)</b>		
For target compounds, was blank contamination >5% of a sample concentration?	No	
If yes, identify Aroclor and samples	NA	
<b>Laboratory Control Samples</b>		
Total #	2	
Frequency	2/21 samples (2 batches)	
Aroclors with outside 50-130% Recovery	None	

<sup>a</sup> According to EPA Region 10, SQ-1 had a spiked value of 170  $\mu\text{g}/\text{kg}$  when created; early historical results averaged 120  $\mu\text{g}/\text{kg}$ . No control limits have been set in this laboratory.



## UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

### Region 6 Laboratory

Environmental Services Branch  
10625 Fallstone Road, Houston, TX 77099  
Phone: (281)983-2100 Fax: (281)983-2248

### Final Analytical Report

Site Name ----- Lower Duwamish Triad / WA

Sample Collection Date(s)-- 08/16/04 - 08/19/04

Contact ----- U.S. EPA Region 10 Lab

Report Date ----- 01/18/05

Project # ----- 05RCRA028

Work Order(s) ----- 0411019

#### Analyses included in this report:

PCB 8082- R10 Solids, Dry Weight

#### Report Narrative

Samples were extracted approximately 3 1/2 months after collection. The samples were received in the laboratory approximately 3 months after collection. Concentrations should be considered minimum values.

Aroclor 1254 is qualified as a tentative identification and estimated on sample 0411019-16. This is because some interfering peaks were present that prevented getting a clear pattern match and made some integrations estimated.

Aroclors 1016 and 1260 recovered below advisory limits in the blank spike B4L0908-BS1. This blank spike applies to samples 0411019-01 to 0411019-10. Aroclor 1260 recovered below advisory limits in blank spike B4L1001-BS1. This blank spike applies to samples 0411019-11 to 0411019-21. Of the Aroclors outside limits, only 1260 was reported. This may indicate a low bias for Aroclor 1260 where reported. Because not all of the Aroclors are spiked, it is unknown if the low bias would apply to the other Aroclors reported (Aroclors 1248 and 1254).

Aroclor 1016 is qualified with a high bias on B4L1001-MS1/MSD1 due to contributions from Aroclor 1248.

Standard procedures for quality assurance and quality control were followed in the analysis and reporting of the sample results. The results apply only to the samples tested. This final report should only be reproduced in full.

Reporting limits are adjusted for sample size and matrix interference.

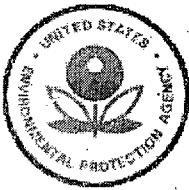
**Report Approvals:**

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Richard McMillin  
Region 6 Laboratory Manager

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Douglas A. Lipka  
Region 6 Laboratory Branch Chief



UNITED STATES ENVIRONMENTAL PROTECTION AGENCY

Region 6 Environmental Services Branch Laboratory

10625 Fallstone Road  
Houston, Texas 77099

**Sample Receipt and Disposal**

Data Management Coordinator: Christy Warren

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Data Management Coordinator Signature

Date

Date Transmitted: \_\_\_\_\_ / \_\_\_\_\_ / \_\_\_\_\_

Please have the U.S. EPA Project Manager/Officer call the Data Management Coordinator at 3-2137 for any comments or questions.

Please sign and date this form below and return it with any comments to:

Christy Warren  
Data Management Coordinator  
Region 6 Laboratory  
6MD-HS

---

Received by and Date

Comments:

The laboratory routinely disposes of samples 90 days after all analyses have been completed. If you have a need to hold these samples in custody longer than 90 days, please sign below.

---

Signature

Date

Please provide a reason for holding:



Environmental Protection Agency  
**Region 6 Laboratory**

10625 Fallstone Road, Houston, TX 77099  
Phone:(281)983-2100 Fax:(281)983-2248

**ANALYTICAL REPORT FOR SAMPLES**

Station ID	Laboratory ID	Matrix	Date Collected	Date Received
04344106	0411019-01	Solid	8/19/04 11:15	11/23/04 08:50
04344107	0411019-02	Solid	8/19/04 11:15	11/23/04 08:50
04344109	0411019-03	Solid	8/19/04 11:45	11/23/04 08:50
04344110	0411019-04	Solid	8/19/04 11:45	11/23/04 08:50
04344129	0411019-05	Solid	8/19/04 18:52	11/23/04 08:50
04344133	0411019-06	Solid	8/19/04 10:40	11/23/04 08:50
04344135	0411019-07	Solid	8/19/04 10:40	11/23/04 08:50
04344137	0411019-08	Solid	8/19/04 9:55	11/23/04 08:50
04344138	0411019-09	Solid	8/19/04 9:55	11/23/04 08:50
04344139	0411019-10	Solid	8/19/04 9:55	11/23/04 08:50
04344148	0411019-11	Solid	8/16/04 18:47	11/23/04 08:50
04344149	0411019-12	Solid	8/19/04 17:33	11/23/04 08:50
04344150	0411019-13	Solid	8/19/04 17:33	11/23/04 08:50
04344151	0411019-14	Solid	8/19/04 17:33	11/23/04 08:50
04344153	0411019-15	Solid	8/18/04 15:25	11/23/04 08:50
04344154	0411019-16	Solid	8/18/04 15:25	11/23/04 08:50
04344161	0411019-17	Solid	8/18/04 11:20	11/23/04 08:50
04344162	0411019-18	Solid	8/18/04 11:20	11/23/04 08:50
04344165	0411019-19	Solid	8/19/04 18:27	11/23/04 08:50
04344160	0411019-20	Solid	8/18/04 11:20	11/23/04 08:50
SQI	0411019-21	Solid	8/18/04 0:00	11/23/04 08:50



Environmental Protection Agency  
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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-01

**Station ID:** 04344106

Batch: B4L0908

Date Collected: 08/19/04

Matrix: Solid

Sample Wet Weight: 10.456g

%Solids: 62.30

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	280		72.9	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	202		52.6	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		15.4	1	12/09/04	12/15/04
Aroclor-1221	U		30.7	"	"	"
Aroclor-1232	U		15.4	"	"	"
Aroclor-1242	U		15.4	"	"	"
Aroclor-1248	76.6		15.4	"	"	01/06/05
Aroclor-1254	346		15.4	"	"	12/13/04
Aroclor-1260	116		15.4	"	"	12/15/04

**Detected Organics**

Target	Sample µg/kg dry	Reported Limit	Reported Value	Reported Limit	Reported Value
1016	U	15.4	15.4	15.4	15.4
1221	U	30.7	30.7	30.7	30.7
1232	U	15.4	15.4	15.4	15.4
1242	U	15.4	15.4	15.4	15.4
1248	76.6	15.4	76.6	15.4	76.6
1254	346	15.4	346	15.4	346
1260	116	15.4	116	15.4	116



Environmental Protection Agency  
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**Aroclors by EPA Method 8082 - GC/ECD**

Lab ID: 0411019-02

Station ID: 04344107

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.738g  
%Solids: 79.09

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	214		72.8	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	217		73.8	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		11.8	1	12/09/04	12/15/04
Aroclor-1221	U		23.5	"	"	"
Aroclor-1232	U		11.8	"	"	"
Aroclor-1242	U		11.8	"	"	"
Aroclor-1248	U		11.8	"	"	01/06/05
Aroclor-1254	86.7		11.8	"	"	12/13/04
Aroclor-1260	40.5		11.8	"	"	12/15/04



Environmental Protection Agency  
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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-03

**Station ID:** 04344109

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.249g  
%Solids: 58.70

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	327		78.6	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	223		53.6	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		16.6	1	12/09/04	12/15/04
Aroclor-1221	U		33.2	"	"	"
Aroclor-1232	U		16.6	"	"	"
Aroclor-1242	U		16.6	"	"	"
Aroclor-1248	U		16.6	"	"	01/06/05
Aroclor-1254	106		16.6	"	"	12/13/04
Aroclor-1260	168		16.6	"	"	12/15/04

**Subrogates**

Result	Analyte	Reporting Limit	Dilution	Prepared	Analyzed
U		16.6	1	12/09/04	12/15/04
U		33.2	"	"	"

**Targets**

Result	Analyte	Reporting Limit	Dilution	Prepared	Analyzed
U		16.6	1	12/09/04	12/15/04
U		33.2	"	"	"

**Subrogates**

Result	Analyte	Reporting Limit	Dilution	Prepared	Analyzed
U		16.6	1	12/09/04	12/15/04
U		33.2	"	"	"
U		10.6	"	"	"
U		16.6	"	"	"
U		16.6	"	"	"
U		16.6	"	"	"
U		16.6	"	"	"



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-04

**Station ID:** 04344110

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.254g  
%Solids: 62.44

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	317		81.3	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	245		62.8	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		15.6	1	12/09/04	12/15/04
Aroclor-1221	U		31.2	"	"	"
Aroclor-1232	U		15.6	"	"	"
Aroclor-1242	U		15.6	"	"	"
Aroclor-1248	U		15.6	"	"	01/06/05
<b>Aroclor-1254</b>	<b>89.3</b>		<b>15.6</b>	"	"	12/13/04
<b>Aroclor-1260</b>	<b>70.4</b>		<b>15.6</b>	"	"	12/15/04



Environmental Protection Agency  
**Region 6 Laboratory**

10625 Fallstone Road, Houston, TX 77099  
Phone:(281)983-2100 Fax:(281)983-2248

**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-05

**Station ID:** 04344129

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.244g  
%Solids: 86.22

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	203		71.7	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	202		71.4	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		11.3	1	12/09/04	12/15/04
Aroclor-1221	U		22.6	"	"	"
Aroclor-1232	U		11.3	"	"	"
Aroclor-1242	U		11.3	"	"	"
Aroclor-1248	U		11.3	"	"	01/06/05
Aroclor-1254	U		11.3	"	"	12/13/04
Aroclor-1260	U		11.3	"	"	12/15/04

**Targets**

Target	Calculated Concentration	Reported Concentration	Dilution	Prepared	Analyzed
1	11.3	11.3	1	12/09/04	12/15/04
2	22.6	22.6	1	12/09/04	12/15/04
3	11.3	11.3	1	12/09/04	12/15/04
4	11.3	11.3	1	12/09/04	12/15/04
5	11.3	11.3	1	12/09/04	12/15/04
6	11.3	11.3	1	12/09/04	12/15/04
7	11.3	11.3	1	12/09/04	12/15/04
8	11.3	11.3	1	12/09/04	12/15/04
9	11.3	11.3	1	12/09/04	12/15/04
10	11.3	11.3	1	12/09/04	12/15/04



Environmental Protection Agency  
Region 6 Laboratory

10625 Fallstone Road, Houston, TX 77099  
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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-06

Station ID: 04344133

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.381g  
%Solids: 57.37

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	342		81.4	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	265		63.1	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		16.8	1	12/09/04	12/15/04
Aroclor-1221	U		33.6	"	"	"
Aroclor-1232	U		16.8	"	"	"
Aroclor-1242	U		16.8	"	"	"
Aroclor-1248	85.0		16.8	"	"	01/06/05
Aroclor-1254	501		16.8	"	"	12/13/04
Aroclor-1260	149		16.8	"	"	12/15/04



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-07

**Station ID:** 04344135

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.441g  
%Solids: 68.29

Sample Qualifiers: R6HTS

**Surrogates**

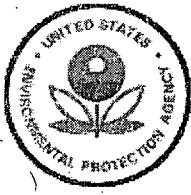
Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	285		81.2	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	219		62.4	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		14.0	1	12/09/04	12/15/04
Aroclor-1221	U		28.0	"	"	"
Aroclor-1232	U		14.0	"	"	"
Aroclor-1242	U		14.0	"	"	"
Aroclor-1248	459		14.0	"	"	01/06/05
Aroclor-1254	2,170		70.1	5	"	12/15/04
Aroclor-1260	436		14.0	1P	"	12/15/04

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	283		81.2	1P	12/09/04	12/15/04
Surr: Decachlorobiphenyl	219		62.4	1P	"	"
Surr: Pentachloro- hexachlorobiphenyl	133		14.0	1P	12/09/04	12/15/04
Surr: Hexachloro- heptachlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Octachloro- nonachlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Nonachloro- decachlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Trideca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Tetradeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Pentadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Hexadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Heptadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Octadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Nonadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Trideca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Tetradeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Pentadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Hexadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Heptadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Octadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04
Surr: Nonadeca- chlorobiphenyl	139		14.0	1P	12/09/04	12/15/04



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-08

**Station ID:** 04344137

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.408g  
%Solids: 59.27

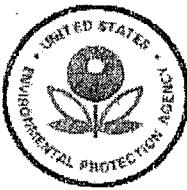
Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	327		80.7	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	267		65.9	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		16.2	1	12/09/04	12/15/04
Aroclor-1221	U		32.4	"	"	"
Aroclor-1232	U		16.2	"	"	"
Aroclor-1242	U		16.2	"	"	"
Aroclor-1248	562		16.2	"	"	01/06/05
Aroclor-1254	755		32.4	2	"	12/15/04
Aroclor-1260	212		16.2	1	"	12/15/04



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-09

Station ID: 04344138

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.168g  
%Solids: 69.16

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	283		79.5	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	284		79.8	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		14.2	1	12/09/04	12/15/04
Aroclor-1221	U		28.4	"	"	12/15/04
Aroclor-1232	U		14.2	"	"	"
Aroclor-1242	U		14.2	"	"	"
Aroclor-1248	6,920		284	20	"	01/06/05
Aroclor-1254	2,830		142	10	"	12/15/04
Aroclor-1260	688		28.4	2	"	12/15/04

Surrogates

Surrogate Analyte	Reporting Limit	Dilution	Prepared	Analyzed
Tetrachloro-meta-xylene	14.2	1	12/09/04	12/15/04
Decachlorobiphenyl	28.4	"	"	"
Octachloro-phenanthrene	14.2	"	"	"

Targets

Target Analyte	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	14.2	1	12/09/04	12/15/04
Aroclor-1221	28.4	"	"	"
Aroclor-1232	14.2	"	"	"
Aroclor-1242	14.2	"	"	"
Aroclor-1248	284	20	"	01/06/05
Aroclor-1254	142	10	"	12/15/04
Aroclor-1260	28.4	2	"	12/15/04



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-10

**Station ID:** 04344139

Batch: B4L0908  
Matrix: Solid

Date Collected: 08/19/04

Sample Wet Weight: 10.113g  
%Solids: 80.08

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	222		71.8	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	230		74.4	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		12.3	1	12/09/04	12/15/04
Aroclor-1221	U		24.7	"	"	"
Aroclor-1232	U		12.3	"	"	"
Aroclor-1242	U		12.3	"	"	"
<b>Aroclor-1248</b>	<b>635</b>		12.3	"	"	01/06/05
<b>Aroclor-1254</b>	<b>799</b>		37.0	3	"	12/15/04
<b>Aroclor-1260</b>	<b>201</b>		12.3	11	"	12/15/04

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	222		71.8	42-119	12/09/04	12/15/04
Surr: Decachlorobiphenyl	230		74.4	43-118	"	"
Surr: Pentachloro- hexachloroethane	100		12.3	"	"	"
Surr: Hexachloro- heptachloroethane	100		12.3	"	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		12.3	1	12/09/04	12/15/04
Aroclor-1221	U		24.7	"	"	"
Aroclor-1232	U		12.3	"	"	"
Aroclor-1242	U		12.3	"	"	"
<b>Aroclor-1248</b>	<b>635</b>		12.3	"	"	01/06/05
<b>Aroclor-1254</b>	<b>799</b>		37.0	3	"	12/15/04
<b>Aroclor-1260</b>	<b>201</b>		12.3	11	"	12/15/04



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-11

Station ID: 04344148

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/16/04  
Sample Wet Weight: 10.393g  
%Solids: 59.66

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	273		67.7	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	331		82.1	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		16.1	1	12/10/04	12/16/04
Aroclor-1221	U		32.3	"	"	"
Aroclor-1232	U		16.1	"	"	"
Aroclor-1242	U		16.1	"	"	"
Aroclor-1248	110		16.1	"	"	01/06/05
Aroclor-1254	1,980		161	10	"	12/16/04
Aroclor-1260	554		16.1	1	"	"



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-12

Station ID: 04344149

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.374g  
%Solids: 56.98

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	355		83.9	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	364		86.1	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		16.9	1	12/10/04	12/16/04
Aroclor-1221	U		33.8	"	"	"
Aroclor-1232	U		16.9	"	"	"
Aroclor-1242	U		16.9	"	"	"
Aroclor-1248	80.2		16.9	"	"	01/06/05
Aroclor-1254	990		33.8	2	"	12/16/04
Aroclor-1260	411		16.9	1	"	"

Surrogates

Surrogate	Min. ID	Reporting Limit	Dilution	Prepared	Analyzed
Tetrachloro-meta-xylene	U	16.9	1	12/10/04	12/16/04
Decachlorobiphenyl	U	33.8	"	"	"
Octachlorobiphenyl	U	16.9	"	"	"
Heptachlorobiphenyl	U	16.9	"	"	"
Hexachlorobiphenyl	U	16.9	"	"	"
Pentachlorobiphenyl	U	16.9	"	"	"
Tetrachlorobiphenyl	U	16.9	"	"	"
Trichlorobiphenyl	U	16.9	"	"	"
Chlorobiphenyl	U	16.9	"	"	"



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-13

Station ID: 04344150

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.292g  
%Solids: 57.16

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	347		81.6	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	326		76.7	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		17.0	1	12/10/04	12/16/04
Aroclor-1221	U		34.0	"	"	"
Aroclor-1232	U		17.0	"	"	"
Aroclor-1242	U		17.0	"	"	"
Aroclor-1248	32.1		17.0	"	"	01/06/05
Aroclor-1254	626		17.0	"	"	12/16/04
Aroclor-1260	576		17.0	"	"	"



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-14

Station ID: 04344151

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.069g  
%Solids: 63.36

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	331		84.4	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	321		81.9	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		15.7	1	12/10/04	12/16/04
Aroclor-1221	U		31.3	"	"	"
Aroclor-1232	U		15.7	"	"	"
Aroclor-1242	U		15.7	"	"	"
Aroclor-1248	U		15.7	"	"	01/06/05
<b>Aroclor-1254</b>	<b>68.9</b>		15.7	"	"	12/16/04
<b>Aroclor-1260</b>	<b>170</b>		15.7	"	"	"



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-15

**Station ID:** 04344153

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.522g  
%Solids: 69.00

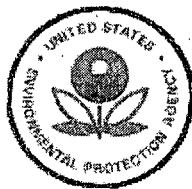
Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	275		79.9	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	312		90.7	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		13.8	1	12/10/04	12/16/04
Aroclor-1221	U		27.5	"	"	"
Aroclor-1232	U		13.8	"	"	"
Aroclor-1242	U		13.8	"	"	"
Aroclor-1248	54.5		13.8	"	"	01/06/05
Aroclor-1254	237		13.8	"	"	12/16/04
Aroclor-1260	460		13.8	"	"	"



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-16

Station ID: 04344154

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.379g  
%Solids: 62.53

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	306		79.5	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	319		82.9	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		15.4	1	12/10/04	12/16/04
Aroclor-1221	U		30.8	"	"	"
Aroclor-1232	U		15.4	"	"	"
Aroclor-1242	U		15.4	"	"	"
Aroclor-1248	U		15.4	"	"	01/06/05
Aroclor-1254	60.4	NJ	15.4	"	"	12/16/04
Aroclor-1260	298		15.4	"	"	"



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-17

**Station ID:** 04344161

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.192g  
%Solids: 53.80

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	372		81.6	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	360		78.9	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		18.2	1	12/10/04	12/16/04
Aroclor-1221	U		36.5	"	"	"
Aroclor-1232	U		18.2	"	"	"
Aroclor-1242	U		18.2	"	"	"
Aroclor-1248	582		18.2	"	"	01/06/05
Aroclor-1254	636		18.2	"	"	12/16/04
Aroclor-1260	173		18.2	"	"	"



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-18

**Station ID:** 04344162

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.159g  
%Solids: 67.32

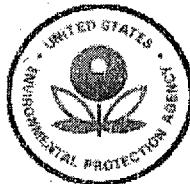
Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	269		73.5	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	294		80.3	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		14.6	1	12/10/04	12/16/04
Aroclor-1221	U		29.2	"	"	"
Aroclor-1232	U		14.6	"	"	"
Aroclor-1242	U		14.6	"	"	"
Aroclor-1248	46.1		14.6	"	"	01/06/05
Aroclor-1254	64.0		14.6	"	"	12/16/04
Aroclor-1260	By 16.0		14.6	"	"	"



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**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-19

**Station ID:** 04344165

Batch:B4L1001  
Matrix: Solid

Date Collected: 08/19/04  
Sample Wet Weight: 10.186g  
%Solids: 67.12

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	291		79.5	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	271		74.0	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		14.6	1	12/10/04	12/16/04
Aroclor-1221	U		29.3	"	"	"
Aroclor-1232	U		14.6	"	"	"
Aroclor-1242	U		14.6	"	"	"
Aroclor-1248	21.5		14.6	"	"	01/06/05
Aroclor-1254	198		14.6	"	"	12/16/04
Aroclor-1260	65.2		14.6	"	"	"



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Aroclors by EPA Method 8082 - GC/ECD

Lab ID: 0411019-20

Station ID: 04344160

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.348g  
%Solids:.55.71

Sample Qualifiers: R6HTS

Surrogates

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	349		80.4	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	335		77.2	43-118	"	"

Targets

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	110	U	17.3	1	12/10/04	12/16/04
Aroclor-1221	110	U	34.7	"	"	"
Aroclor-1232	110	U	17.3	"	"	"
Aroclor-1242	110	U	17.3	"	"	"
<b>Aroclor-1248</b>	<b>1,830</b>		86.7	5	"	01/06/05
Aroclor-1254	782		52.0	3	"	12/16/04
Aroclor-1260	161		17.3	"	"	"



Environmental Protection Agency  
**Region 6 Laboratory**

10625 Fallstone Road, Houston, TX 77099  
Phone:(281)983-2100 Fax:(281)983-2248

**Aroclors by EPA Method 8082 - GC/ECD**

**Lab ID:** 0411019-21

**Station ID:** SQI

Batch: B4L1001  
Matrix: Solid

Date Collected: 08/18/04  
Sample Wet Weight: 10.268g  
%Solids: 62.45

Sample Qualifiers: R6HTS

**Surrogates**

Analyte	Result µg/kg dry	Analyte Qualifiers	%Recovery	%Recovery Limits	Prepared	Analyzed
Surr: Tetrachloro-meta-xylene	297		76.2	42-119	12/10/04	12/16/04
Surr: Decachlorobiphenyl	268		68.7	43-118	"	"

**Targets**

Analyte	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Dilution	Prepared	Analyzed
Aroclor-1016	U		15.6	1	12/10/04	12/16/04
Aroclor-1221	U		31.2	"	"	"
Aroclor-1232	U		15.6	"	"	"
Aroclor-1242	U		15.6	"	"	"
Aroclor-1248	U		15.6	"	"	01/06/05
<b>Aroclor-1254</b>	<b>82.3</b>		<b>15.6</b>	"	"	<b>12/16/04</b>
Aroclor-1260	U		15.6	"	"	"



Environmental Protection Agency  
Region 6 Laboratory

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Aroclors by EPA Method 8082 - GC/ECD - Quality Control

Batch: B4L0908

Matrix: Solid

**Blank (B4L0908-BLK1)**

Prepared: 12/09/04 Analyzed: 12/15/2004

**Surrogates**

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC Limits
Surr: Tetrachloro-meta-xylene	201		249	80.7	42-119
Surr: Decachlorobiphenyl	176		249	70.7	43-118

**Blank (B4L0908-BLK1)**

Prepared: 12/09/04 Analyzed: 12/15/2004

**Targets**

ANALYTE	Result µg/kg dry	Analyte Reporting Qualifiers	Reporting Limit
Aroclor-1016	U	9.9	
Aroclor-1221	U	19.9	
Aroclor-1232	U	9.9	
Aroclor-1242	U	9.9	
Aroclor-1248	U	9.9	
Aroclor-1254	U	9.9	
Aroclor-1260	U	9.9	

**LCS (B4L0908-BS1)**

Prepared: 12/09/04 Analyzed: 12/15/2004

**Surrogates**

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC Limits
Surr: Tetrachloro-meta-xylene	193		246	78.5	42-119
Surr: Decachlorobiphenyl	185		246	75.2	43-118



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Aroclors by EPA Method 8082 - GC/ECD - Quality Control

Batch: B4L0908

Matrix: Solid

LCS (B4L0908-BS1)

Prepared: 12/09/04 Analyzed: 12/15/2004

Targets

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	%REC %REC	Limits
Aroclor-1016	610		19.7	986	61.9#	70-130
Aroclor-1260	590		19.7	986	59.8#	70-130

Matrix Spike (B4L0908-MS1)

Source: 0411019-05

Prepared: 12/09/04 Analyzed: 12/15/2004

Surrogates

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC
Surr: Tetrachloro-meta-xylene	252		286	88.1	42-119
Surr: Decachlorobiphenyl	227		286	79.4	43-118

Matrix Spike (B4L0908-MS1)

Source: 0411019-05

Prepared: 12/09/04 Analyzed: 12/15/2004

Targets

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	Source Result	%REC %REC	Limits
Aroclor-1016	810		22.9	1,440		71.1	50-150
Aroclor-1260	710		22.9	1,440		62.3	50-150

Matrix Spike Dup (B4L0908-MSD1)

Source: 0411019-05

Prepared: 12/09/04 Analyzed: 12/15/2004

Surrogates

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC
Surr: Tetrachloro-meta-xylene	280		286	97.9	42-119
Surr: Decachlorobiphenyl	242		286	84.6	43-118



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Aroclors by EPA Method 8082 - GC/ECD - Quality Control

Batch: B4L0908

Matrix: Solid

Matrix Spike Dup (B4L0908-MSD1)

Source: 0411019-05

Prepared: 12/09/04 Analyzed: 12/15/2004

Targets

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	Source Result	%REC %REC	RPD Limits	RPD Limit	
Aroclor-1016	902		22.9	1,140		79.1	50-150	10.7	25
Aroclor-1260	766		22.9	1,140		67.2	50-150	7.59	25



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Aroclors by EPA Method 8082 - GC/ECD - Quality Control

Batch: B4L1001

Matrix: Solid

**Blank (B4L1001-BLK1)**

Prepared: 12/10/04 Analyzed: 12/16/2004

**Surrogates**

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC Limits
<i>Surr: Tetrachloro-meta-xylene</i>	166		246	67.5	42-119
<i>Surr: Decachlorobiphenyl</i>	186		246	75.6	43-118

**Blank (B4L1001-BLK1)**

Prepared: 12/10/04 Analyzed: 12/16/2004

**Targets**

ANALYTE	Result µg/kg dry	Analyte Reporting Qualifiers	Reporting Limit
Aroclor-1016	U		9.8
Aroclor-1221	U		19.7
Aroclor-1232	U		9.8
Aroclor-1242	U		9.8
Aroclor-1248	U		9.8
Aroclor-1254	U		9.8
Aroclor-1260	U		9.8

**LCS (B4L1001-BS1)**

Prepared: 12/10/04 Analyzed: 12/16/2004

**Surrogates**

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC	%REC Limits
<i>Surr: Tetrachloro-meta-xylene</i>	220		246	89.4	42-119
<i>Surr: Decachlorobiphenyl</i>	199		246	80.9	43-118



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Aroclors by EPA Method 8082 - GC/ECD - Quality Control

Batch: B4L1001

Matrix: Solid

LCS (B4L1001-BS1)

Prepared: 12/10/04 Analyzed: 12/16/2004

Targets

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	%REC %REC	Limits
Aroclor-1016	724		19.7	983	73.7	70-130
Aroclor-1260	666		19.7	983	67.8*	70-130

Matrix Spike (B4L1001-MS1)

Source: 0411019-20

Prepared: 12/10/04 Analyzed: 12/16/2004

Surrogates

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC %REC	Limits
Surr: Tetrachloro-meta-xylene	402		444	90.5	42-119
Surr: Decachlorobiphenyl	371		444	83.6	43-118

Matrix Spike (B4L1001-MS1)

Source: 0411019-20

Prepared: 12/10/04 Analyzed: 12/16/2004

Targets

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	Source Result	%REC %REC	Limits
Aroclor-1016	1,630	K	88.9	1,780		91.6	50-150
Aroclor-1260	1,330		35.5	1,780	161	65.7	50-150

Matrix Spike Dup (B4L1001-MSD1)

Source: 0411019-20

Prepared: 12/10/04 Analyzed: 12/16/2004

Surrogates

ANALYTE	Result µg/kg dry	Analyte Qualifier	Spike Level	%REC %REC	Limits
Surr: Tetrachloro-meta-xylene	418		425	98.4	42-119
Surr: Decachlorobiphenyl	324		425	76.2	43-118



Environmental Protection Agency  
**Region 6 Laboratory**

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**Aroclors by EPA Method 8082 - GC/ECD - Quality Control**

Batch: B4L1001

Matrix: Solid

**Matrix Spike Dup (B4L1001-MSD1)**

Source: 0411019-20

Prepared: 12/10/04 Analyzed: 12/16/2004

**Targets**

ANALYTE	Result µg/kg dry	Analyte Qualifiers	Reporting Limit	Spike Level	Source Result	%REC %REC	%REC Limits	RPD RPD Limit
Aroclor-1016	1,480	K	85.1	1,700		87.1	50-150	9.65 25
Aroclor-1260	1,250		34.0	1,700	161	64.1	50-150	6.20 25



**Environmental Protection Agency  
Region 6 Laboratory**

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**Notes and Definitions**

NJ	There is presumptive evidence that the analyte is present; the analyte is reported as a tentative identification. The reported value is an estimate.
K	The identification of the analyte is acceptable; the reported value may be biased high. The actual value is expected to be less than the reported value.
R6A	This sample was extracted at a single acid pH.
R6HTS	Sample was prepared and/or analyzed past recommended holding time. Concentrations should be considered minimum values.
AES	Atomic Emission Spectrometer
CVAA	Cold Vapor Atomic Absorption
ECD	Electron Capture Detector
GC	Gas Chromatograph
GFAA	Graphite Furnace Atomic Absorption
ICP	Inductively Coupled Plasma
MS	Mass Spectrometer
NA	Not Applicable
NPD	Nitrogen Phosphorous Detector
NR	Not Reported
TCLP	Toxicity Characteristic Leaching Procedure
U	Undetected
#	Out of QC limits

Initial pressure in air analyses is the pressure at which the canister was received in psia (pounds per square inch absolute pressure).

The pH reported for Volatile liquid samples was tested using a 0-14 pH indicator strip for the purpose of verifying chemical preservation.

## **APPENDIX H: Field Records**

## **Appendix H: Sediment Sampling Field Notebooks**



1414 DEXTER AVENUE NORTH  
SUITE 200  
SEATTLE, WASHINGTON 98109  
(206) 281-7604  
FAX 281-7651

## SEDIMENT SAMPLE RECORD

Page 1 of 1

PROJECT NAME/NUMBER: PLANT 2 CO-01802-006 CC2-CC1

DATE: 8/18/04

LOCATION: Duramisit River

CREW: JOHN NAVARRO GEAR: VIBRATED  
ROS ZISKE  
ALEX SVANDER

### OBSERVATIONS:

### Notes:



HERRERA  
ENVIRONMENTAL  
CONSULTANTS

1414 DEXTER AVENUE NORTH  
SUITE 200  
SEATTLE, WASHINGTON 98109  
(206) 281-7604  
FAX 281-7651

## SEDIMENT SAMPLE RECORD

Page 1 of 1

PROJECT NAME/NUMBER:

Boeing Plant 2 00-01802 (Triad)

DATE: 8/18/04

LOCATION: Duwamish River

CARPORT: EISCH  
CREW: Eaton, Putnam GEAR: Vibracorer  
Waterman (recorder)  
Filutsonique (observer)

OBSERVATIONS:

SAMPLE NUMBER	STA. NO.	REP. NO.	TIME	WATER DEPTH	SEDIMENT DEPTH	CHARACTERISTICS (COLOR,TYPE,DEBRIS,ODOR)
SD-317	317	A	0926	NA	<2 ft	Black mud, sandy gravelly silt
SD-318-22	318-22	A	1012	6.9m	2 ft	Noticing to report (refusal)
SD-317-322	317-322	B	1030	6.8m	3 ft	Black fine mud/silt
SD-317-322	317-322	B	1120	6.5m	6'4"	Black fine mud
SD-321	321	A	1525	7.1m	2 ft	4.5' core good black fine mud
SD-319	319	A	1610	9.6m	2'4"	Sample went in but in open "
SD-314	314	A	1750	4.1m	2 ft	"
SD-314	314	B	1807	4.2m	2 ft	"
SD-315	315	A	1700	2.8m	2'4"	"
SD-313	313	A	1904	4.2m	~2 ft	(sheen)
313 A SD-313A						
Notes:						

J. J. H. Williamson

8/18/04



1414 DEXTER AVENUE NORTH  
SUITE 200  
SEATTLE, WASHINGTON 98109  
(206) 281-7604  
FAX 281-7651

## SEDIMENT SAMPLE RECORD

Page 1 of 1

PROJECT NAME/NUMBER: Boeing Plant 2 00-01802-006 (Triad)

DATE: 8/19/04

LOCATION: Duwamish River

Eaton, Putnam,  
CREW: Barker, GEAR: vibracorer  
Carpenter, Wickman

### OBSERVATIONS:

SAMPLE NUMBER	STA. NO.	REP. NO.	TIME	WATER DEPTH	SEDIMENT DEPTH	CHARACTERISTICS (COLOR,TYPE,DEBRIS,ODOR)
SD-307	307	A	0830	3' m	2 ft	water likely full over core - likely full over
		B	0950	2.5' m	< 0.5 ft	core follows
			0900	4' m	4 ft	core bent but got recovery
SD-312	312	A	0955	11' 9"	6' 3"	core bent, some recovery
SD-314	314	A	1040	11' 3"	7' (full)	core cut to 7' (true 4.5') before sampling
SD-309	309	A	1115	10' 6"	7' (full)	pre-cut to 7', full
SD-310	310	A	1145	8' 2"	7' 5' 8"	recovery
SD-311	311	A	1615	7' 1"	7'	full recovery, good recovery, very black silt 16/8/04
SD-312	312	A	1735	9' 4"	7' 6"	good recovery 17/8/04
SD-311	311	A	1615	7' 1"	7' 6"	full recovery, very black silt
SD-312	312	A	1700	16' 7"	4' 8"	*good recovery, very black, fine silt
SD-312	312	A	1733	9' 0"	6' 3"	good recovery, black fine silt
SD-323	323	A	1822	11' 5"	4' 5"	good recovery, black fine silt, woody debris
SD-315	315	A	1832	16' 5"	5' 33"	good recovery, very watery, black fine silt, less fine than
SD-313	313	B	1933	+5' 6"	3' 10"	more sand
SD-313	313	B	1959	14' 4"	4' 2"	very sandy, sheer, recovery 50%
						better recovery, woody debris

Notes:

\*good recovery = > 50%

Adams  
8/19/04

S

SS

## **Field Notes**

8/18/0419

Wednesday August 18, 2004.

00-01802-066 Plant 2

Rob Zisette (R2), Bruce Carpenter (E)

0600 Arrived at site and prepared vibrocorer. Needed work on vibrocorer switch.

0815 Left dock to pick up Anna Filutawik and John Wateman at South Park Marina. We had to fix electric switch for vibrocorer. Dick Sylvester came from Golder office in Redmond to do this.

We set up core processing equipment and table while switch was repaired.

0850 Bruce Carpenter gave 86 foot bungee.

0900h - 0915h - Deployed. Due to shallow water, removed weight carriage from vibrocorer.

0926h Station 317 began

A: Did not get good <sup>soil</sup> <sub>depth</sub> Generator

on/off switch malfunction. Less than

2 hr of recovery. Sample not retained

- 1005 Removed core liner, placed

residue in bucket for 1Dw

1012 → station 322 - new core 322(A)

Depth 5.9 m (water)

No recovery of sample.

- 1030h 322(B)

about 6.8 ft bottom (approx)

3 ft of core - 6.5 ft penetration

(10.5 ft - 4.0 ft)

= 3/6.5 recovery

Core not retained (to 1Dw)

redo,

- 1120h Advanced ~~322~~ again 322(C)

(6.5 m approx)

6 ft of mud penetration no recovery

8 ft 8" penetration

(11 ft) = 0% recovery

Cored core horizontal white transparent

to Harbor Island Marina. (unable

to split core on deck due to electric

saw that blows the circuit breaker)

Arrived at Harbor Is. Marina 1135h

20

processed core starting at 1025h.

Picture 2-18 (miss fire)  
17:00

2-18 > SD-322

2-19 / depths as shown  
(core appears winnowed (porous) below  
3 ft)

SD322- 0010m black silty clay

0020 -	" "	" clay with woody
0030 -	" "	Sand w/ woody <sup>char.</sup>
0040 -	" "	" sand
0050 -	" "	"

3-3.5 Wood predominates; below it is  
Sand.

\* Decided to cap cores for transport to lab.  
Left 140h to return to site

-1523 SD321 Depth 6.47m

76" depth of penetration = 6' 6"  
4' 6" recovery

8/18/04 21

Note to sample - rule of thumb

322 was to be a 6' sample,  
but only 4.5' was recovered.  
P.S. try to get a 1013 sample  
at least from 4-4.5' if  
possible. TOC a secondary  
but preferred.

- John Wahrman

1610h SD-319

Tom says 3.8m to bottom.  
(this seems to a deep)

Charlie's bathymetry was 13.5'  
but this is apparently a slope  
8' 9" penetration  
6' 9" ~~penetration~~ recovery

SD 318

1700h - 2.8m depth  
Core bent - only 2' 4" recovery

1730h - SD 314

water depth 4m  
penetration 9' 3"  
no recovery (less than 1 foot)

22

8/18/64

Removed catcher and reinstalled, disconnected

Tribal members informed us that

8pm begins gill net fishing

It extends to 8am on 8/19/64.

1807 h SD 314 (B)

(4.2 m to bottom

~~7.5 m to sediment bottom~~ I measured

Because core has been used ~~before~~

cannot estimate recovery from

mark on barrel. About 8' of

mud on barrel.  $3.3 \text{ m} = 10' \text{ in sc.}$

foot compared to deck measure

Recovery 4' 10" but lost a  
little at bottom on breaking

1907 h SD 313

~~Depth to bottom 4.2 m~~

" to depth of ~~core~~ 7.2 m

(measured) This is an overestimate!

Bottom Sheen - Picture 2-20

droplets of sheen 1/2-1"

Pb and Cu word chips - reddish

that penetration is poor.

23

Recovery  $\approx 2 \text{ ft}$

Took 1-2 samples. We should

revisit this site and try

again. If a better 1-2

sample occurs, we can write it

Remove 11950 h tridol at Harbor Is

*John S. Waterson*

John S. Waterson, US Army  
Corps of Engineers

24

August 19, 2004 00-01802-006 sunny ±75

0700 Meet @ Marina, Harbor Island  
 Rebekah Barker - USACE, writer  
 Bruce Carpenter - HEC, sampler  
 Charlie Eaton - captain  
 Tom Putnam - deck hand, writer

0723 Leave slip/marina  
 after loading ice, equip  
 Prepare corer (attach head, etc.)

0755 Reach South Park Marina

0807 Pick up John Waterman - USACE  
 Health & Safety briefing

0828 Arrive @ station SD-307

0830 zero core  
 take sample Rep A SD-307  
 water depth ~ 3 m 11' 6"

~~Max~~ penetration ~ 5.5' 10' (full)  
 recovery ~ 2'  
 core fell over (too much slack),  
 will lower slower next rep), deem

0850 SD-307 Rep B  
 water depth = 2.5 m  
 penetration ~ 6 m 3' 0"  
 recovery = < 0.5'  
 core likely fell over again

MWHSI Shm 8/19/04

25

0819/04 00-01802-006

SD-307 Rep C, wash tube, zero @ surface  
 water depth = 4 m  
 penetration = 6.2 m  
 recovery = 4'  
 Rep C → nose off shore 15' (Sm)  
 tough to pull core out, possibly  
 driven in at an angle —  
 core bent just over 6' up

SD-307 Rep D 16 8/19

Arrive 16 8/19

Prepare corer for next sample  
 decon core catcher, attach

0955 Arrive @ station SD-317

0955 zero for Rep B (A on 8/18/04)  
 water depth = 11' 9"  
 penetration = 8'  
 recovery = 6' 3"  
 Based on conversations with  
 Captain Charlie Eaton (earlier),  
 will be moving ~ 5 fm offshore  
 for these nearshore samples

1040 Arrive @ station SD-316  
 zero for Rep A  
 water depth = 11' 3"

MWHSI Shm 8/19/04

26

8/19/04 00-01802-006 sunny ±80°F

penetration = 7'

recovery = 316A = 7'

cut tube down to a 7' core  
in an attempt to stave off  
bending

1115 Arrive @ station SD-309

core tube pre-cut to 7'

SD - 309 A

water depth = 10' 6"

penetration = (5.5m max depth)

More recovery = 7' penetration

recovery = 7' (full)

1145 Arrive @ station SD-310

pre-cut core tube to 7'

zero @ water surface

water depth = 8' 2"

penetration = 7"

recovery = 5' 8"

1205 Stop @ South Park Marina,  
John Wakeman Disembark

1240 Arrive @ Harbor Island Marina

1240 Arrive back

1500 Reconvene @ Harbor Island Marina  
as tides begin to come back in

Whale Shri 8/19/04

27

8/19/04 00-01802-006 sunny ±80°F breezy

Prepare equipment

1517 Leave Marina (Harbor Island)

1555 Arrive @ station 311

- Prior to arrival @ station

while passing South Park Marina

we passed through a diesel  
fuel spill extending the entire  
length of the Marina and  
through the channel

- spray paint on outside of  
tube to better note penetration  
depth

- had to drill holes for rivets  
in tube

- sheet extends south of Marina,  
also we are downwind = strong odor

1600 - station is too shallow presently  
we will wait 20 minutes

- A plans  $\Rightarrow$  since tide is  
coming in we'll attempt sampling  
now & float off if we go aground

1615 SD - 311 Rep A

water depth = 7' 11"

penetration = 7' 6"

Whale Shri 8/19/04

28

8/19/04 00-01802-006 sunny 58°F  
windy

recovery = 7' 6" (full)

Note: fuel slick migrating south  
towards sampling sites with  
incoming tide

SD-311 RepA tube bent

1700 Arrive @ station 312

water depth = 7.7'

penetration = 90" = 7' 6" (7.5')

recovery = 58" = 4' 10" (4.8')

- spray painting outside was not effective because sediment is too soft to scratch the paint

1733 Arrive @ station SD-320

water depth = 9.4' - 0.4 = 9.0'

penetration = 7.5'

recovery = 75" = 6' 3" (6.25')

1827 Arrive @ station SD-323

water depth = ~~11' 8"~~ / ~~(11.73')~~ - 0.4

penetration = 7.5'

recovery = 4.5' (60%)

1852 Arrive @ station SD-315 A

water depth = ~~11.3'~~ / ~~0.1~~ = 10.7'

penetration = 7.5'

recovery = 64" = 5' 4" (5.33')

WhaleShoe 8/19/04

29

8/19/04 00-01802-006

very watery; some lost through  
core catcher, still 5' 4" remain

1923 Arrive @ station 313, Rep B

water depth =  $13.6' - 0.4 = 13.2'$

penetration = 7.5'

recovery =  $46" = 3' 10"$

Returning to SD-323 SD-313 B

wholesome but poor recovery  
yesterday

- \*\* Due to presence of small rock & algae lodged at top, we conclude that there was full penetration

- Definite shear in sediment and strong hydrocarbon smell

- Will move off shore 5 m & take SD-313 C since recovery was just over 50%

1958 Arrive @ SD-313 C

water depth = 14.4'

penetration = 7.5'

recovery = 4' 2"

WhaleShoe 8/19/04

## **Core Processing Notes**

12

27 FEB 04

- 08:00 ARRIVED ON-SITE  
 08:15 BACK HOE OPERATOR BEGAN WORKING IN-RIVER TO REMOVE OUTFALL CHP  
 08:45 ROCK-UP LAB SAMPLE  
 08:34 ROCK DOWN LAB SAMPLE  
 08:44 TURBIDITY ANALYSIS.  
 -NEAR HATCH TURBIDIMETER #1 4.99 NTU ± 5.04 NTU  
 -ROCK UP 1.01 NTU  
 -ROCK DOWN 1.25 NTU  
 08:55 BACK HOE OP. RE-POSITIONED THE BACK HOE, CAUSED A SHORT TERM (~1 MIN) TURBIDITY PLUME = UNABLE TO CATCH THE PLUME DOWNRIVER  
 09:05 ROCK DOWN: 1.01 NTU  
 09:07 WHILE ANALYZING ROCK-DOWN SAMPLE, CAP WAS FINALLY REMOVED, BACK HOE WAS IMMEDIATELY WASHED OUT OF RIVER WITH CAS AND ASSOC. CABLES.  
 09:15 IN-RIVER WORK COMPLETED, NO TURBIDITY PLUME VISIBLE. IT SAID THE POSSIBLE WORK WOULD BE DONE IN THE HARRISON PONDS.

13

19 AUG 04. -AS-

- Longer Duramish Sediment Sample  
 - CORES TAKEN YESTERDAY (8/18)  
 IN THE AFTERNOON. CORES STORED OVERNIGHT IN STORAGE ROOM ON ICE, CUSTODY SEALED.
- 10:44 SD - 319  
 - ONLY ABOUT 1 1/2" HEAD SPACE ON TOP, SMALL AMOUNT OF EXCESS WASTE REMOVED  
 - PHOTO 2-21-24), FROM TOP TO BOTTOM  
 - 1-2 FT  
 1-2 FT: BLACK CLAYEY SILT.  
 - PHOTO 2-26 2-3 FT  
 2-3 FT: BLACK CLAYEY, SANDY SILT WITH A BLACK SANDY LAYER FROM 2.4 FT TO 2.5 FT  
 PHOTO 2-27 3-4 FT  
 3-4 FT: FROM 3.0' TO 3.5' BLACK CLAYEY SILT  
 FROM 3.5 TO 4.0 FT, SILTY SAND (BLACK)  
 NOTE: CONTINUES BLACK SILTY SAND TO BOTTOM OF CORE (5.8 FT)
- 13:20 SD - 321  
 - ABOUT 8 1/2" HEAD SPACE ON TOP  
 - PICTURES 2-28, 29 OF CORE (TOP TO BOTTOM)

14

19 AUG 04

PHOTO 230: 1-2 FT

1.0 FT TO 1.1 FT BLACK CLAYEY SILT

1.1 FT TO 2.8 FT BLACK SILTY SAND

1.8 FT TO 2.0 FT BLACK CLAYEY SILT

PHOTO 234 - 2-3 FT

2.0-3.0 FT: BLACK CLAYEY SILT w/ SOME SAND

PHOTO 232 - 3 - 3.0 FT

- 3.0-3.5 FT: BLACK SILTY SAND

15:45 SD 314

- CORE PICTURES: PHOTO 233, 234 (TOP)  
BOTH

- VISIBLE STRATIGRAPHIC SECTION

\*NOTE: UNABLE TO ALIGN CORE

AND TUBING WITH DEPTH MEASUREMENT  
ON SIDEWALL. THUS, 1 FT T-CRSD-314 WILL BE 0 FT, WITH AN  
OFFSET OF 1 FT

- PHOTO 235: MIXED 1-2 FT SECTION

- 1.0 TO 1.1 BROWN SILTY CLAY

- 1.1 TO 2.0 DARK BROWN SILTY SAND

- PHOTO 236: MIXED 2-3 FT SECTION

- 2.0 TO 2.1 LIGHT GRAY CLAYEY SILT

- 2.1 TO 3.0 DE. GRAY/BLACK F-RD M-GRAINED  
SAND

15

SD 313 19 AUG 04

PHOTO 317: 3.0 FT TO 3.3 FT

3.0 TO 3.3 FT: DARK BROWN SILTY SAND

NOTE: DUE TO SMALL SECTION STRIPS (4),  
IRON WHICH SEDIMENT LEFT FOR

TCC - SEE JOURNAL

PHOTO 3-1: 1.0 TO 1.8 FT: SD 313

1.0 TO 1.8 FT - Black silty fine-to  
medium-grained sandJohn Bakeman - COE PM -  
due to HC odor & sheen noted  
in field, he requested SVOCs at  
this depth.

18:05 SD 318 2 sec 8/19/04

- CORE PHOTO 3-2

- 1.0-1.5 very dark gray, fine-  
grained sand w/ mica. NO  
debris.photo 3-3 1.0-1.5 mixed in  
bowl

19:20 SD-307 sec C

- core photo 3-4, 3-5, 3-6

1-2 Very dark gray, fine-to-med-grained  
sand. Sample was not in

16.

19 AUG 04

tube solidly → space 2 Limited

sample volume:

SD-307-0001 photo 3-7

Mixed up in bowl.

2.0' to 3.0' - VERY DK. GRAY FINE-GRAINED  
SAND

PHOTO 3-8 - SD-307-0001 @ 2.0 to 3.0'  
in bowl

3.0 to 3.8' - Very DK. Gray Fine-grained  
SAND

PHOTO 3-9 - SD-307-0001 @ 3.0 to 3.8'  
in bowl

20:24 SD-310 - CORES 0'-5'

Core photos 3-10, 3-11, 3-12

0-3.3' V. DK GRAY/BLACK alternate  
layers of fine-grained sand / organic  
silt / clayey silt.

3.3 to 4.5' V. DK. Gray fine-grained sand  
with some medium-grained sand,  
mica flakes.

4.5 to 5.0' Fine-grained sand matrix w/  
wood chips fragments.

PHOTO 3-13 Sample 1.0 to 2.0' SD-310  
(clayey silt)

19 AUG 04

PHOTO 3-14 Sample 2.0 to 3.0' SD-310  
SILT & Fine-grained SAND

PHOTO 3-15 Sample 3.0 to 4.0' SD-310  
Fine-grained sand

Clean up lab / decon bowls &  
spoons for tomorrow's

Sample processing  
10:25 Done

19 Aug

04

19-AUG-04

18

08/20/04, FRIDAY LOWER DUWAMISH  
SEDIMENT SAMPLING - SAMPLE  
PROCESSING CORES COLLECTED YESTERDAY  
8/19/04 - DP, AS, GC, BC, PJ

07:15 DP SETS UP SAMPLING AREA;  
GC SETS UP TO PROCESS SAMPLES  
FOR SHIPPING; AS/BC/PJ SETS UP  
CORES COLLECTED YESTERDAY

0855 SD 311 - Start CUTTING CORE

PHOTO 3-16 - AS cutting SD 311 core

CORE PHOTOS 3-17, 3-18, 3-19, 3-20, &  
3-21

Black clayey silt w/ tr. gravel 0-4"

Black silty SAND 4" to 8"

0-1 Black clayey silt cf. gravel

1-3' 2" Black clayey silt + gravel 3" φ

PHOTO 3-22 Sample 1.0 to 2.0 SD 311  
in bowl,

3' 2" to 8" Black silty SAND

36" brick fragment

photo 3-23 1' 2"-3' SD-311 in bowl

photo 3-24 3' to 4' SD-311 in bowl

10:10 Boeing Consultants stop by  
to watch sample processing

10:21 SD 313 Rep. C core

Diana M. Phula

19

SD 313C Plant 2

313C top is a brick fragment  
0'-0.5' clayey silt Black

0.5-1.0 Black silty SAND

1-2.3 Black clayey silt

2.3-4 Black silty SAND

photo 4-1 Core photo of 313C after  
samples collected

photo 4-2 fine-grained sandy silt

\*SD 313C 1.0 to 2.0' Sample in bowl

photo 4-3 SD 313C 2.0' to 3.0'

in bowl wet fine to med-grained  
SAND w/ gravel

photo 4-4 SD 313C 3.0' to 4.0'

sample in bowl - V.DEGAAT FINE TO  
11:10 SD 309A core

core photos 4-5, 4-6, & 4-7

rehomogenize 1.0'-2.0' sample 313C  
to fill the fourth jar for grain size

photo 4-8 1.0' to 2.0' 309A

Sample in bowl.

SD-309A 0-0.5' Black silty SAND

0.5-2.75 Black clayey silt, fragment

2.75 woody debris, black silty SAND

(2.75-7.0)

Diana M. Phula

20

## 20 Aug 04. Plant 2

photo 4-9 2.0' to 3.0' SD 309A

Sample in bowl. clayey silt w/<sup>sand</sup>  
Collected field duplicate (SD 327)  
2.0' to 3.0' sample from 309A  
Core

photo 4-10 3.0' to 4.0' SD 309A

Sample in bowl - DK Gray F-M SAND  
12:15 SD - <sup>300</sup> core cap mislabeled in  
core photos 4-11, 4-12, & 4-13 field

photo 4-14 Sample 1.0' to 2.0' SD <sup>327</sup>  
in bowl (clayey silt)

SD 315 320

0'-14" Black clayey SILT w/  
debris, brick, Shag, plastic wire.  
Coating 14" same as above w/ some  
Sand, 3.1.3' Bone clay 3.3' to 6.3'  
Black silty clay.

Not collected photo 4-15 Sample 2 - 2.0' to 3.0' SD 320

in bowl. (Clayey silt) <sup>DMP</sup>

Photo 4-16 Sample 3 - 3.0' to 4.0' SD <sup>320</sup>

in bowl (clayey silt)

Collected field duplicate SD 328-3

3.0' to 4.5' sample of SD 312 <sup>DMP</sup> 320

13:00 - 14:00 break

Diana M Phelan

21

## 20 Aug 04 Plant 2

14:20 SD 317B Core

Core photos 4-15, 4-16, & 4-17  
photo 4-18 - Sample 1 - 1.0' to 2.0'  
of SD 317B in Bowl.

photo 4-19 - Sample 2 - 2.0' to 3.0'  
of SD 317B in bowl

0.4' - 3.0' Black silty clay  
1.4' - 3.0' Black sandy silt w/  
trace clay

3.0' - 4.0' Black sandy silt w/  
gravel

4.0' - 6.6' Black silty sand

photo 4-20 Sample 3 - 3.0' to 4.0'  
of SD 317B in bowl.

16:05 SD 323 core being cut

Core photos 4-21, 4-22, & 4-23

photo 5-1 Sample 1 - 1.0' to 2.0' of  
core SD 323 in bowl

Collected field duplicate SD 326-0001

from 1.0' to 2.0' sample of SD 323

photos 5-2 Sample 2 - 2.0' to 3.0'

of SD 323

not collected photo 5-3 Sample 3 - 3.0' to 4.0' of  
SD 323

Diana M. Phelan

22

20 AUG. 04 Plant 2

SD 323

0.0-0.7' Black clayey silt w/ pieces  
of slag

0.7'-0.11' Black sandy silt w/ gravel

2.1'-2.5' Black silty clay, trace amounts  
of organic material2.5'-3.1' Black silty clay w/ large pieces  
of wood debris

3.1 to 4.5' Black silty sand

1650 SD 316A Core

core photos 5-43, 5-4, &amp; 5-5

photo 5-6 Sample 1 1.0' to 2.0' of  
SD 316A in bowlphoto 5-7 Sample 2 0.0' to 3.0' of  
SD 316A in bowlphoto 5-8 Sample 3 3.0' to 4.0' of  
SD 316A in bowl

0-0.3' Black silty sand

0.3'-3.2' Black silty clay

3.2'-4.2' Black silty sand, trace clay  
2" piece of asphalt-like material

4.2'-6.2' Dark grayish brown silty sand.

17:58 SD 315 Core

core photos 5-9, 5-10, &amp; 5-11

photo 5-12 Sample 1 0 to 2.0' of

Diana M. Phelan SD 315 in bowl

23

20 AUG. 04 Plant 2

photo 5-13 Sample 2 2.0' to 3.0'  
of SD 315 core in bowlphoto 5-14 Sample 3 3.0' to 4.0'  
of SD 315 core in bowl0-1.2' dk. brownish gray sandy  
silt trace wood debris

1.2'-4.8' Black silty sand

0-0.5' Core - no sediment

19150 SD 312 Core

core photos 5-15, 5-16, &amp; 5-17

photo 5-18 SAMPLE 1 1.0' to 2.0'  
of SD 312 core in bowlphoto 5-19 Sample 2 2.0' to 3.0'  
of SD 312 core in bowl (taken  
after sample collected).photo 5-20 Sample 3 3.0' to 4.0'  
of SD 312 core in bowl0-1.8' Black sandy silt w/ large  
4" wood piece (black) and smaller-  
sized wood pieces.1.8' to 4.8' Black silty sand  
1" dark black stain @ 2.7'  
or layer

Cleaned up, impact remaining cores

22:25 Done Diana M. Phelan

**Supplement to Appendix G  
Tables Showing Data Quality Review  
PCB Analyses by Method 8082**

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 10**

Field Sample ID	Lab Sample ID	Sample Description	Analys	Result <sup>a</sup>	Qualifier	Sample RT	Project RL Met?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK?	LSC % Rec.		MS % Rec.	MS CLs Met?	MSD % Rec.	MSD CLs Met?	MS/MSD RPD CL OK?	Dup RPD	Preservation Met?	Container OK?	Sample Date		Prep Hold Time Met?	Analysis Data	Analysis Hold Time Met?	Reference Sample CL Met?	% Completeness OK <sup>c</sup>
<b>Acceptance = 48-149%</b>																												
SD-311-0000	4344112	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	900 900 1800 900 900 3300 900 -	U U U U U J U -	900 900 1600 900 900 3300 900 353	No No No No No Yes No No													Yes	Yes	08/16/04	11/18/04	Yes	01/03/05	Yes	Yes	
SD-314-0000	4344124	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Decachlorobiphenyl Tetrachloromethylene	95 95 195 95 95 870 95 -	U U U U U U U -	95 95 195 95 95 870 95 112	No No No No No Yes No Yes													Yes	Yes	08/17/04	11/18/04	Yes	12/07/04	Yes	Yes	
SD-315-0000	4344128	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	65 65 145 65 65 150 110 110	U U U U U U J U	65 65 145 65 65 150 110 104	No No No No No Yes Yes Yes													Yes	Yes	08/17/04	11/18/04	Yes	12/05/04	Yes	Yes	
SD-317-0000	4344136	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	100 105 110 105 105 800 100 100	U U U U U U U U	100 105 110 105 105 800 100 100	No No No No No Yes No Yes													- Yes	Yes	08/16/04	11/18/04	Yes	12/05/04	Yes	Yes	
SD-319-0000	4344144	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	160 160 330 160 160 3100 160 160	U U U U U U U U	160 160 330 160 160 3100 160 160	No No No No No Yes No Yes													Yes	Yes	08/16/04	11/18/04	Yes	12/05/04	Yes	Yes	
SD-320-0000	4344148	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	760 760 1600 760 760 4200 760 760	U U U U U U U U	760 760 1600 760 760 4200 760 760	No No No No No Yes No No													Yes	Yes	08/16/04	11/18/04	Yes	12/15/04	Yes	Yes	
SD-321-0000	4344152	Reg sample	Arode-1018 Arode-1221 Arode-1232 Arode-1242 Arode-1248 Arode-1254 Arode-1260 Arode-1290 Decachlorobiphenyl Tetrachloromethylene	91 91 160 91 91 220 91 91	U U U U U J U U	91 91 160 91 91 220 91 91	No No No No No Yes No No													Yes	Yes	08/16/04	11/18/04	Yes	12/05/04	Yes	Yes	

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
EPA REGION 10**

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
FRA REGION 10**

TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
EPA REGION 10

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Result*	Qualifier	Sample RT	Project RL Met?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK	LSC % Rec.	MS % Rec.	MS CLs Met?	MSD % Rec.	MSD CLs Met?	MS/MSD RPD CL OK?	DUP RPD					Prep Head Time Met?	Analysis Date	Analysis Hold Time Met?	Reference Sample CL Met?**	% Completeness OK?***
							1221 > 49 ppm Others < 30 ppm																				
			Anodin-1254	610	U	Yes																					
			Anodin-1261	73	U	No																					
			Decachlorobiphenyl						129	Yes																	
			Tetrachloromethylene						30	Yes																	
SD-307-0000	4344100	Reg sample	Anodin-1018	190	U	190	No																				
			Anodin-1221	190	U	190	No																				
			Anodin-1232	300	U	300	No																				
			Anodin-1242	190	U	190	No																				
			Anodin-1248	190	U	190	No																				
			Anodin-1254	2000		2000	Yes																				
			Anodin-1260	190	U	190	No																				
			Decachlorobiphenyl						39	Yes																	
			Tetrachloromethylene						74	Yes																	
SD-310-0000	4344108		Anodin-1018	99	U	100	No																				
			Anodin-1221	100	U	100	No																				
			Anodin-1232	200	U	200	No																				
			Anodin-1242	100	U	100	No																				
			Anodin-1248	100	U	100	No																				
			Anodin-1254	500		Yes																					
			Anodin-1260	100	U	100	No																				
			Decachlorobiphenyl						353	No																	
			Tetrachloromethylene						97	Yes																	
SD-322-0000	4344158	Reg sample	Anodin-1018	98	U	No																					
			Anodin-1221	95	U	No																					
			Anodin-1232	198	U	No																					
			Anodin-1242	95	U	No																					
			Anodin-1248	98	U	No																					
			Anodin-1254	110		Yes																					
			Anodin-1260	98	U	No																					
			Decachlorobiphenyl						127	Yes																	
			Tetrachloromethylene						102	Yes																	
SD-323-0000	4344168		Anodin-1018	9.8	U	Yes																					
			Anodin-1221	9.8	U	Yes																					
			Anodin-1232	18	U	Yes																					
			Anodin-1242	9.8	U	Yes																					
			Anodin-1248	9.8	U	Yes																					
			Anodin-1254	4	J	Yes																					
			Anodin-1260	9.8	U	Yes																					
			Decachlorobiphenyl						127	Yes																	
			Tetrachloromethylene						53	Yes																	
SD-323-0003	4344157	Reg sample	Anodin-1018	5	U	Yes																					
			Anodin-1221	5	U	Yes																					
			Anodin-1232	10	U	Yes																					
			Anodin-1242	5	U	Yes																					
			Anodin-1248	5	U	Yes																					
			Anodin-1254	1.2	J	Yes																					
			Anodin-1260	5	U	Yes																					
			Decachlorobiphenyl						123	Yes																	
			Tetrachloromethylene						75	Yes																	
SD-339-0000	4354021	Reg sample	Anodin-1018	87	U	No																					
			Anodin-1221	87	U	No																					
			Anodin-1232	178	U	No																					
			Anodin-1242	87	U	No																					
			Anodin-1248	87	U	No																					
			Anodin-1254	488		Yes																					
			Anodin-1260	87	U	No																					
			Decachlorobiphenyl						130	Yes																	
			Tetrachloromethylene						90	Yes																	
			Reference Sample																								
	B	Sequel Reference	Anodin-1254	124					128	Yes																	

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
EPA REGION 10**

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**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 10**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Result <sup>a</sup>	Qualifier	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK?	LSC % Rec.	MS % Rec.	MS CLs Met?	MSU % Rec.	MSD CLs Met?	MS/MSD RPD	MS/MSD RPD CL OK?	Dup RPD	Preservation Met?	Container OK?	Sample Date	Prep Hold Time Met?	Analysis Date	Analytic Hold Time Met?	Reference Sample CL Met?	% Completeness OK <sup>b</sup>		
				(22) + 48 ppb Others = 20 ppb								35-110%		30-100%		40-100%		40-100%		40-100%		37°C	3M vial		30 days (predicted)		6 months (analyzed)	Reference Sample CL Met?	% Completeness OK <sup>b</sup>
SD-313-0002	4344122	Reg sample	Aroclor-1018	5.9	U	Yes																Yes	Yes	08/18/04	12/08/04	Yes	12/15/04	Yes	Yes
			Aroclor-1221	5.9	U	Yes																							
			Aroclor-1232	12	U	Yes																							
			Aroclor-1242	5.9	U	Yes																							
			Aroclor-1248	5.9	U	Yes																							
			Aroclor-1254	5.9	U	Yes																							
			Aroclor-1260	5.9	U	Yes																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-318-0001	4344141	Reg sample	Aroclor-1018	6.6	U	Yes																							
			Aroclor-1221	6.6	U	Yes																							
			Aroclor-1232	13	U	Yes																							
			Aroclor-1242	6.6	U	Yes																							
			Aroclor-1248	6.6	U	Yes																							
			Aroclor-1254	6.6	U	Yes																							
			Aroclor-1260	6.6	U	Yes																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-314-0002	4344127	Reg sample	Aroclor-1018	61	U	No																							
			Aroclor-1221	61	U	No																							
			Aroclor-1232	120	U	No																							
			Aroclor-1242	6.6	U	No																							
			Aroclor-1248	61	U	No																							
			Aroclor-1254	139	J	Yes																							
			Aroclor-1260	94	J	Yes																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-318-0001	4344145	Reg sample	Aroclor-1018	82	U	No																							
			Aroclor-1221	82	U	No																							
			Aroclor-1232	160	U	No																							
			Aroclor-1242	82	U	No																							
			Aroclor-1248	82	U	No																							
			Aroclor-1254	129	U	Yes																							
			Aroclor-1260	82	U	No																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-318-0002	4344147	Reg sample	Aroclor-1018	82	U	Yes																							
			Aroclor-1221	82	U	Yes																							
			Aroclor-1232	13	U	Yes																							
			Aroclor-1242	82	U	Yes																							
			Aroclor-1248	82	U	Yes																							
			Aroclor-1254	6.6	U	No																							
			Aroclor-1260	5.3	U	Yes																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-344-0000	4354026	Reg sample	Aroclor-1018	720	U	No																							
			Aroclor-1221	720	U	No																							
			Aroclor-1232	1400	U	No																							
			Aroclor-1242	720	U	No																							
			Aroclor-1248	720	U	No																							
			Aroclor-1254	1190	U	No																							
			Aroclor-1260	720	U	No																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										
SD-335-0000	4354017	Reg sample	Aroclor-1018	93	U	No																							
			Aroclor-1221	93	U	No																							
			Aroclor-1232	180	U	No																							
			Aroclor-1242	93	U	No																							
			Aroclor-1248	93	U	No																							
			Aroclor-1254	93	U	Yes																							
			Aroclor-1260	400	U	Yes																							
			Decachlorobiphenyl																										
			Tetrachloromethane																										

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
EPA REGION 10**

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 10**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Result <sup>**</sup>	Qualifier	Sample RL	Project RL Met?	Dilution	Surrogate Recovery	Surrogate CLs OK?	Method Blank CLs OK?	LSC % Rec.	MS % Rec.	MS CLs Met?	MSD % Rec.	MSD CLs Met?	MS/MSD RPD CL OK?	Dup RPD	Preservation Met?***	Container OK?	Sample Date	Prep Hold Time Met?	Analysis Date	Analysis Hold Time Met?	Reference Sample CL Met?***	% Completeness OK?****
			Anodex-1016	550	U	No						55-100%	20-100%	20-100%	40-100%	40-100%	40-100%	<10%	N/A	N/A		363 days (extended)	6 months (extended)	State = 70% Accuracy = 40-100%	82%	
			Anodex-1221	540	U	No																				
			Anodex-1232	>129	U	No																				
			Anodex-1242	58	U	No																				
			Anodex-1248	58	U	No																				
			Anodex-1254	58	U	No																				
			Anodex-1290	58	U	No																				
SD-314-0002	4344176	Reg sample																								
			Anodex-1016	48	U	No																				
			Anodex-1221	48	U	No																				
			Anodex-1232	81	U	No																				
			Anodex-1242	48	U	No																				
			Anodex-1248	48	U	No																				
			Anodex-1254	48	U	Yes																				
			Anodex-1290	48	U	Yes																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
SD-315-0002	4344130	Reg sample																								
			Anodex-1016	48	U	No																				
			Anodex-1221	48	U	No																				
			Anodex-1232	81	U	No																				
			Anodex-1242	48	U	No																				
			Anodex-1248	48	U	No																				
			Anodex-1254	48	U	Yes																				
			Anodex-1290	48	U	No																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
SD-330-0000	4354012	Reg sample																								
			Anodex-1016	84	U	No																				
			Anodex-1221	81	U	No																				
			Anodex-1232	189	U	No																				
			Anodex-1242	84	U	No																				
			Anodex-1248	84	U	No																				
			Anodex-1254	660	J	Yes																				
			Anodex-1290	94	U	No																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
SD-336-0000	4354018	Reg sample																								
			Anodex-1016	71	U	No																				
			Anodex-1221	71	U	No																				
			Anodex-1232	140	U	No																				
			Anodex-1242	71	U	No																				
			Anodex-1248	71	U	Yes																				
			Anodex-1254	169	J	Yes																				
			Anodex-1290	71	U	No																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
SD-340-0000	4354022	Reg sample																								
			Anodex-1016	76	U	No																				
			Anodex-1221	76	U	No																				
			Anodex-1232	150	U	No																				
			Anodex-1242	76	U	No																				
			Anodex-1248	76	U	No																				
			Anodex-1254	230	J	Yes																				
			Anodex-1290	76	U	No																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
SD-433	4354033	Reg sample																								
			Anodex-1016	97	U	No																				
			Anodex-1221	97	U	No																				
			Anodex-1232	139	U	Yes																				
			Anodex-1242	97	U	Yes																				
			Anodex-1248	97	U	Yes																				
			Anodex-1254	97	U	Yes																				
			Anodex-1290	97	U	Yes																				
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							
	0884344A1	Blank	Anodex-1016	-	A																					
			Anodex-1221	-	A																					
			Anodex-1232	-	A																					
			Anodex-1242	-	A																					
			Anodex-1248	-	A																					
			Anodex-1254	-	A																					
			Anodex-1290	-	A																					
			Decachlorobiphenyl																							
			Tetrachloromethylenes																							

**TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**PCB REGION 10**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Result <sup>a</sup>	Qualifier	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK	LSC % Rec.	MS % Rec.	MS CLs Met?	MSD % Rec.	MSD CLs Met?	MS/MSD RPD CL OK?	Dup RPD	Preservation Met?	Container OK?	Sample Date	Prep Hold Time Met?	Analysis Date	Reference Sample CL Met?**	% Completeness OK?***		
<b>Method Blank</b>																											
	OBS4344A2	Blanks	Aroclor-1016	-	A																						
			Aroclor-1221	-	A																						
			Aroclor-1232	-	A																						
			Aroclor-1242	-	A																						
			Aroclor-1248	-	A																						
			Aroclor-1254	-	A																						
			Aroclor-1260	-	A																						
			Decachlorobiphenyl	-																							
			Tetrachloromethylylene	-																							
									137	Yes																	
									65	Yes																	
<b>Laboratory Control Sample</b>																											
		LCS	Aroclor-1016	-																							
			Aroclor-1260	-																							
			Decachlorobiphenyl	-																							
			Tetrachloromethylylene	-																							
<b>Matrix Spike</b>																											
SD-315-0002 - 4344130		Matrix Spike	Aroclor-1016	94	U	-	No						89	Yes													
			Aroclor-1260	94	U	-	No						112	Yes													
			Decachlorobiphenyl	-																							
			Tetrachloromethylylene	-					135	Yes																	
									10	No																	
<b>Matrix Spike Duplicate</b>																											
SD-315-0002 - 4344130		Matrix Spike	Aroclor-1016	94	U	-	No						79	Yes	11.0	Yes											
			Aroclor-1260	94	U	-	No						115	Yes	2.6	Yes											
			Decachlorobiphenyl	-																							
			Tetrachloromethylylene	-					142	No																	
									3	No																	
<b>Primary Samples</b>																											
SD-333-0000 - 4354015		Reg sample	Aroclor-1016	94	U	-	No																				
			Aroclor-1221	94	U	-	No																				
			Aroclor-1232	100	U	-	No																				
			Aroclor-1242	94	U	-	No																				
			Aroclor-1248	94	U	-	No																				
			Aroclor-1254	1000	U	-	No																				
			Aroclor-1260	94	U	-	No																				
			Decachlorobiphenyl	-					133	Yes																	
			Tetrachloromethylylene	-					68	Yes																	
SD-333-0000 - 4354015		Duplicate	Aroclor-1016	110	U	-	No																				
			Aroclor-1221	110	U	-	No																				
			Aroclor-1232	230	U	-	No																				
			Aroclor-1242	110	U	-	No																				
			Aroclor-1248	110	U	-	No																				
			Aroclor-1254	840	U	-	No																				
			Aroclor-1260	110	U	-	No																				
			Decachlorobiphenyl	-					178	Yes																	
			Tetrachloromethylylene	-					75	Yes																	
SD-333-0000 - 4354015		Reg sample	Aroclor-1016	88	U	-	No																				
			Aroclor-1221	88	U	-	No																				
			Aroclor-1232	130	U	-	No																				
			Aroclor-1242	65	U	-	No																				
			Aroclor-1248	88	U	-	No																				
			Aroclor-1254	290	U	-	No																				
			Aroclor-1260	88	U	-	No																				
			Decachlorobiphenyl	-					126	Yes																	
			Tetrachloromethylylene	-					79	Yes																	
SD-333-0000 - 4354020		Reg sample	Aroclor-1016	86	U	-	No																				
			Aroclor-1221	86	U	-	No																				
			Aroclor-1232	170	U	-	No																				
			Aroclor-1242	88	U	-	No																				
			Aroclor-1248	86	U	-	No																				
			Aroclor-1254	270	U	-	No																				
			Aroclor-1260	109	U	-	No																				
			Decachlorobiphenyl	-					121	Yes																	
			Tetrachloromethylylene	-					80	Yes																	

TABLE G-1 - LOWER DUWAMISH TRIAD SAMPLING EVENT  
PCB DATA REVIEW  
EPA REGION 10

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Result <sup>a</sup>	Qualifier	Sample #	Project RL Met?	Date Taken	Surrogate Recovery	Surrogate CLs OK?	Blank CLs OK?	LSC % Rec.	MS % Rec.	MS CLs Met?	MSD % Rec.	MSD CLs Met?	MS/MSD RPD	MS/MSD RPD CL OK?	Dup RPD	Preservation Met? <sup>b</sup>	Container OK?	Sample Date	Prep Hold Time Met?	Analysis Date	Analysis Hold Time Met?	Reference Sample CL Met? <sup>c</sup>	% Completeness OK? <sup>d</sup>
<b>Reference Sample</b>																											
SO-1	C	Sediment Reference Sample (spiked at 170 ppb)	Aroclor-1254	134					1221 = <0 ppb Others = 20 - 40 ppb	40	40	40-120%	35-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	40-120%	
			Decachlorobiphenyl						131	Yes																	
			Tetrachloromethylenes						76	Yes																	
<b>Method QC</b>																											
<b>Blank</b>																											
			Aroclor-1016	-	A																						
			Aroclor-1211	-	A																						
			Aroclor-1232	-	A																						
			Aroclor-1242	-	A																						
			Aroclor-1248	-	A																						
			Aroclor-1254	-	A																						
			Aroclor-1260	-	A																						
			Decachlorobiphenyl						139	Yes																	
			Tetrachloromethylenes						0	No																	
<b>Orphaned Result</b>																											
			Aroclor-1016	0.4	U	0.4	Yes																				
			Aroclor-1211	0.4	U	0.4	Yes																				
			Aroclor-1232	0.8	U	0.8	Yes																				
			Aroclor-1242	0.4	U	0.4	Yes																				
			Aroclor-1248	0.4	U	0.4	Yes																				
			Aroclor-1254	0.4	U	0.4	Yes																				
			Aroclor-1260	0.4	U	0.4	Yes		82	Yes																	
			Decachlorobiphenyl						87	Yes																	
<b>Orphaned Result</b>																											
			Aroclor-1016										115	Yes													
			Aroclor-1260										116	Yes													
			Decachlorobiphenyl						48	Yes																	
			Tetrachloromethylenes						82	Yes																	
<b>Orphaned Result</b>																											
			Aroclor-1016	87	U	87	No																				
			Aroclor-1221	87	U	87	No																				
			Aroclor-1232	0.8	U	0.8	Yes																				
			Aroclor-1242	0.8	U	0.8	Yes																				
			Aroclor-1248	0.8	U	0.8	Yes																				
			Aroclor-1254	0.8	U	0.8	Yes																				
			Aroclor-1260	0.8	U	0.8	Yes																				
			Decachlorobiphenyl						133	Yes																	
			Tetrachloromethylenes						49	No																	
<b>Orphaned Result</b>																											
			Aroclor-1016	0.4	U	0.4	Yes																				
			Aroclor-1221	0.4	U	0.4	Yes																				
			Aroclor-1232	0.8	U	0.8	Yes																				
			Aroclor-1242	0.4	U	0.4	Yes																				
			Aroclor-1248	0.4	U	0.4	Yes																				
			Aroclor-1254	0.4	U	0.4	Yes																				
			Aroclor-1260	0.4	U	0.4	Yes																				
			Decachlorobiphenyl						75	Yes																	
			Tetrachloromethylenes						87	Yes																	

Blank cells = data is not available

\*Simpatico, LCS's and NSMSI's are only reported in % recovery in the Region 10 LIMS.

\*\*Preservation based on USEPA requirements for organic parameters (hold time is 1 year when frozen).

\*\*\*Sediment reference limits derived from information received from EPA Region 10 (Steve Reimer - SO-1 had a spiked value of 170 ug/kg when created, early historical results averaged 120 ug/kg. No control limits have been set in this laboratory.)

\*\*\*\*Assume 100% per John Wakeman

NO = value not calculated

Table G-2 - Lower Duwanish Triad Sampling Event  
PCB Data Review  
EPA Region 6

Field Sample ID	Sample Name	Lab Sample ID	Sample Description	Batch	Analyte	Results*	Qualifier	Sample RL	Project RL Mat?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK?	LCS % Rec	MS CLs Mat?	MS % Rec	MS/MSD RPD CLs Mat?	MSD % Rec	MSD RPD Mat?	Field Dup RPD Mat?	Preserved on Mat?**	Container OK?	Sample Date	Prep Date	Prep Hold Time Mat?	Analysis Date	Analysis Hold Time Mat?	Reference Sample#	% Concentration Increment DKT***							
SD-309-000	04344106	0411019-01	Primary	B4L0908	Aroclor-1016	15.4	U	15.4	Yes	1																										
					Aroclor-1211	25.7	U	31.7	Yes	1																										
					Aroclor-1232	15.4	U	15.4	Yes	1																										
					Aroclor-1242	15.4	U	15.4	Yes	1																										
					Aroclor-1248	76.6	U	15.4	Yes	1																										
					Aroclor-1254	34.0	U	15.4	Yes	1																										
					Aroclor-1278	15.4	U	15.4	Yes	1																										
					Decachlorobiphenyl	263				1	53.6	Yes																								
					Tetrachloro-methyl-ylene	280				1	72.9	Yes																								
SD-303-000	04344107	0411019-02	Primary	B4L0908	Aroclor-1016	11.8	U	11.8	Yes	1																										
					Aroclor-1211	25.5	U	27.5	Yes	1																										
					Aroclor-1232	11.8	U	11.8	Yes	1																										
					Aroclor-1242	11.8	U	11.8	Yes	1																										
					Aroclor-1248	11.8	U	11.8	Yes	1																										
					Aroclor-1254	89.7																														
					Aroclor-1278	11.8	U	11.8	Yes	1																										
					Decachlorobiphenyl	217				1	71.8	Yes																								
					Tetrachloro-methyl-ylene	214				1	72.6	Yes																								
SD-310-000	04344109	0411019-03	Primary	B4L0908	Aroclor-1016	15.5	U	15.5	Yes	1																										
					Aroclor-1211	33.2	U	33.2	Yes	1																										
					Aroclor-1232	15.6	U	15.6	Yes	1																										
					Aroclor-1242	16.5	U	15.6	Yes	1																										
					Aroclor-1248	16.5	U	15.6	Yes	1																										
					Aroclor-1254	106																														
					Aroclor-1278	15.5	U	15.5	Yes	1																										
					Decachlorobiphenyl	223				1	53.6	Yes																								
					Tetrachloro-methyl-ylene	227				1	78.6	Yes																								
SD-310-000	04344110	0411019-04	Primary	B4L0908	Aroclor-1016	15.5	U	15.5	Yes	1																										
					Aroclor-1211	31.2	U	31.2	Yes	1																										
					Aroclor-1232	15.6	U	15.6	Yes	1																										
					Aroclor-1242	16.5	U	15.6	Yes	1																										
					Aroclor-1248	16.5	U	15.6	Yes	1																										
					Aroclor-1254	106																														
					Aroclor-1278	15.5	U	15.5	Yes	1																										
					Decachlorobiphenyl	245				1	62.6	Yes																								
					Tetrachloro-methyl-ylene	317				1	81.3	Yes																								
SD-310-000	04344110	0411019-05	Primary	B4L0908	Aroclor-1016	15.5	U	15.5	Yes	1																										
					Aroclor-1211	31.2	U	31.2	Yes	1																										
					Aroclor-1232	15.6	U	15.6	Yes	1																										
					Aroclor-1242	15.6	U	15.6	Yes	1																										
					Aroclor-1248	15.5	U	15.5	Yes	1																										
					Aroclor-1254	254																														
					Aroclor-1278	20.4	U	15.6	Yes	1																										
					Decachlorobiphenyl	245				1	62.6	Yes																								
					Tetrachloro-methyl-ylene	317				1	81.3	Yes																								
SD-316-000	04344133	0411019-06	Primary	R4L0908	Aroclor-1016	15.5	U	15.5	Yes	1																										
					Aroclor-1211	33.6	U	33.5	Yes	1																										
					Aroclor-1232	15.6	U	15.6	Yes	1																										
					Aroclor-1242	16.5	U	15.6	Yes	1																										
					Aroclor-1248	15.5	U	15.5	Yes	1																										
					Aroclor-1254	201																														
					Aroclor-1278	149																														
					Decachlorobiphenyl	2170				70.1	Yes	2																								
					Tetrachloro-methyl-ylene	265				1	81.3	Yes																								
					Aroclor-1016	14	U	14	Yes	1																										
					Aroclor-1211	28	U	28	Yes	1																										
					Aroclor-1232	14	U	14	Yes	1																										
					Aroclor-1242	14	U	14	Yes	1																										
					Aroclor-1248	15	U	15	Yes	1																										
					Aroclor-1254	2170																														
					Decachlorobiphenyl	2170																														
					Tetrachloro-methyl-ylene	285																														
SD-316-000	04344135	0411019-07	Primary	B4L0908	Aroclor-1016	16.2	U	16.2	Yes	1																										
					Aroclor-1211	32.4	U	32.4	Yes	1																										
					Aroclor-1232	18.2	U	18.2	Yes	1																										
					Aroclor-1242	16.2	U	16.2	Yes	1																										
					Aroclor-1248	15	U	15	Yes	1																										
					Aroclor-1254																															

Table G-2 - Lower Dowlanski Triad Sampling Event  
PCB Data Review  
EPA Region 6

Field Sample ID	Sample Name	Lab Sample ID	Sample Description	Batch	Analyte	Results*	Qualifier	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Surrogate CLs OK?	Method Blank CLs OK?	LCS % Rec	LCS CLs Met?	MS % Rec	MS CLs Met?	MHD % Rec	MSD % Rec	MS/MSD RPD	Field Dup RPD Met?	Preservat on Met?	Container OK?	Sample Date	Prep Date	Prep Hold Time Met?	Analysis Date	Analysis Hold Time Met?	Reference Sample***	% Concentration Increment OK?****				
					Aroclu-1018 127.0 ± 40 not Other	11.3	U	11.3	Yes	1	80.7	Yes	DCS/P-65 < 20% of reported value or	50-130%	40-140%	40-140%	<50	<50%	<50%	<0°C					365 days (infract)	40 days (analysis)	77 days Average	120	95%					
					Tetrachloro-methyl-ethylene	327				1																	12/15/2004							
SD-315-000	04344129	0411019-05	Primary	B4L0908	Aroclu-1018 122.6 Aroclu-1232 11.3 Aroclu-1242 11.3 Aroclu-1248 11.3 Aroclu-1254 11.3 Aroclu-1260 11.3 Decachlorobiphenyl 202 Tetrachloro-methyl-ethylene	11.3 22.6 11.3 11.3 11.3 11.3 11.3 203	U	11.3 27.6 11.3 11.3 11.3 11.3 11.3 71.4 71.7	Yes Yes Yes Yes Yes Yes Yes Yes	1 1 1 1 1 1 1 1																12/15/2004 12/15/2004	Yes	Yes	8/19/2004	12/9/2004	Yes	Yes	Yes	Yes
SD-317-002	04344138	0411019-05	Primary	B4L0908	Aroclu-1018 28.4 Aroclu-1221 14.2 Aroclu-1232 14.2 Aroclu-1242 14.2 Aroclu-1248 60.0 Aroclu-1254 2830 Aroclu-1260 688 Decachlorobiphenyl 284 Tetrachloro-methyl-ethylene	14.3 28.4 14.2 14.2 14.2 60.0 14.2 2830 28.4 283 2.2	U	14.2 28.4 14.2 14.2 14.2 60.0 14.2 2830 28.4 283 79.5	Yes Yes Yes Yes Yes Yes Yes Yes Yes Yes Yes	1 1 1 1 1 20 1 1 1 1 1														12/15/2004 12/15/2004	Yes	Yes	8/19/2004	12/9/2004	Yes	Yes	Yes	Yes		
SD-317-003	04344139	0411019-10	Primary	B4L0908	Aroclu-1018 24.7 Aroclu-1221 12.3 Aroclu-1232 12.3 Aroclu-1242 12.3 Aroclu-1248 12.3 Aroclu-1254 279 Aroclu-1260 291 Decachlorobiphenyl 230 Tetrachloro-methyl-ethylene	12.3 24.7 12.3 12.3 12.3 12.3 12.3 279 12.3 291 74.4	U	12.3 24.7 12.3 12.3 12.3 12.3 12.3 279 12.3 291 74.4	Yes Yes Yes Yes Yes Yes Yes Yes Yes Yes Yes	1 1 1 1 1 1 1 1 1 1 1														12/15/2004 12/15/2004	Yes	Yes	8/19/2004	12/9/2004	Yes	Yes	Yes	Yes		
<b>Method QC</b>																																		
<b>Method Blank</b>																																		
					Aroclu-1018 Aroclu-1221 Aroclu-1232 Aroclu-1242 Aroclu-1248 Aroclu-1254 Aroclu-1260 Decachlorobiphenyl Tetrachloro-methyl-ethylene	9.9 19.8 9.9 9.9 9.9 9.9 9.9 176 201	U	19.3 19.8 19.8 19.8 19.8 19.8 19.8 70.7 68.7	Yes Yes Yes Yes Yes Yes Yes Yes Yes	1 1 1 1 1 1 1 1 1																12/9/2004	12/15/2004	Yes	Yes					
<b>Laboratory Control Sample</b>																																		
					LCS	B4L0908- BS1	Aroclu-1018 Aroclu-1250 Decachlorobiphenyl Tetrachloro-methyl-ethylene	610 590 165 193	U	19.7 19.7 19.8 19.5			61.9 59.8	Yes Yes													12/9/2004	12/15/2004	Yes	Yes				
<b>Matrix Spike</b>																																		
					MS	B4L0908- MS1	Aroclu-1018 Aroclu-1250 Decachlorobiphenyl Tetrachloro-methyl-ethylene	710 710 242 252	U	22.8 22.8 22.8 88.1			71.1 62.3	Yes Yes													12/9/2004	12/15/2004	Yes	Yes				
<b>Matrix Spike Duplicate</b>																																		
					MSD	B4L0908- MSD	Aroclu-1018 Aroclu-1250 Decachlorobiphenyl Tetrachloro-methyl-ethylene	902 766 242 260	U	22.9 22.9 22.8 87.9			78.1 67.2	Yes Yes	10.7 7.6	Yes Yes											12/9/2004	12/15/2004	Yes	Yes				
<b>Primary Samples</b>																																		
SD-310-000	04344149	0411019-12	Primary	B4L1001	Aroclu-1018 Aroclu-1221 Aroclu-1242 Aroclu-1248 Aroclu-1254 Aroclu-1260 Decachlorobiphenyl Tetrachloro-methyl-ethylene	16.9 33.8 16.9 16.9 33.8 41.1 254 355	U	16.9 33.8 16.9 16.9 33.8 16.9 254 355	Yes Yes Yes Yes Yes Yes Yes Yes	1 1 1 1 1 1 1 1																12/15/2004 12/15/2004 12/15/2004 12/15/2004 12/15/2004 12/15/2004 12/15/2004 12/15/2004	Yes	Yes	8/19/2004	12/10/2004	Yes	Yes	Yes	Yes

Table G-2 - Lower Duwamish Triad Sampling Event  
PCB Data Review  
EPA Revision 6

Table G-2 - Lower Duwamish Triad Sampling Event  
PCB Data Review  
EPA Review 6

Blank cells = data not available

\*Surrogates, LCS's and MS/MSD's are only reported in % recovery in the Region 1 CLIMs.

**\*\*Preservation based on PSEP requirements for organic parameters (hold time is 1 year when frozen).**

\*\*\*Sequin sediment reference limits derived from information received from EPA Region 10 (Steve Reimer of EPA stated, 'SQ-1 had a spiked value of 170 ug/kg when created, early historical results averaged 120 ug/kg. No control limits have been set in this laboratory. The

\*\*\*Not stated in laboratory

**TABLE G-3 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 8**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Results*	Qualifier (Lab Qualifier)	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Preservation Met?**	Container OK?	Analysis Date	Analysis Hold Time Met?	Reference Sample***	% Completeness OK?****
								20 ppb						5 months (analysis)	Spike = 170 ppb Average = 120 Acceptance = 40-140%
<b>Primary Samples</b>															
SD-307-0001	4344101	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	20.2			Yes								Yes
			Aroclor-1260	61.6			Yes								
			Decachlorobiphenyl					114							
SD-307-0002	4344102	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	30.1			Yes								Yes
			Aroclor-1260	132			Yes								
			Decachlorobiphenyl					107							
SD-307-0003	4344103	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	27.2			Yes								Yes
			Aroclor-1260	43.2			Yes								
			Decachlorobiphenyl					125							
SD-309-0001	4344105	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	184			Yes								Yes
			Aroclor-1260	69.1			Yes								
			Decachlorobiphenyl					118							
SD-310-0003	4344111	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	29.8			Yes								Yes
			Aroclor-1260	35.4			Yes								
			Decachlorobiphenyl					101							
SD-311-0001	4344113	Reg Sample	Aroclor-1248	259			Yes					Yes	12/30/2004		
			Aroclor-1254	110			Yes								Yes
			Aroclor-1260	217			Yes								
			Decachlorobiphenyl					94							
SD-311-0002	4344114	Reg Sample	Aroclor-1248	1280			Yes					Yes	12/30/2004		
			Aroclor-1254	4090			Yes								Yes
			Aroclor-1260	419			Yes								
			Decachlorobiphenyl					113							
SD-311-0003	4344115	Reg Sample	Aroclor-1248	703		20	Yes					Yes	12/30/2004		
			Aroclor-1254	3370			Yes								Yes
			Aroclor-1260	373			Yes								
			Decachlorobiphenyl					120							
SD-312-0001	4344117	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	1450			Yes								Yes
			Aroclor-1260	409			Yes								
			Decachlorobiphenyl					103							
SD-313-0001	4344121	Reg Sample	Aroclor-1248	20	U	20	Yes					Yes	12/30/2004		
			Aroclor-1254	64.2			Yes								Yes
			Aroclor-1260	20	U	20	Yes								
			Decachlorobiphenyl					117							

**TABLE G-3 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 8**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Results*	Qualifier (Lab Qualifier)	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Preservation Met?**	Container OK?	Analysis Date	Analysis Hold Time Met?	Reference Sample***	% Completeness OK?****
								20 ppb						6 months (analysis)	Spike = 170 ppb Average = 120 Acceptance > 40-140%
SD-314-0001	4344126	Reg Sample	Aroclor-1248	20	U	20	Yes		119	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	20	U	20	Yes								
			Aroclor-1260	20	U	20	Yes								
			Decachlorobiphenyl												
SD-316-0002	4344134	Reg Sample	Aroclor-1248	146			Yes		104	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	513			Yes								
			Aroclor-1260	120			Yes								
			Decachlorobiphenyl												
SD-319-0002	4344146	Reg Sample	Aroclor-1248	20		20	Yes		127	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	36.2			Yes								
			Aroclor-1260	96.7			Yes								
			Decachlorobiphenyl												
SD-321-0003	4344155	Reg Sample	Aroclor-1248	20	U	20	Yes		124	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	109			Yes								
			Aroclor-1260	183			Yes								
			Decachlorobiphenyl												
SD-322-0001	4344159	Reg Sample	Aroclor-1248	1420			Yes		128	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	482			Yes								
			Aroclor-1260	61.2			Yes								
			Decachlorobiphenyl												
SD-322-0005	4344183	Reg Sample	Aroclor-1248	63.2			Yes		114	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	147			Yes								
			Aroclor-1260	21.6			Yes								
			Decachlorobiphenyl												
SD-326-0001	4344170	Reg Sample	Aroclor-1248	179			Yes		105	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	693			Yes								
			Aroclor-1260	99.3			Yes								
			Decachlorobiphenyl												
SD-327-0002	4344171	Reg Sample	Aroclor-1248	111			Yes		91	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	349			Yes								
			Aroclor-1260	90.8			Yes								
			Decachlorobiphenyl												
<b>Primary Samples</b>															
SD-207-0000 (dup SD-435-0000, not analyzed)	4354001	Reg Sample	Aroclor-1248	20	U	20	Yes		62	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	104			Yes								
			Aroclor-1260	41.6			Yes								
			Decachlorobiphenyl												
SD-208-0000	4354002	Reg Sample	Aroclor-1248	34.7			Yes		90	Yes	Yes	12/30/2004			Yes
			Aroclor-1254	190			Yes								
			Aroclor-1260	116			Yes								
			Decachlorobiphenyl												

**TABLE G-3 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 8**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Results*	Qualifier (Lab Qualifier)	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Preservation Met?**	Container OK?	Analysis Date	Analysis Hold Time Met?	Reference Sample***	% Completeness OK?****
SD-209-0000	4354003	Reg Sample	Aroclor-1248	20	U	20	Yes							Spike = 170 ppb Average = 120 Acceptance = 40-140%	95%
			Aroclor-1254	57.5			Yes								
			Aroclor-1260	20.2			Yes								
			Decachlorobiphenyl					74							Yes
SD-212-0000	4354006	Reg Sample	Aroclor-1248	20	U	20	Yes								
			Aroclor-1254	48.9			Yes								
			Aroclor-1260	20	U	20	Yes								
			Decachlorobiphenyl					63							Yes
SD-217-0000	4354011	Reg Sample	Aroclor-1248	48.3			Yes								
			Aroclor-1254	201			Yes								
			Aroclor-1260	43.4			Yes								
			Decachlorobiphenyl					78							Yes
SD-331-0000	4354013	Reg Sample	Aroclor-1248	DNR											
			Aroclor-1254	DNR											
			Aroclor-1260	DNR											
			Decachlorobiphenyl					dnr							Yes
SD-332-0000	4354014	Reg Sample	Aroclor-1248	75.9			Yes								
			Aroclor-1254	227			Yes								
			Aroclor-1260	58.3			Yes								
			Decachlorobiphenyl					102							Yes
SD-337-0000	4354019	Reg Sample	Aroclor-1248	215			Yes								
			Aroclor-1254	719			Yes								
			Aroclor-1260	304			Yes								
			Decachlorobiphenyl					77							Yes
SD-341-0000	4354023	Duplicate of SD-333-0000	Aroclor-1248	20	U	20	Yes								
			Aroclor-1254	1170			Yes								
			Aroclor-1260	228			Yes								
			Decachlorobiphenyl					113							Yes
SD-342-0000	4354024	Duplicate of SD-338-0000	Aroclor-1248	113			Yes								
			Aroclor-1254	3290			Yes								
			Aroclor-1260	236			Yes								
			Decachlorobiphenyl					76							Yes
SD-345-0000	4354027	Reg Sample	Aroclor-1248	20		20	Yes								
			Aroclor-1254	124			Yes								
			Aroclor-1260	58.1			Yes								
			Decachlorobiphenyl					120							Yes
SD-431 (dup SD-333-0000, not analyzed)	4354031	Reg Sample	Aroclor-1248	20	U	20	Yes								
			Aroclor-1254	238			Yes								
			Aroclor-1260	65.4			Yes								
			Decachlorobiphenyl					108							Yes

**TABLE G-3 - LOWER DUWAMISH TRIAD SAMPLING EVENT**  
**PCB DATA REVIEW**  
**EPA REGION 8**

Field Sample ID	Lab Sample ID	Sample Description	Analyte	Results*	Qualifier (Lab Qualifier)	Sample RL	Project RL Met?	Dilution	Surrogate % Recovery	Preservation Met?**	Container OK?	Analysis Date	Analysis Hold Time Met?	Reference Sample***	% Completeness OK?****										
								20 ppb						6 months (analysis)	Spike = 170 ppb Average = 120 Acceptance = 40-140%										
<b>Reference Samples</b>																									
SQI		Sequin Reference (spiked at 7 ppb)	Aroclor-1248	20	U	20	Yes			Yes	Yes			No	Yes										
			Aroclor-1254	46.9			Yes																		
			Aroclor-1260	20	U	20	Yes																		
			Decachlorobiphenyl																						
SQI		Sequin Reference (spiked at 7 ppb)	Aroclor-1248	20	U	20	Yes			Yes	Yes			No	Yes										
			Aroclor-1254	57.6			Yes																		
			Aroclor-1260	20	U	20	Yes																		
			Decachlorobiphenyl																						
<b>Method QC</b>																									
<b>Matrix Spike</b>																									
		MS	Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
<b>Matrix Spike Duplicate</b>																									
		MSD	Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
<b>Matrix Spike</b>																									
		MS	Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
<b>Matrix Spike Duplicate</b>																									
		MSD	Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
<b>Laboratory Control Sample</b>																									
		LCS	Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
<b>Method Blanks</b>																									
		Blank	Aroclor-1248																						
			Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						
		Blank	Aroclor-1248																						
			Aroclor-1254																						
			Aroclor-1260																						
			Decachlorobiphenyl																						

Blank cells - data is not available

\*Surrogates, LCS's and MS/MSD's are only reported in % recovery in the Region 8 LIMS.

\*\*Preservation based on PSEP requirements for organic parameters (hold time is 1 year when frozen)

\*\*\*Sequin sediment reference limits derived from information received from EPA Region 10 (Steve Reimer - "SQ-1 had a spiked value of 170 ug/kg when created, early historical results averaged 120 ug/kg. No control limits have been set in this laboratory.")

\*\*\*\*Assume 100% per John Wakeman

NC = value not calculated

DNR - did not report; sample lost during preparation at the laboratory